



## 23 **Abstract**

24 The diversity of small shelly fossils (SSF) demonstrates that multicellular organisms underwent  
25 large-scale radiation at the beginning of the Cambrian, which is highlighted by the coexistence of  
26 various metazoans and the occurrence of their embryo fossils. However, little is known about early  
27 Cambrian eukaryotic multicellular algae, the primary producers that replaced oxygenic cyano-  
28 bacteria and played a crucial ecological role in matter cycling and energy dynamics in marine  
29 ecosystems. In this study, hundreds of microscopic three-dimensionally preserved multicellular  
30 agglomerate fossils were obtained from the early Cambrian Kuanchuanpu Formation (535 Ma) in  
31 southern Shaanxi, South China, which consisted of several tightly-packed multicellular clusters  
32 encapsulated within a thin organic membrane. Synchrotron tomography analysis further revealed  
33 that the cells of the whole agglomerate, although partitioned into different subunits by a gelatinous  
34 membrane, were distinctly differentiated into an outer conical cell layer and an inner spherical-cell  
35 layer, thus suggesting of a cortex-medulla-like differentiation. These characteristics resemble  
36 those of multicellular algae (e.g. *Wengania*, *Gremiphyca*, and *Thallophyca*) from the Ediacaran  
37 Weng'an biota (South China) in morphology, size, and internal cell structure. Furthermore, a po-  
38 tential asexual life cycle for these membranous algae was proposed based on their morphological  
39 and structural characteristics. Our findings support an evolutionary continuity of the multicellular  
40 algae from the Ediacaran to the early Cambrian Period.

## 41 **1. Introduction**

42 The chlorophyll-based photosynthesis of eukaryotic multicellular algae transformed the  
43 Proterozoic ocean-atmosphere system into its modern state (Butterfield, 2015; Tang et al., 2020).  
44 Particularly, the rise of algae, approximately 1,900 to 900 million years (Myr) ago  
45 (Sanchez-Baracaldo et al. 2017), in comparison with cyanobacteria made food webs more efficient  
46 in nutrient and energy transfer, driving the occurrence of larger and increasingly complex organ-  
47 isms (Brock et al., 2017). Recent studies on molecular biomarkers have indicated a rapid increase  
48 of multicellular algae (e.g. red algae and green algae) after the Cryogenian (Brocks et al., 2017).  
49 Moreover, abundant multicellular algae fossils dating from the Mesoproterozoic to the Ediacaran  
50 period were identified in the Indian plate (Kumar, 2001; Sharma et al., 2009; Sharma and Shukla,  
51 2009, Bengtson et al., 2017), North America (Walter et al., 1976, 1990), and the Yangtze plate  
52 (Zhu et al., 2016; Ding et al., 1992; Tang et al., 1997; Wang et al., 2007; Zhang, 1989; Xiao et al.,  
53 2002; Yuan et al., 2001, 2016; Zhao et al., 2004; Tang et al., 2007). However, most algae fossils  
54 are usually preserved as macroscopic compressed carbonaceous films without cellular structures,  
55 except for the fossils obtained from the Lower Vindhyan in central India (Bengtson et al., 2017)  
56 and the Ediacaran Doushantuo Formation in South China (Zhang, 1989; Zhang and Yuan, 1992).  
57 In particular, the Doushantuo phosphorites contain an exceptional record of multicellular algae  
58 featuring many anatomical and reproductive features seen in the modern marine flora such as  
59 conceptacle, spermatangia, and a cortex-medulla differentiation (Zhang, 1989; Zhang and Yu-  
60 an, 1992; Zhang et al., 1998; Xiao et al., 1999). These fossils provide an exceptional opportunity

61 to understand the evolution and diversification of algae in Neoproterozoic marine ecosystems  
62 during the advent of the Ediacaran metazoan radiation.

63 Cambrian fossil algae are quite abundant. They are preserved as macroscopic compressed  
64 carbonaceous films in Burgess shale-type Lagerstätten in south China, including the Chengjiang  
65 (Hou et al., 2017), Balang (Yang et al., 1999, 2001), Yanjiahe (Guo et al., 2011), and Qingjiang  
66 biotas (Fu et al., 2019). Additionally, they were also found in cherts (Yin, 1987; Shang et al., 2020)  
67 or phosphatized rocks (Shen, 1987) alongside various small shelly fossils. Although algae have  
68 been an essential component of the Cambrian marine ecosystem, they were largely neglected, and  
69 their role during Cambrian metazoan radiation was poorly investigated.

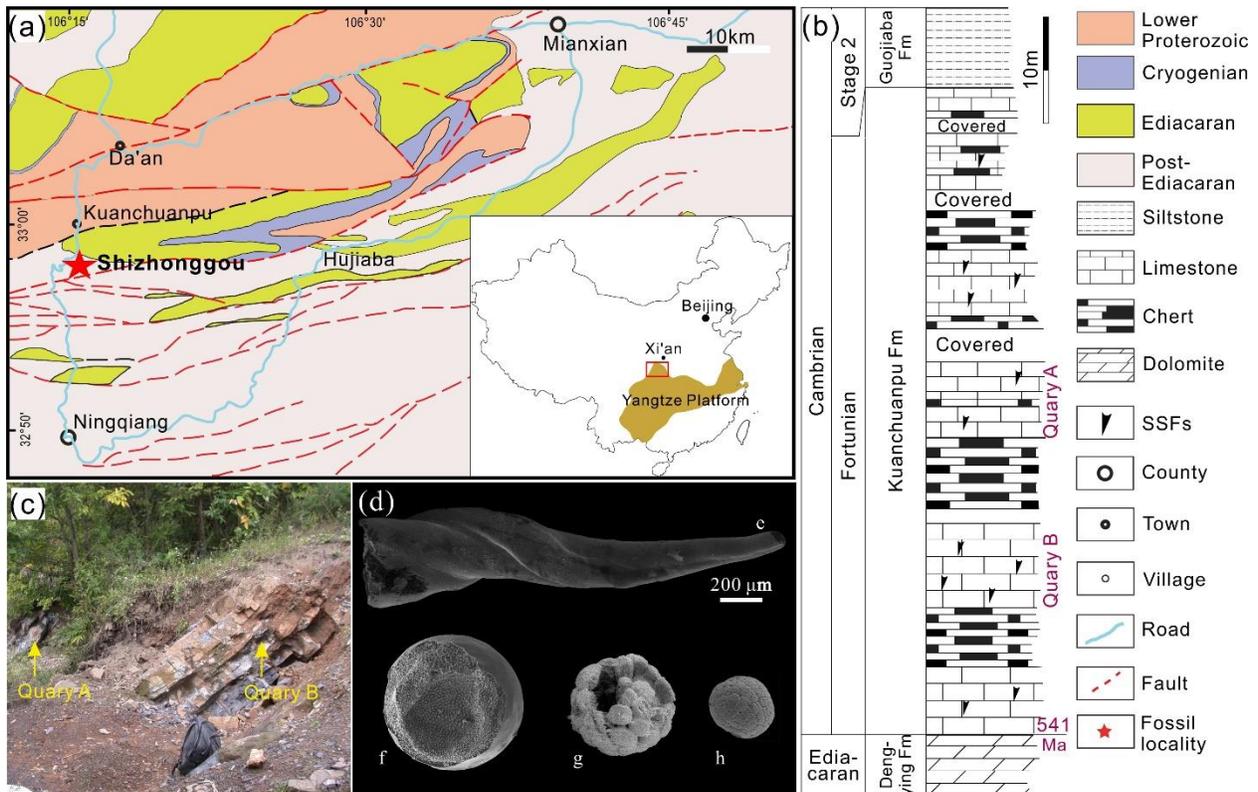
70 The early Cambrian Kuanchuanpu biota in Southern Shaanxi Province China (Fig. 1a–c)  
71 constitutes a complete ecosystem, which not only contains small shell fossils (Qian, 1999; Li,  
72 1992) and animal embryos (Yue and Bengtson, 1998; Bengtson and Yue, 1997; Zhang et al., 2015;  
73 Han et al., 2017)(Fig. 1d), but also up to ten species of microbial cyanobacteria (Ding, 1986; Yin,  
74 1987; Qian, 2007; Liu et al., 2014; Yang et al., 2017; Cui et al., 2020), and spherical coccoidal  
75 bacteria colonies (Steiner et al., 2004a). However, the algae were seldom documented, and hardly  
76 be distinguished from animal embryos in the cleavage stage. Zheng et al. (2017) reported multi-  
77 cellular spherical fossils with tetrad division and cellular structure, which were highly comparable  
78 to eukaryotic algae fossils from the Weng'an biota. Nevertheless, this study only characterized  
79 basic morphological features and did not demonstrate the internal structures for further biological  
80 classification.

81 Here we presented the morphology and internal microstructure of multicellular mem-  
82 brane-bearing algae from the early Cambrian Kuanchuanpu biota using a scanning electron mi-  
83 croscope (SEM) and synchrotron radiation X-ray tomographic microscopy (SRXTM). Our find-  
84 ings reveal a cell differentiation of algae thalli, and provide a theoretical basis for the systematic  
85 classification of these multicellular algae in comparison with those of the Weng'an biota.

## 86 **2. Stratigraphy**

87 All samples were collected from the Shizhonggou section of the Lower Cambrian  
88 Kuanchuanpu Formation in Ningqiang County, southern Shaanxi, China (Fig. 1a–c). This region  
89 comprises a series of shallow sea carbonate deposits of the Yangtze Platform, which is in turn  
90 primarily composed of dark grey or black phosphorus limestones interbedded with thin layered  
91 chert. In this section, the bottom of the Kuanchuanpu Formation is disconformably underlain by  
92 the gray-white dolomite at the top of the Ediacaran Dengying Formation (Bengtson and Yue,  
93 1992). There is also a disconformity between the Kuanchuanpu Formation and the overlying si-  
94 liciclastic Lower Cambrian Guojiaba Formation characterized by gray-dark silty shale. The  
95 Kuanchuanpu Formation is approximately 60–70 meters thick and rich in small shelly fossils and  
96 animal embryos (Fig. 1d), and is chronologically equivalent to the first small shelly fossil as-

97 semblage zone (i.e., the *Anabarites trisulcatus*–*Protohertzina anabarica* Zone) of the Fortunian  
 98 Stage and Terreneuvian Series (see Fig. 2 in Steiner et al., 2004a).



99

100 **Figure 1.** Geological map, stratigraphy, and typical fossils from the Shizhonggou section in  
 101 Ningqiang County, Shaanxi Province. (a), a geological map showing the Cambrian strata and the  
 102 fossil locality (modified from Han et al., 2016a). (b), stratigraphic column of the Shizhonggou  
 103 section in Ningqiang. (c), A photograph showing two quarries of the rock samples. (d), Mi-  
 104 cro-fossils from the Shizhonggou section: e, *Anabarites*, specimen No. ELISN001-K31-102; f,  
 105 an embryo of *Olivoooides*, No. ELISN138-5; g, an animal embryo in the gastrulation stage (From  
 106 Yasui et al., 2013); h, an embryo-like fossil, No. ELISN012-K31-098. Fm: formation.

### 107 3. Materials and methods

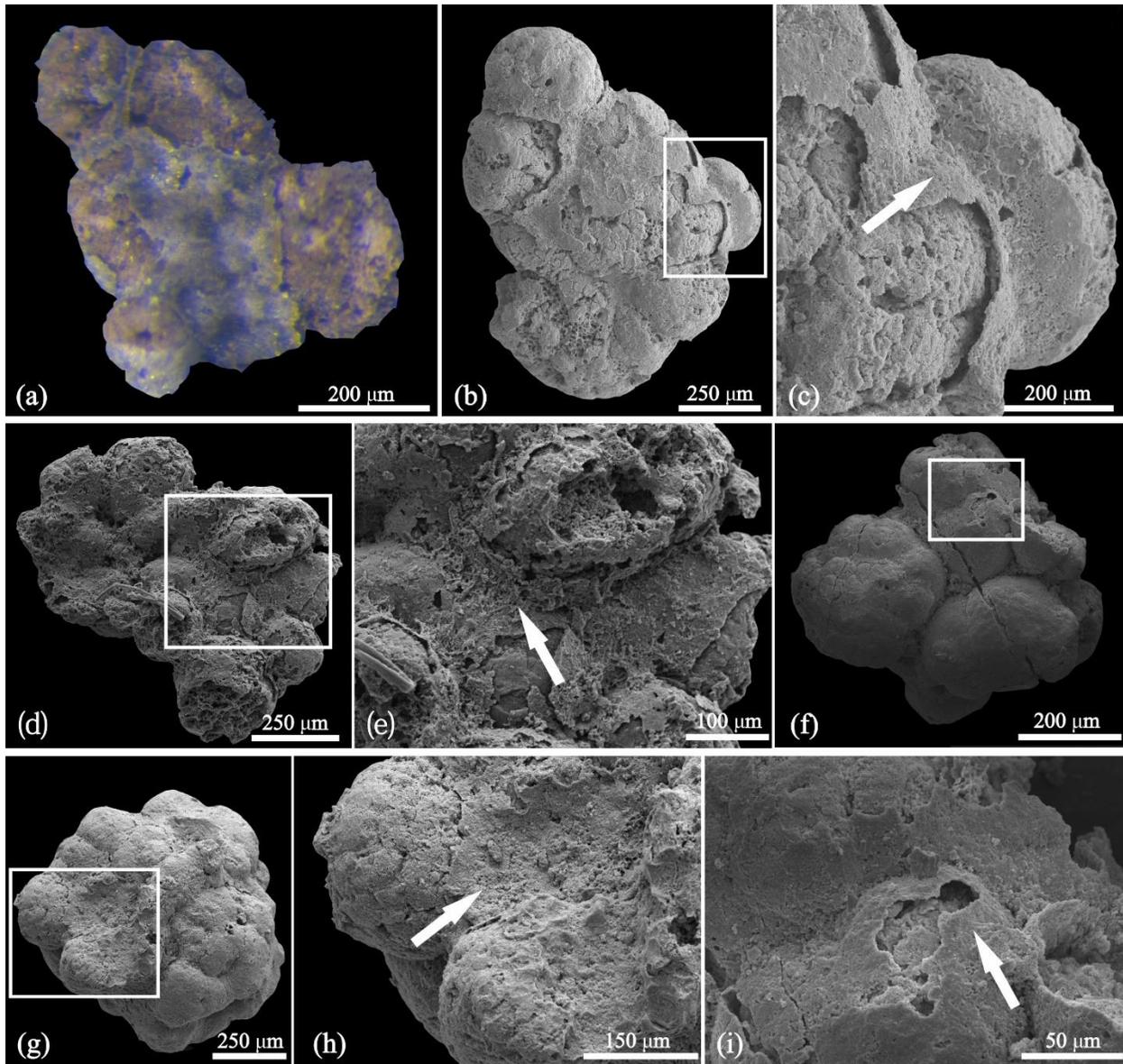
108 The rock samples were treated with 5%–10% acetic acid and manually selected under a  
 109 binocular stereoscopic microscope. The 117 selected specimens, coated with gold, were then ob-  
 110 served and measured using a SEM of FEI Quanta 400 FEG with a 20 kV voltage. Two samples  
 111 (ELISN004-51, ELISN159-297) were analyzed using SRXTM at SPring-8 (Hyogo, Japan). The  
 112 specimens were imaged through 360° rotation. 3D reconstructions of X-ray data and movies, were  
 113 conducted using the Dragonfly 4.1 software. All specimens are deposited at the SKLELE of  
 114 Northwest University, Xi'an, China.

115 **4. Results**

116 4.1. External morphology of the multicellular agglomerate fossils

117 The multicellular membrane-bearing agglomerate fossils are found to be amorphous in  
118 shape (Figs. 2, 3; Movie S1, S2), variable in size, and are constituted by several multicellular  
119 sphere-shaped or clump-shaped multicellular units that formed a secondary population (Figs. 2,3;  
120 Movie S1, S2). The length of the 117 specimens studied herein ranges from 500  $\mu\text{m}$  to 2900  $\mu\text{m}$   
121 (blue bar in Fig. 4) and are 1230  $\mu\text{m}$  on average, but most specimens range from 800 to 1400  $\mu\text{m}$ .  
122 Their widths range from 350 to 2000  $\mu\text{m}$  (red bar in Fig. 4) with a 500–1000  $\mu\text{m}$  dominance and an  
123 average of 820  $\mu\text{m}$  (Fig. 4). Some of the multicellular units in the agglomerates are regular  
124 spheroid (Fig. 3c), papillary (arrows in Fig. 3a, h), or rather irregular in shape (Figs. 3a, h and 5a).  
125 These multicellular units range from 30 to 500  $\mu\text{m}$  in diameter (350  $\mu\text{m}$  on average).

126 Light microscopy imaging shows that the microfossil agglomerate is encapsulated by a  
127 thin and smooth primrose yellow or white membrane layer, closely attached to the multicellular  
128 units (Fig. 2a). The organic membrane, approximately 1  $\mu\text{m}$  in thickness, is mostly visible in the  
129 recessed area (Fig. 2d–h). The membrane is inconspicuous between two multicellular clumps (Fig.  
130 6d, white arrow), but became much thicker at the triangular intersection between three or four  
131 multicellular clumps (Fig. 6i).



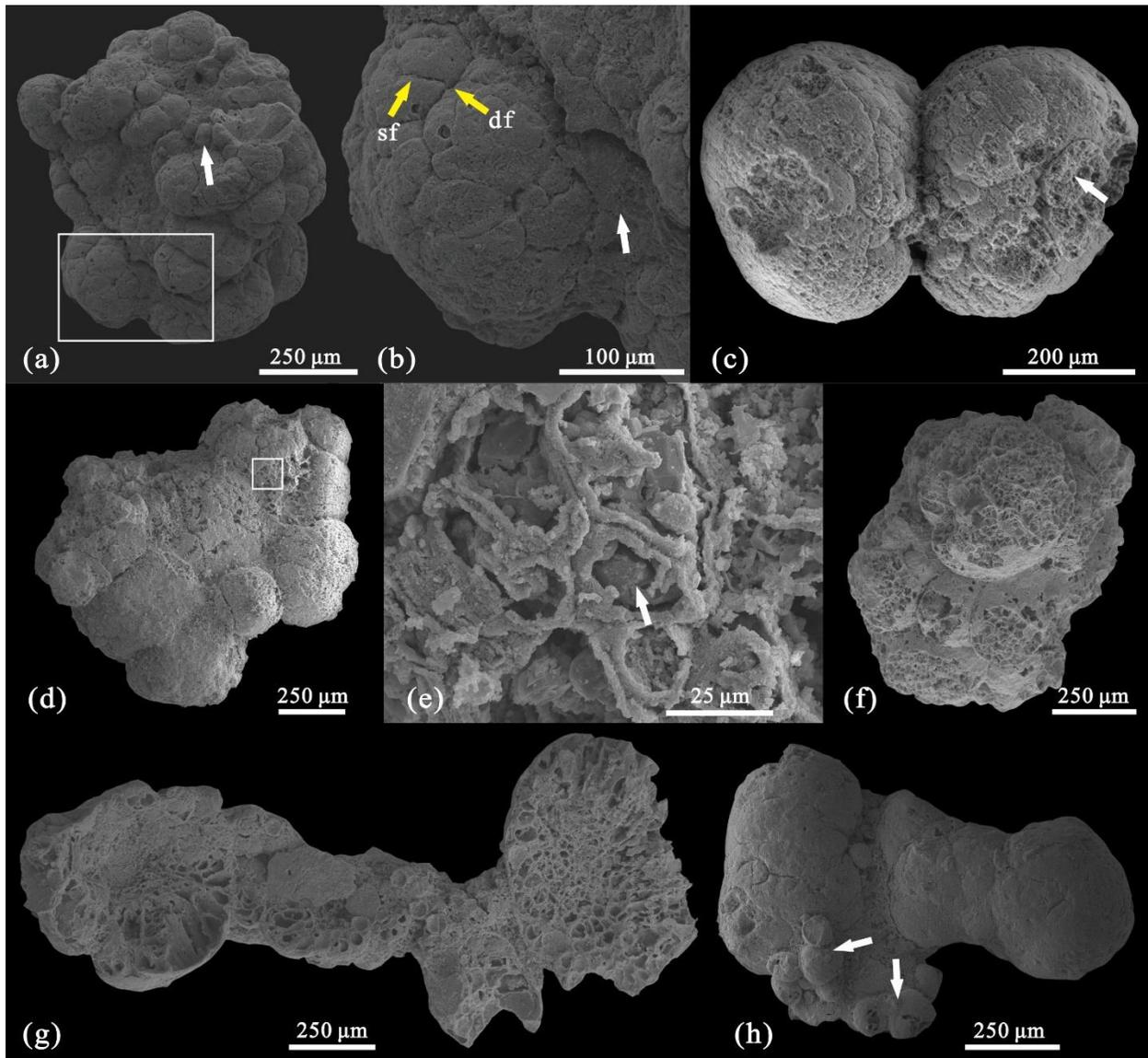
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133 **Figure 2.** External membrane of multicellular agglomerate fossils from the Cambrian  
 134 Kuanchuanpu Formation. (a), optical microscope imaging shows a smooth pale-yellowish external  
 135 membrane. (b–i), SEM imaging shows the decay of the external membrane. (c, e, h, and i),  
 136 close-up views of the areas marked by frames in (b, d, g, and f), respectively. (a–c): specimen No.  
 137 ELISN160; (d and e): specimen Nos. ELISN160 and ELI SN140-224; (f and i): specimen No.  
 138 ELISN160-203; (g and h): specimen No. ELISN159-297.

139 The multicellular units in the agglomerate fossils are tightly or loosely connected. Some  
 140 irregularly shaped units, bordered by evident cleavage depressions, are closely interconnected to  
 141 each other (Fig. 3c, f); whereas some larger units are loosely connected to the agglomerate and

142 even partitioned by the organic membrane, making them an independent unit within the agglom-  
143 erate (Fig. 3g).

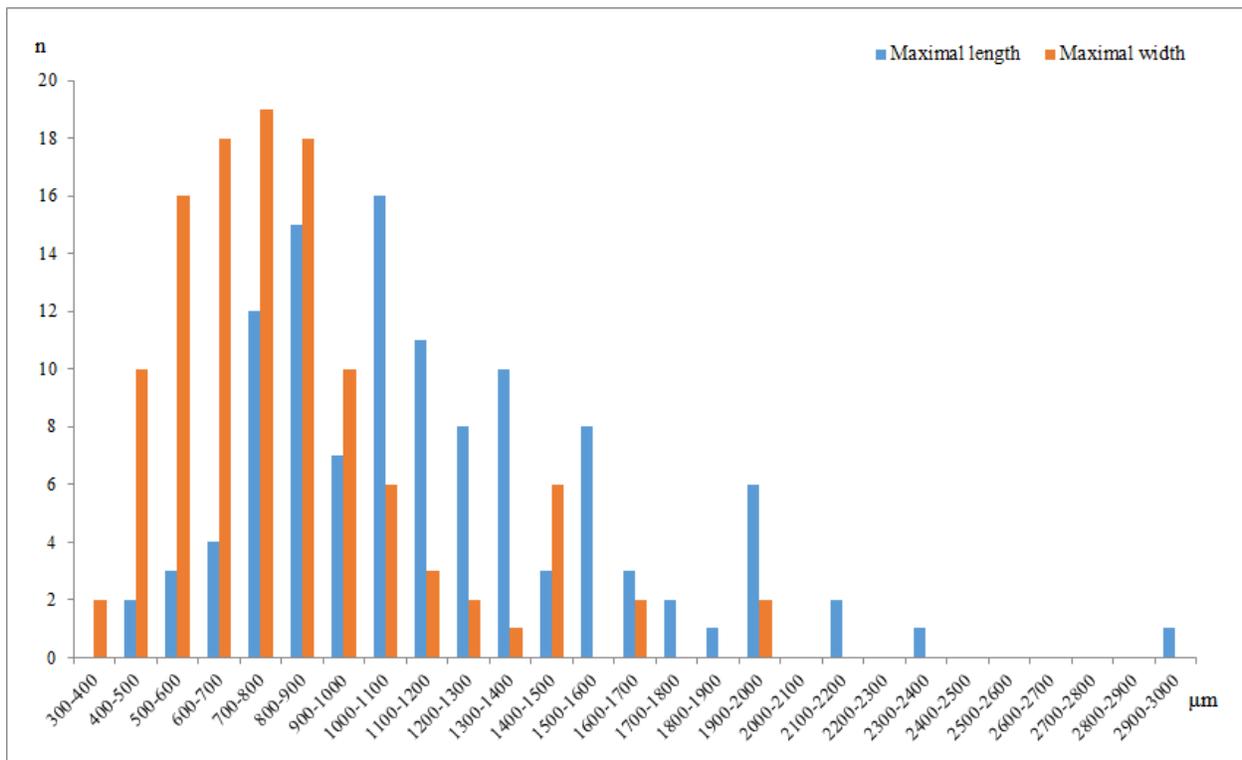
144 On the surface of some large multicellular units, the cleavage furrows are discernible in a  
145 hierarchy based on length and depth (Fig. 3b). The deeper and broader furrows (df) divided the  
146 unit into subunits, and the shallower, shorter ones (sf) in the subunit continued to divide the cell  
147 groups into clear-cut polygonal single cells approximately 30  $\mu\text{m}$  in width (Fig. 3e), which is  
148 reminiscent of cell division sequences. Some inconspicuous cleavage furrows could be recognized  
149 due to the presence of prominent hemispherical or clump-shaped subunits on the agglomerate  
150 surface (Fig. 3b, c).



151

152 **Figure 3.** External morphology of multicellular agglomerate fossils from the Cambrian  
153 Kuanchuanpu Formation. (a–h), Multicellular agglomerate fossils (ELISN160-413,

154 ELISN160-k31, ELISN164-294, ELISN167-244, ELISN160-153, ELISN160-310). (a) Multi-  
 155 cellular clump and mammillary clump (white arrow) size variations. (b) close-up view of the area  
 156 marked by the frame in A, which shows the fragmentary membrane between the units (white ar-  
 157 row) and different depth cleavage wrinkles (yellow arrows). (c), double spherical-like agglomerate  
 158 fossils and smaller multicellular units (white arrow) in the sphere. (d and e), multicellular ag-  
 159 gglomerate. (e), close-up view of (d) and polygonal cell (white arrow). (f), multicellular agglom-  
 160 erate without a preserved membrane, the latticed holes indicate single cells with broken walls. (g),  
 161 loosely connected agglomerate and the organic membrane (white arrow) that divides the multi-  
 162 cellular units. (h), agglomerate with nipple-like multicellular units (white arrows). df: deep furrow;  
 163 sf: shallow furrow.



164

165 **Figure 4.** Size distribution of agglomerate fossils from the early Cambrian Kuanchuanpu For-  
 166 mation in South China. Agglomerate fossil length (blue bars); agglomerate fossil width (red bars).

167 4.2. Internal morphology of multicellular agglomerate fossils

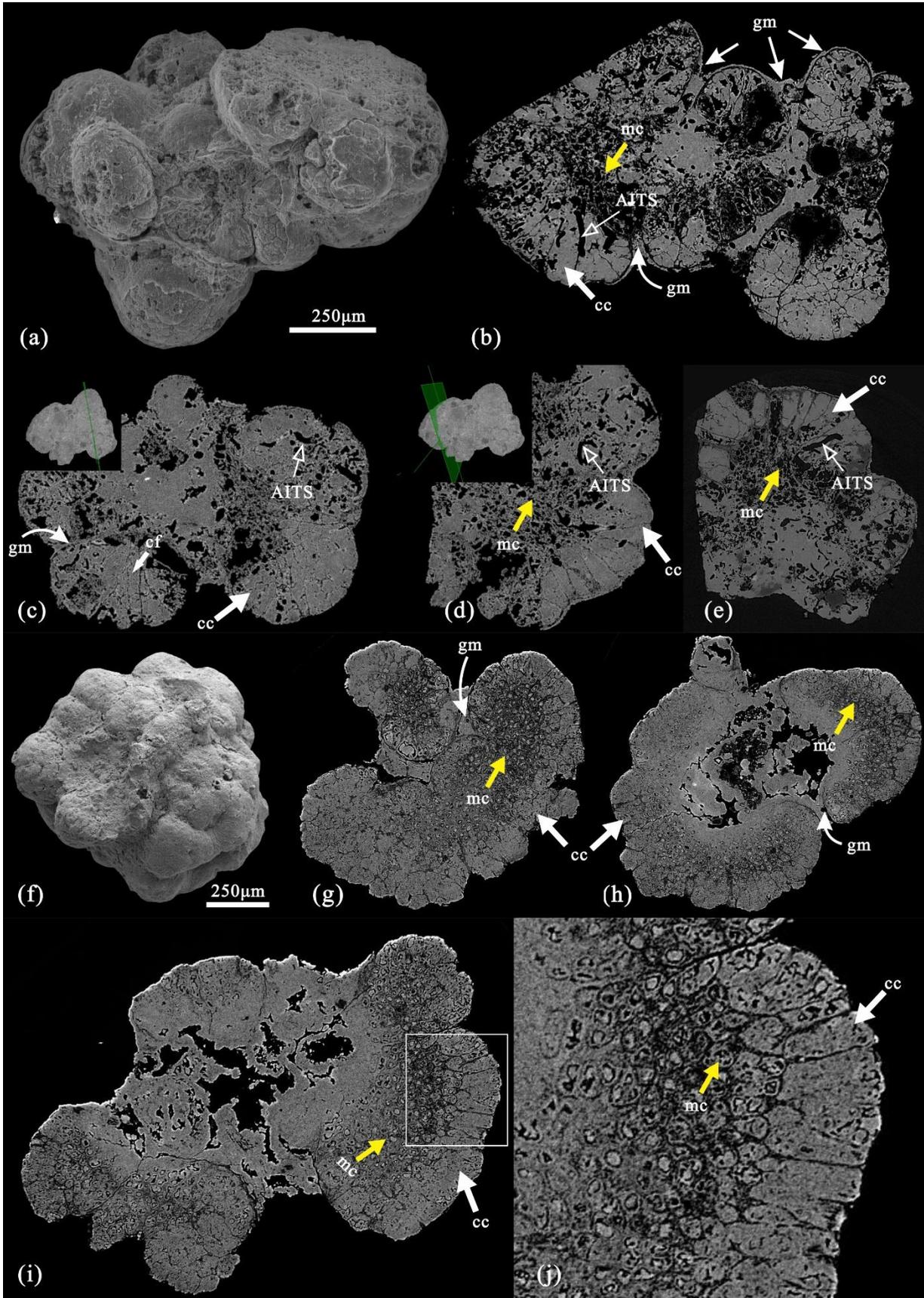
168 The SRXTM results are mostly consistent with the light microscopy or SEM observations  
 169 that the amorphous agglomerate is constituted by several different-sized multicellular  
 170 clump-shaped units with distinct boundaries (Fig. 5; Movie S1, S2). Moreover, the clump-shaped  
 171 units in the virtual cross section are further divisible as superficially triangular or trapezoid sec-  
 172 tions (Fig. 5b–e, g–i). On loosely connected multicellular units, the outer membrane may more or

173 less extend into the lumps, and the converging of the thin membrane in adjacent multicellular units  
174 may fuse into a substantially thicker non-cellular gelatinous membrane (gm) (Fig. 5b, c, g).

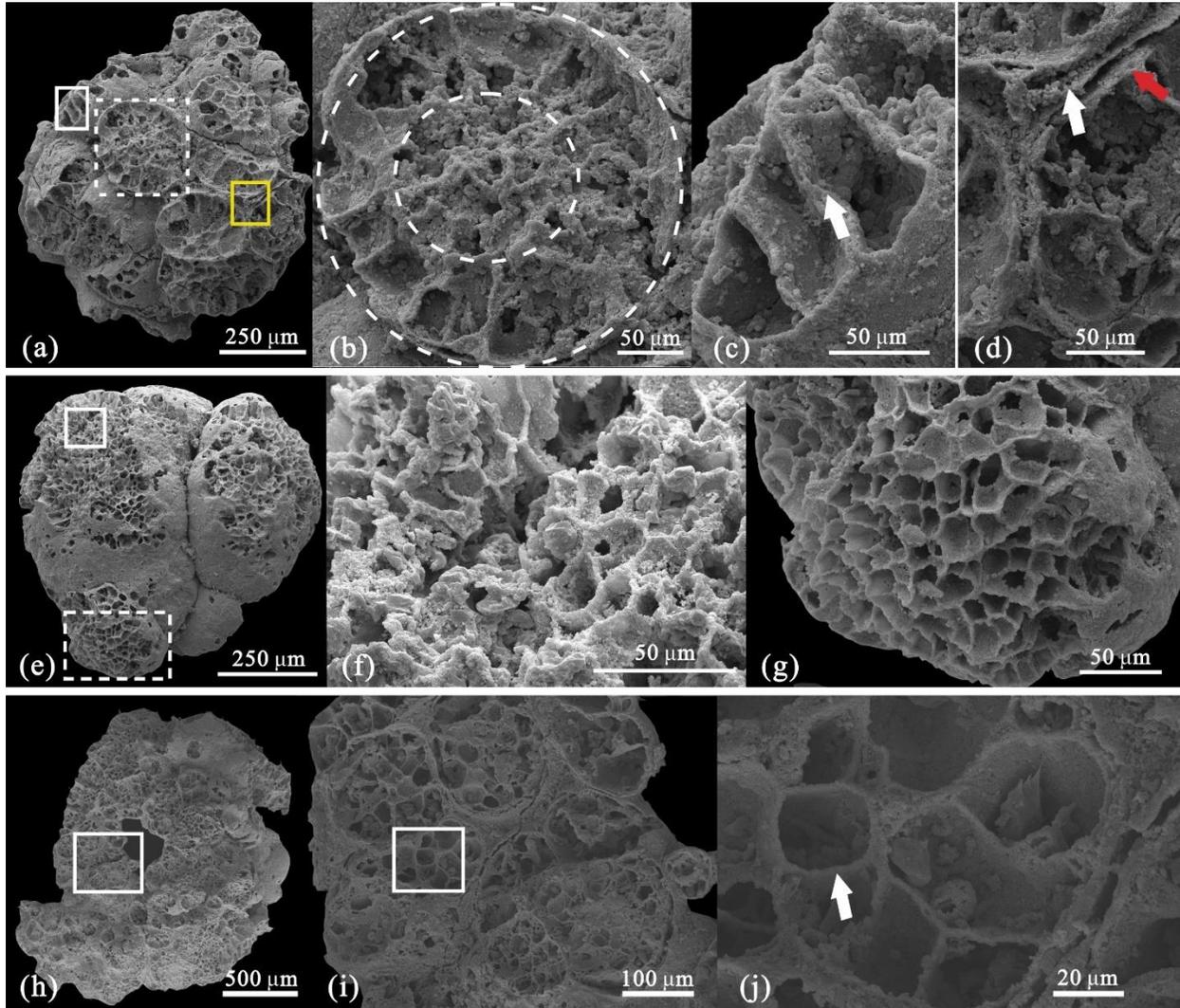
175 Interiorly, however, the cell clusters of the whole agglomerate, although partitioned into  
176 different subunits by the gelatinous membrane, are distinctly differentiated into an outer and an  
177 inner layer (Movie S1, S2). The outer cells are closely aligned at the peripheral region of the ag-  
178 glomerate with clear axial intercellular boundaries directed towards the agglomerate center (Fig.  
179 5i). These structures are equivalent to the above-mentioned triangular or trapezoid sections in the  
180 virtual cross section, which are approximately 15  $\mu\text{m}$  in width and 50  $\mu\text{m}$  in axial length, with a  
181 length-width ratio ranging from 1:3 to 1:4. The outer cells contribute approximately half of the  
182 agglomerate volume. The triangular or trapezoid sections are further subdivided into multiple  
183 cells. In contrast, the cells in the inner layer of the whole agglomerate or multicellular clump are  
184 indivisible into secondary groups. These cells are spherical or spherical-like/spindle-like, exhib-  
185 ited uniform shape, and are approximately 5  $\mu\text{m}$  in diameter, significantly smaller and uniform in  
186 size in comparison with the outer layer cells (Fig. 5e, j). Notably, the cell boundaries in some areas  
187 are obscured by the development of numerous hollow wormhole-shaped ambient inclusion trails  
188 (AITs) (Fig. 5b–e) that were produced in the early diagenesis (see Yang et al., 2017).

#### 189 4.3. Organic outer cell wall of the agglomerate fossils

190 The cellular structure in the multicellular clumps with broken surfaces is well exposed  
191 using SEM (Fig. 6). The outer layer within the clump interior is composed of a tightly-packed ring  
192 of cylindrical or square-column cells (indicated by two concentric circles in Fig. 6b) with lateral  
193 cell walls directed toward the clump center; whereas the surface view of the clump exhibits a  
194 honeycomb pattern like traditional Chinese "window lattice" (Fig. 6e–j) as the lacking of cyto-  
195 plasm and organelles. Each honeycomb structure represents a single cell, approximately 50  $\mu\text{m}$  in  
196 width. A shared cell wall, approximately 1.5  $\mu\text{m}$  in thickness, is visible between two adjacent  
197 cells. Whereas in some specimens, a narrow gap is observed between adjoining cell walls (white  
198 arrow in Fig. 6c–d). In contrast, the inner cell walls, as viewed via SEM and SRXTM, are much  
199 smaller; they are spherical in shape and are arranged in a reticular pattern (Fig. 6b, i; Movie S1,  
200 S2).



202 **Figure 5.** Interior anatomy of multicellular agglomerate fossils from the Cambrian Kuanchuanpu  
 203 Formation. (a–e), No. ELISN004-k30-51, multicellular agglomerate fossils. (a) SEM image; (b–  
 204 e), virtual sections of SRXTM of (a). (f–j), No. ELISN159-297, multicellular agglomerate fossils;  
 205 (f) SEM image; (g–i), virtual sections of SRXTM of (f); (j), close-up views of areas marked by  
 206 frames in (b3), which show cellular units. AITs: ambient inclusion trails; cc: cortical cells; cf: "cell  
 207 fountain"; gm: gelatinous membranes; mc: medullary cells.



208

209 **Figure 6.** The exposed cell walls of multicellular aggregate fossils from the Cambrian  
 210 Kuanchuanpu Formation. (a–d), No. ELISN166-86, multicellular aggregate fossils, the image  
 211 shows the structure of the multicellular clumps; (b), (c), (d) are magnifications of the white solid  
 212 frame, white dotted frame, and yellow frame in (a), respectively. (b) Cylindrical and square  
 213 columns of the outer cells that are radially arranged between two concentric circles; the spheri-  
 214 cal-like inner cells are arranged in a reticular pattern within the small concentric circles; (c) the gap  
 215 between adjacent cells (white arrow); (d) the organic matter membrane between multicellular

216 clumps (white arrow), and the cell wall (red arrow). (e–g), No. ELISN166-422, an aggregate fossil  
217 with cells arranged in a honeycomb pattern; (f), (g) are close-up view of the white solid frame,  
218 white dotted frame in (e), respectively. (h–j), morphology and cell arrangement of amorphous  
219 aggregate fossil specimen No. ELISN167-201; (i), (j) are magnifications of the white solid frame  
220 in (h) and (i), respectively.

## 221 **5. Discussion**

### 222 **5.1. Biological affinities**

#### 223 5.1.1 Cyanobacterial hypothesis

224 Spheriform colonial fossils are widespread in the Proterozoic to Cambrian rocks and are  
225 frequently interpreted as bacterial colonies (Butterfield, 2015; Sergeev and Schopf, 2010). For  
226 example, many sphaeromorphic colonial fossils (e.g. *Myxococcoides*) from the Kuanchuanpu  
227 Formation, including diverse microfossils of heterogeneous origin (e.g. cyanobacteria) and un-  
228 known hollow microbial sheaths (Cui et al., 2020), have been difficult to classify taxonomically.  
229 The aggregate fossils described herein superficially resemble cyanobacteria based on the external  
230 colloid sheath/gelatinous membrane and the aggregate preservation. Therefore, it comes first to  
231 determine whether the aggregate fossils described herein represented prokaryotes or eukaryotes by  
232 comparing their size, cellular differentiation, and individuality.

233 Size often takes a central role in discerning fossil eukaryotes and prokaryotes. The cells in  
234 modern spheroidal cyanobacteria are approximately 1–55  $\mu\text{m}$  and commonly about 4  $\mu\text{m}$  in di-  
235 ameter (Schopf and Oehler, 1976). The cells in cyanobacterial colonies are typically uniform in  
236 size, except for a few enlarged heterocysts (Golubic et al., 2000). Cyanobacteria fossils such as  
237 *Gloeodiniopsis* in the Kuanchuanpu Formation are approximately 9–20  $\mu\text{m}$  (Cui et al., 2020). In  
238 contrast, the cells in the aggregate fossil described herein range from 5 to 50  $\mu\text{m}$  in size, and such a  
239 wide variation was unknown in cyanobacterial colonies.

240 Cell differentiation is one of the most important criteria to distinguish multicellular or-  
241 ganisms from the colonial form of unicellular prokaryotes (Zhang, 1989). We can recognize  
242 morphologically two distinct types of cells in each multicellular unit of the aggregate fossils. The  
243 outer cells are mostly elongate cylindrical in shape, arranged in a radiate palisade along the margin  
244 of each multicellular unit; and they are much bigger and dramatically different from the fusiform  
245 inner cells. Therefore, the agglomerate fossils examined herein undoubtedly represent a multi-  
246 cellular organism.

#### 247 5.1.2 Metazoan embryo hypothesis

248 As discussed above, the aggregate fossils described herein likely represent higher eukar-  
249 yotes, particularly animal embryos or multicellular algae. In the Kuanchuanpu biota, metazoan  
250 embryos fossils from blastocyst to larvae were reported and generally accepted for their synap-  
251 omorphies (Yue and Bengtson, 1998; Bengtson and Yue, 1997). These agglomerate fossils exhibit

252 a distinct "cleavage-like" structure, which is superficially similar to metazoan embryo fossils in the  
253 early cleavage stage. Although determining the biological identities of aggregate fossils remains a  
254 considerable challenge, the aggregate fossils studied herein can still be identified based on their  
255 size, shape, cell cleavage, external membrane, and cell wall.

256 (1) Size: the aggregate fossils studied herein are inconsistent with co-occurred embryo  
257 fossils in size. Most animal embryo fossils from the Kuanchuanpu Formation are approximately  
258 590–910  $\mu\text{m}$  in diameter, and only a few embryos range from 1250 to 1690  $\mu\text{m}$  (Steiner et al.,  
259 2004b, 2014). The aggregate fossils range from 500  $\mu\text{m}$  to 2900  $\mu\text{m}$  in maximal length and 350–  
260 2000  $\mu\text{m}$  in maximal width, much larger than the co-occurred animal embryos.

261 (2) Shape: the Cambrian animal embryos with a cleavage-like structure were known to  
262 possess an egg envelope even in thickness (see Han et al., 2013, 2016b). Therefore, the soft tissue  
263 is restricted to a spherical egg envelope. In contrast, although the aggregate fossils are encapsu-  
264 lated by a thin organic membrane (Fig. 2), its amorphous shape and growth in random directions  
265 (Fig. 3) are inconsistent with animals or animal embryos with defined configurations.

266 (3) Cell cleavage: the cell cleavage pattern is among the most important factors to deter-  
267 mine the biological affinities of microfossils. Metazoan embryos generally exhibit a pattern of  
268 palintomic cell cleavage without cytoplasmic growth. Through successive cell proliferation, a  
269 single cell may build a stereoblastula-like structure with hundreds of daughter cells within. These  
270 daughter cells are spherical with smooth edges and loosely arranged, making it difficult to trace the  
271 sequence of cell divisions as lacking identifiable major division furrows (Stern, 2004). By con-  
272 trast, the cells of the aggregate fossils are tightly packed with each other and are delimited by  
273 polygonal boundaries or furrows (Fig. 6f); moreover, the sequence of multistage cell divisions is  
274 quite distinct from that of animals.

275 (4) External membrane: the entire metazoan embryo during the cell division stages is  
276 generally covered by an independent egg envelope with a constant thickness (Fig. 1f) and it is  
277 more resistant than soft-tissue of the enclosed embryos. By contrast, the external organic mem-  
278 brane of the agglomerate fossils can extend into the gap between multicellular units (Fig. 5b, g, h),  
279 unlike the egg envelope of an animal embryo. It is worthy of noting that the organic membrane is  
280 unlikely a diagenetic coating constituted by one or several layers of acicular apatite crystals (see  
281 Fig. 4.1 in Steiner et al., 2014) with regard to the exclusive occurrence of the homogenous com-  
282 position in current aggregate fossils and its absence in other co-occurred small shelly fossils.  
283 Therefore, the examined structures most likely represent the organic structure originally secreted  
284 by the organisms.

285 (5) The cell wall is the most fundamental difference between agglomerate fossils and  
286 animals. Animal cells are devoid of cell walls and are generally only equipped with a protein cell  
287 membrane, 7–8 nm in thickness. In the agglomerate fossils, some outer walls appear as square  
288 columns (Fig. 6j), indicating that the outer organic wall had a certain thickness and stiffness

289 comparable to that of a cell wall. If this is true, the interpretation of the agglomerate fossils as  
290 animal embryos appears unlikely.

### 291 5.1.3 Multicellular algae hypothesis

292 Based on the external membranes, cell differentiation, and cell wall structure, the fossil  
293 aggregates are most likely multicellular algae.

294 First, the organic membrane of the agglomerate fossils is very common in eukaryotic algae  
295 (Barsanti and Gualtieri, 2006), whose cells are encapsulated by a colloid coating that is very sim-  
296 ilar to the organic membrane in the aggregate fossils. Second, the cells of the multicellular units  
297 differentiate into outer and inner layers in the agglomerates (Fig. 5). In the outer layer, the  
298 close-packed pyramidal cells diverge radially from the sphere center to the surface; the vertical  
299 cell rows expand outward to form a fountain-like anatomical array (Fig. 5c), which is reminiscent  
300 of a "cell fountain", a distinctive hallmark of pseudo-parenchymatous growth in extant red and  
301 brown algae (see Bold and Wynne, 1985). The cells in the inner layer are spherical, spheroidal, or  
302 spindle-shaped, and are significantly smaller than those in the outer layer and irregularly arranged  
303 (Fig. 5g–j). This double-layered structure is similar to the cortex and medulla of multicellular  
304 algae (Zhang, 1989). Finally, the inferred cell wall, pyramidal shape, distinct stiffness, and decay  
305 resistance of the fossil aggregates favor the interpretation of the aggregate fossils as the multi-  
306 cellular algae. However, a more detailed comparison with modern algae cannot be conducted due  
307 to the lack of biochemistry and subcellular fraction evidence. Nonetheless, a comparison between  
308 our samples and exceptionally preserved Precambrian algae (e.g. the Weng'an biota in the  
309 Doushantuo phosphorites, China) may provide informative clues.

310 The Weng'an biota is remarkably rich in cellularly preserved multicellular algae including  
311 *Wengania globosa*, *Wengania exquisite*, *Thallophyca corrugate* (Zhang, 1989), *Gremiphyca*  
312 *corymbiata* (Zhang et al., 1998), *Thallophycoides phloeatus* (Zhang and Yuan, 1992). Based on the  
313 characteristics of well-preserved pseudoparenchyma, apical meristem, parenchyma, and complex  
314 thallus with cortex-medulla differentiation, as well as the reproductive tissues (carpospores, car-  
315 posporangia, and conceptacles), *Wengania*, *Gremiphyca*, *Thallophycoides* and *Thallophyca* in  
316 Weng'an biota were classified as Florideophyceae (Rhodophyta) (Zhang et al., 1989; Zhang et al.,  
317 1998; Xiao, 2004; Yuan et al., 2002).

318 There are striking similarities between the above-mentioned Weng'an algae and aggregate  
319 fossils in the Kuanchuanpu biota. (1) Thallus shape similarities. Although the shape of the thallus  
320 in Weng'an biota is ambiguous based on random thin section (i.e. two-dimensional) observations,  
321 the consistently circular outline of thalli observed in the thin sections suggests that *Wengania* tend  
322 to be nodular and lumpy or that other more complex thalli may co-exist in the same assemblage  
323 (Yuan et al., 2002; Xiao et al., 2004). Moreover, the aggregate fossils in the Kuanchuanpu biota  
324 are constituted of sphere-shaped or lump-shaped units. Particularly, the double-sphere shape of  
325 *Wengania globosa* obtained in the Doushantuo Formation in Shaanxi Province (Fig. 9 in Xiao et

326 al., 2004) is congruent with the specimen in Fig. 2c. Additionally, the spatial configuration of  
327 multicellular units in *Gremiphyca corymbiata* (Figs. 12–14 in Xiao, 2004), resembles that of the  
328 agglomerate fossils of the Kuanchuanpu biota (Figs. 3, 6). (2) Size similarities. *W. globosa* thalli in  
329 thin sections range from 70 to 750  $\mu\text{m}$  in diameter (Xiao et al., 2002). In the Kuanchuanpu biota,  
330 although the size of the lumps in the agglomerate fossils range from 30 to 500  $\mu\text{m}$  (i.e. slightly  
331 smaller than the dimensions associated with *W. globosa*), they are still very close to the sizes  
332 expected in *W. globosa*. (3) Cell differentiation. The samples had a similar "cell fountain"-like  
333 arrangement and double-layer differentiation, forming the cortex and medulla of the thallus (Figs.  
334 4, 5 in Xiao et al., 2004).

335 Notably, there are some distinctions between the agglomerate fossils in the Kuanchuanpu  
336 biota and the multicellular algae in the Weng'an biota. (1) The organic membrane is absent in  
337 Weng'an algae, although a potential of a taphonomical artifact cannot be excluded. (2) The apical  
338 meristem and reproductive tissues (carospores, carposporangia, and conceptacles) are absent in  
339 the agglomerate fossils studied herein.

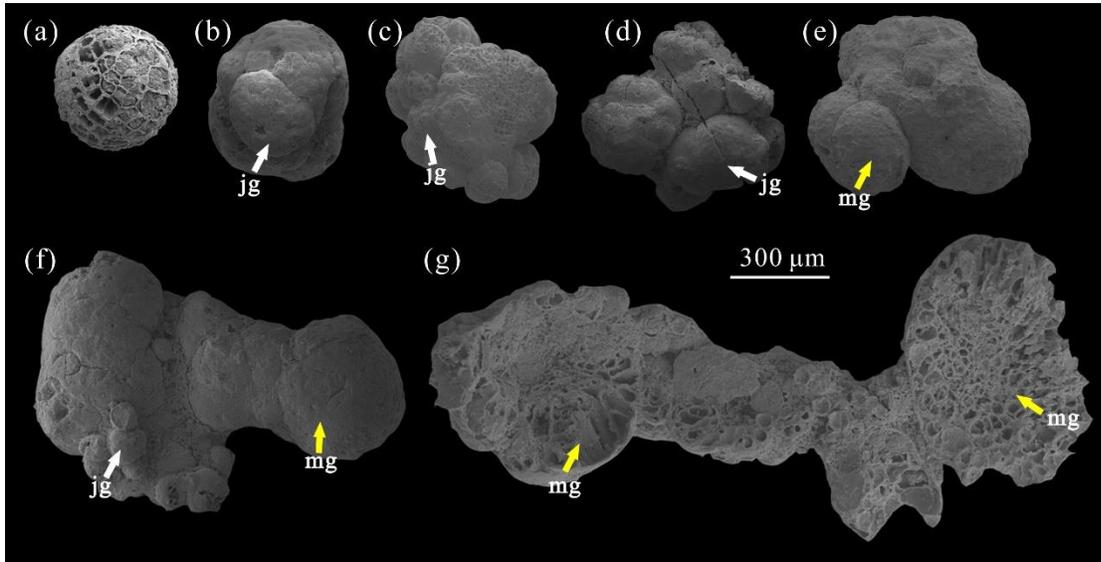
340 In summary, although the agglomerate fossils in the Kuanchuanpu biota exhibit some  
341 differences from those in the Weng'an biota, their external morphology, spatial configuration, size,  
342 pseudoparenchyma, 'cell fountains', and cortex-medulla differentiation are all suggestive of a close  
343 affinity with the Weng'an algae. Therefore, the discovery of agglomerate fossils in the  
344 Kuanchuanpu and Weng'an biota tend to support continuous evolution of multicellular algae from  
345 the Ediacaran to the early Cambrian, consistent with the evidence provided by the skeletal fossils  
346 in Ediacaran-Cambrian boundary (Cai et al., 2019; Laflamme et al., 2013; Yang et al., 2016; Zhu  
347 et al., 2017).

## 348 **5.2. A hypothesized life cycle of asexual reproduction**

349 Although the full-grown, mature forms remain obscure, the large abundance of the ag-  
350 gregate fossils from the Kuanchuanpu Formation allows us to reconstruct their asexual life cycle  
351 based on three lines of evidence. First, the sphere-shaped or lump-shaped unit of the aggregate  
352 fossils, which is well delimited from adjacent units, is most likely proliferated from a single cell.  
353 Second, the proliferation in random direction of these the aggregate fossils and their units can  
354 produce a broad scope of morphological variations. Third, the aggregate fossils most likely be-  
355 long to the same taxon based on amorphous morphology, continuous range of size (Fig. 4), cell  
356 division, and the cell differentiation pattern. These fossils can be grouped into three distinct stages  
357 of life cycle based on their increasing size and number of multicellular units (Figs. 7, 8).

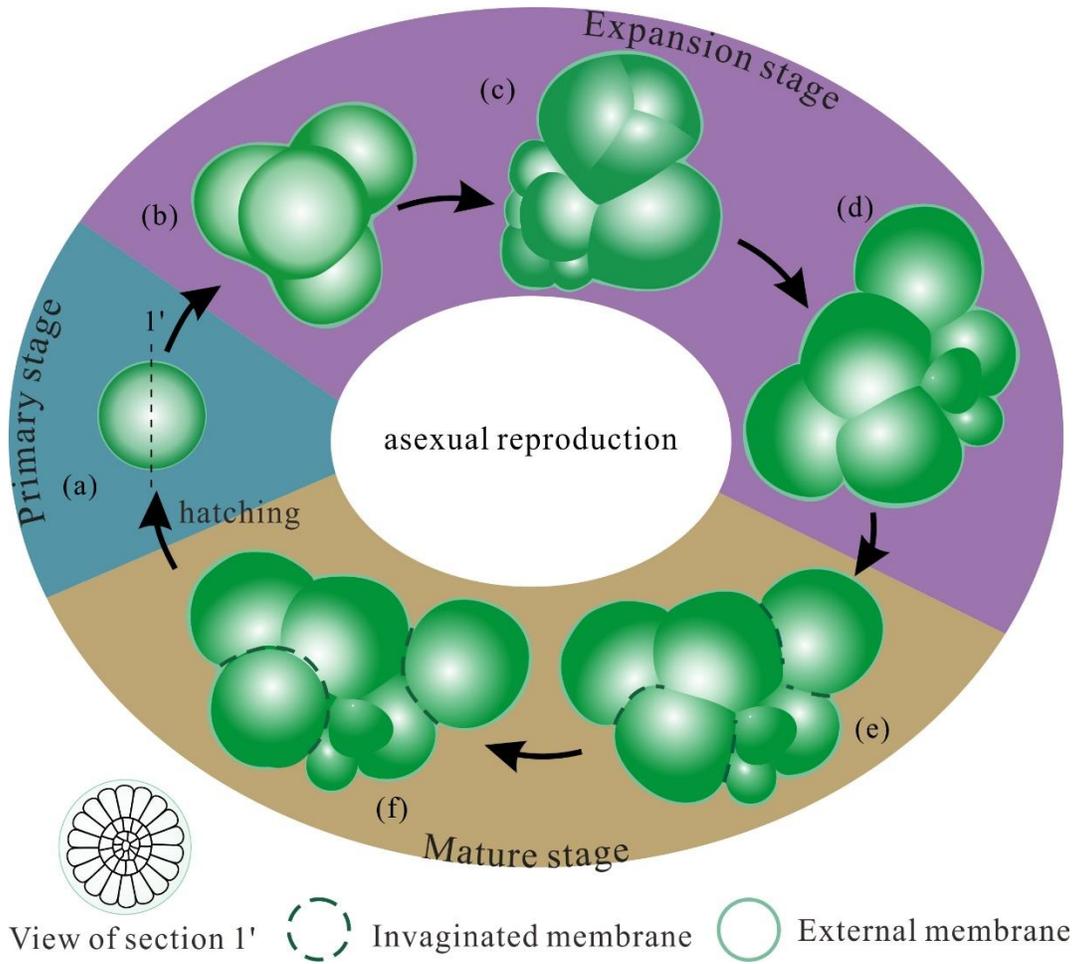
358 In the primary stage, the aggregate membrane-bearing algae are spherical or sub-spherical  
359 (Figs. 7a, 8a) with uniform size of external cells, indicating a constant speed of proliferation. In  
360 the expansion stage, a portion of cells in the outer layer of a single sphere may develop succes-  
361 sively inside a common membrane by successive proliferation, thereby forming a non-spherical  
362 cauliflower-like elevated bump structure; and lastly, growing into a prominent spherical multi-

363 cellular unit interpretable as a gemmule. The random formation of new bumps, probably mediated  
364 by allometric growth, leads to an amorphous agglomerate configuration composed of multiple  
365 multicellular units or gemmules (Figs. 7b–d, 8b–d). In this stage, the organic membrane is re-  
366 stricted to the external surface of the agglomerate. In the relatively mature stage, the outer mem-  
367 brane extends into space among multicellular units. Subsequently, the unit becomes enveloped by  
368 its own organic membrane (Figs. 7e–g, 8e–f) that finally detach from the agglomerate and form an  
369 independent individual (Figs. 7a, 8a).



370

371 **Figure 7.** Aggregate fossils in the putative life cycle. (a), an algae fossil in the initial growth stage;  
 372 (b), an algae fossil in the reproductive stage, with a juvenile gemmule structure (yellow arrow); (c–  
 373 d), algae fossils in the subsequent allometric growth stage with larger gemmules; (e–g), algae  
 374 fossils in the relative mature stage exhibiting a mature gemmule (white arrow), which will readily  
 375 separate from the matrix. All these images share the same scale. jg: juvenile gemmule; mg: mature  
 376 gemmule.



378 **Figure 8.** Schematic drawing of the asexual life cycle of the multicellular aggregate algae.

### 379 6. Conclusion

380 Based on the basic morphology, external membranes, spatial configuration, cell differen-  
 381 tiation, and particularly the presence of a cell wall, it is more reasonable to interpret the fossil  
 382 aggregates in the Kuanchuanpu biota as multicellular algae, rather than cyanobacteria or metazoan  
 383 embryos. The characteristics of the size, morphology, and cortex-medulla differentiation pattern  
 384 of these agglomerate fossils favor an affinity of Rhodophyta as multicellular algae from the  
 385 Weng'an biota. We also reconstructed a putative asexual life cycle of these aggregate fossils. The

386 discovery of agglomerate algae in the Kuanchuanpu biota may provide critical insights for a better  
387 understanding of the evolution and fate of the multicellular algae during the Ediacaran–Cambrian  
388 transition period. It may provide new evidence on the continuous evolutionary process of Edia-  
389 caran and Cambrian creatures.

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