

Relationship between genetic and phenotypic variations in natural populations of perennial and biennial sagebrush

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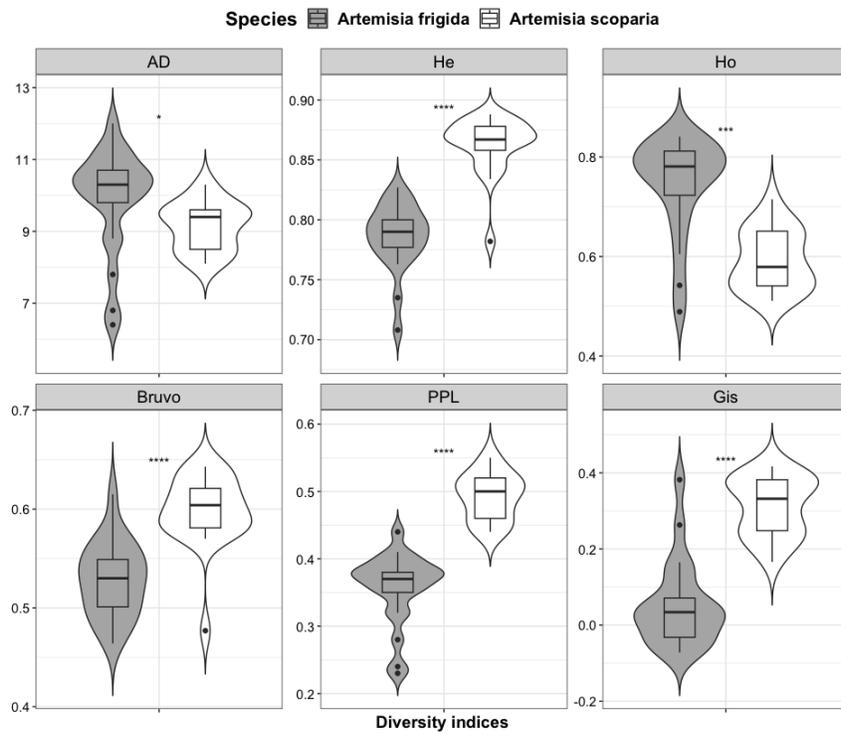
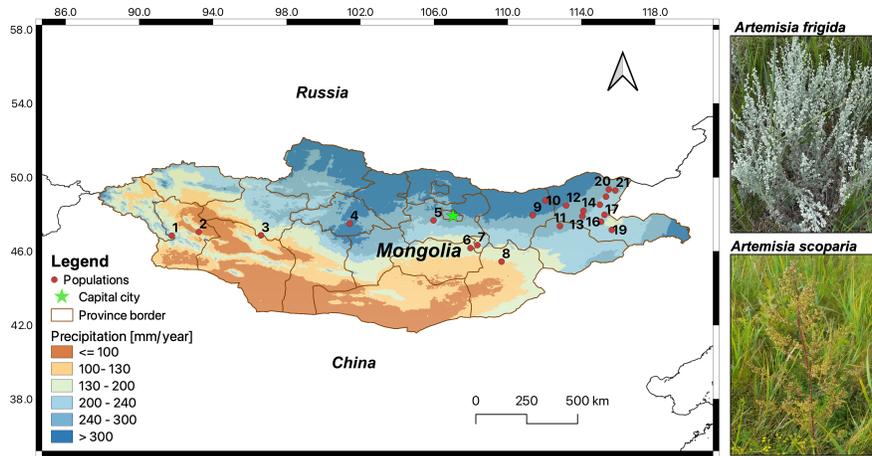
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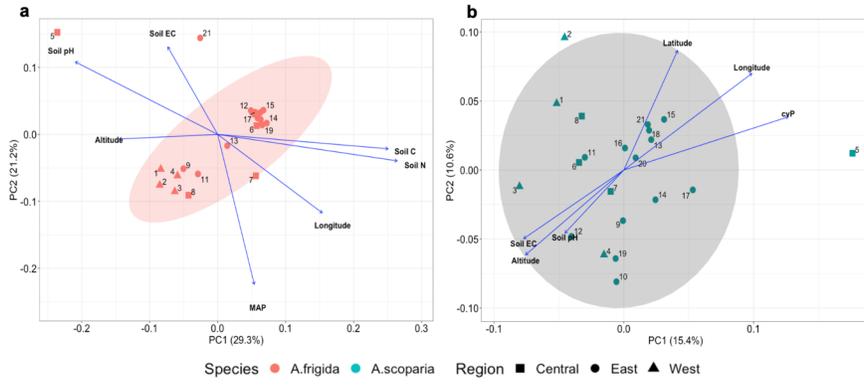
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Abstract

Plant responses to environmental heterogeneity depend on life-history traits, which could relate to phenotypical and genetic characteristics. To elucidate this relationship, we examined the variation in population genetics and functional traits of short- and a long-lived *Artemisia* species that are co-occurring in the steppes of Mongolia. Mongolian steppes represent stressful, waterlimited habitats demanding phenotypic modifications in the short term and/or genetic adaptation in the long term. However, detailed knowledge is missing about both plant phenotypic and genetic differentiation and their inter-relationships in temperate grasslands. Here, we investigated 21 populations of the widely distributed subshrub *A. frigida* and the herbaceous biennial *A. scoparia*. Genetic variation was assessed with newly developed Simple Sequence Repeats (SSRs) markers. Functional trait data was collected from each individual, and data on environmental variables was collected for each population. We detected significantly higher genetic diversity in the biennial species ($H E = 0.86$) compared to the perennial ($H E = 0.79$). For both species, the largest share of genetic variation was partitioned within populations (96%). Population genetic structure in the biennial *A. scoparia* was weak, while the perennial *A. frigida* showed some spatial genetic structure, which was impacted by geographical factors, soil nutrients, and precipitation. Morphology-related functional traits (i.e., plant height) were predominantly associated with environmental variables rather than with genetic variation, while physiology-related traits (i.e., specific leaf area) were partly genetically determined.





1 **Relationship between genetic and phenotypic variations**
2 **in natural populations of perennial and biennial sagebrush**

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14
15 **Abstract**

16
17 Plant responses to environmental heterogeneity depend on life-history traits, which could
18 relate to phenotypical and genetic characteristics. To elucidate this relationship, we examined the
19 variation in population genetics and functional traits of short- and long-lived *Artemisia* species
20 that are co-occurring in the steppes of Mongolia. Mongolian steppes represent stressful, water-
21 limited habitats demanding phenotypic modifications in the short term and/or genetic adaptation
22 in the long term. However, detailed knowledge is missing about both plant phenotypic and genetic
23 differentiation and their inter-relationships in temperate grasslands. Here, we investigated 21
24 populations of the widely distributed subshrub *A. frigida* and the herbaceous biennial *A. scoparia*.
25 Genetic variation was assessed with newly developed Simple Sequence Repeats (SSRs) markers.
26 Functional trait data was collected from each individual, and data on environmental variables was
27 collected for each population. We detected significantly higher genetic diversity in the biennial
28 species ($H_E=0.86$) compared to the perennial ($H_E=0.79$). For both species, the largest share of
29 genetic variation was partitioned within populations (96%). Population genetic structure in the
30 biennial *A. scoparia* was weak, while the perennial *A. frigida* showed some spatial genetic
31 structure, which was impacted by geographical factors, soil nutrients, and precipitation.
32 Morphology-related functional traits (i.e., plant height) were predominantly associated with
33 environmental variables rather than with genetic variation, while physiology-related traits (i.e.,
34 specific leaf area) were partly genetically determined.

35
36 Keywords: growth-form, *Artemisia*, inter-relationship among variations of genetic, functional
37 traits and environment

38 1. Introduction

39 It is widely acknowledged that a species' genetic diversity and its variation are associated
40 with life-history traits, such as life form, breeding system, seed dispersal mechanism, and
41 geographical range (Hamrick & Godt 1996; Nybom & Bartish 2000; Reisch & Bernhardt-
42 Römermann 2014). Species with outcrossing and mixed-mating systems tend to have higher levels
43 of genetic variation than selfing species (Nybom 2004). Short-lived, non-woody, self-compatible,
44 and early-successional species, i.e., annuals/biennials, are characterized by higher genetic variation
45 between populations, but lower genetic variation within populations. In contrast, long-lived,
46 woody, outcrossing, and late-successional species, i.e., many perennials, have higher genetic
47 variation within populations (Reisch & Bernhardt-Römermann 2014). However, comparative
48 studies such as that of Heeleman *et al.* (2015) found lower within-population variation in perennial
49 *Eriosephalus africanus* L. than in the annual species *Hemimeris racemosa* (Houtt.) Merrill. A
50 comparison of perennial and annual wild species of the genus *Oryza* L. discovered that perennial
51 species had higher population level genetic diversity but less genetic variation among populations
52 than annuals (Zhou *et al.* 2008).

53 Even within a species, plant phenotypic variation is often high. Functional trait plasticity
54 related to morphology (e.g., plant height), (eco)physiology (e.g., specific leaf area), and life history
55 (e.g., flowering time and seed traits) was found to be under genetic control in some model plants
56 (Locascio *et al.* 2009; Hughes *et al.* 2019). Several studies detected correlations between
57 phenotypic traits (morphological and functional trait variation) and genetic variation (Waite &
58 Levin 1998; Karbstein *et al.* 2019; Csilléry *et al.* 2020). In particular, Waite & Levin (1998)
59 presented a meta-study demonstrating a positive correlation between the genetic and phenotypic
60 character traits of 27 species. However, trait variation does not necessarily coincide with genetic
61 variation, especially if the trait is completely plastic (Chevin & Hoffmann 2017). Plasticity, i.e.,
62 phenotypic modification, allows for long term adaptation to the local environment and/or short
63 term (reversible) responses. However, how genetic diversity and intraspecific functional traits
64 interact at the population level, particularly in natural environments, remains poorly understood.

65 *Artemisia* L. (sagebrush) is a large and diverse genus that comprises over 500 taxa of
66 annuals/biennials, perennial herbs, and shrubs or subshrubs distributed across temperate regions
67 of the northern hemisphere (Riggins & Seigler 2012). Many species are clearly wind-pollinated;
68 however, some indication of insect pollination was observed (colorful capitula and sticky pollen;
69 Vallès & McArthur 2001). *Artemisia* spp. inhabits arid, semi-arid and mesic environments
70 spanning deserts to tundras, and their range of phenotypic diversity is broad (morphological,
71 (eco)physiological, and reproductive traits), as is their range of ploidy levels ($2n=16$ or 18 up to
72 $2n=144$; Sanz *et al.* 2008). Although the genus offers ample opportunities for comparison, studies
73 on genetic diversity and life history traits are hardly available. Al-Ajmi *et al.* (2021) compared
74 seven species of *Artemisia* and found a positive interspecific correlation between similarities in
75 genetic variation among species. However, we do not know of any study that addressed
76 intraspecific variation in traits and genetic structures.

77 *Artemisia frigida* Willd. and *A. scoparia* Waldst. & Kit. are both outbreeding and wind
78 pollinated species (Vallès et al. 2011) with a range of phenotypic variations. In this study, we
79 aimed to test the effects of environment on genetic variation and genetic structure of the short-
80 lived biennial *A. scoparia* and the long-lived sub-shrub *A. frigida*, which are co-occurring in the
81 steppes of Mongolia. The flora of Mongolia lists 103 native *Artemisia* species (Baasanmunkh *et*
82 *al.* 2022), among which species growing in dry steppes and forest steppes are the most numerous.
83 Mongolia has one of the world's largest steppes, covering 1.2 million km² and being home to
84 thousands of steppe species (Munkhzul *et al.* 2021; Baasanmunkh *et al.* 2022). The continuous
85 plain steppe of Mongolia allows for sufficient genetic exchanges between plant populations, as
86 shown by former studies on the perennial grass *Stipa glareosa* P.A.Smirn. (Oyundelger *et al.* 2020)
87 and on *Artemisia frigida* (Oyundelger *et al.* 2021, 2023). In these studies, we detected moderate
88 genetic structuring, which was mostly attributed to the differences in climate and edaphic
89 conditions of the local populations rather than the geographical distance. However, the present
90 study covers an even larger area of Mongolia, ranging from the western Altai Mountains to the
91 eastern Mongolian Steppes. Specifically, we aimed to answer the following questions: i) How do
92 genetic diversity and population structure differ between the two *Artemisia* species? ii). Do
93 environmental factors relate to the genetic variation of the species across the Mongolian steppe?
94 iii). Are functional traits related to genetic diversity and/or abiotic habitat heterogeneity?

95

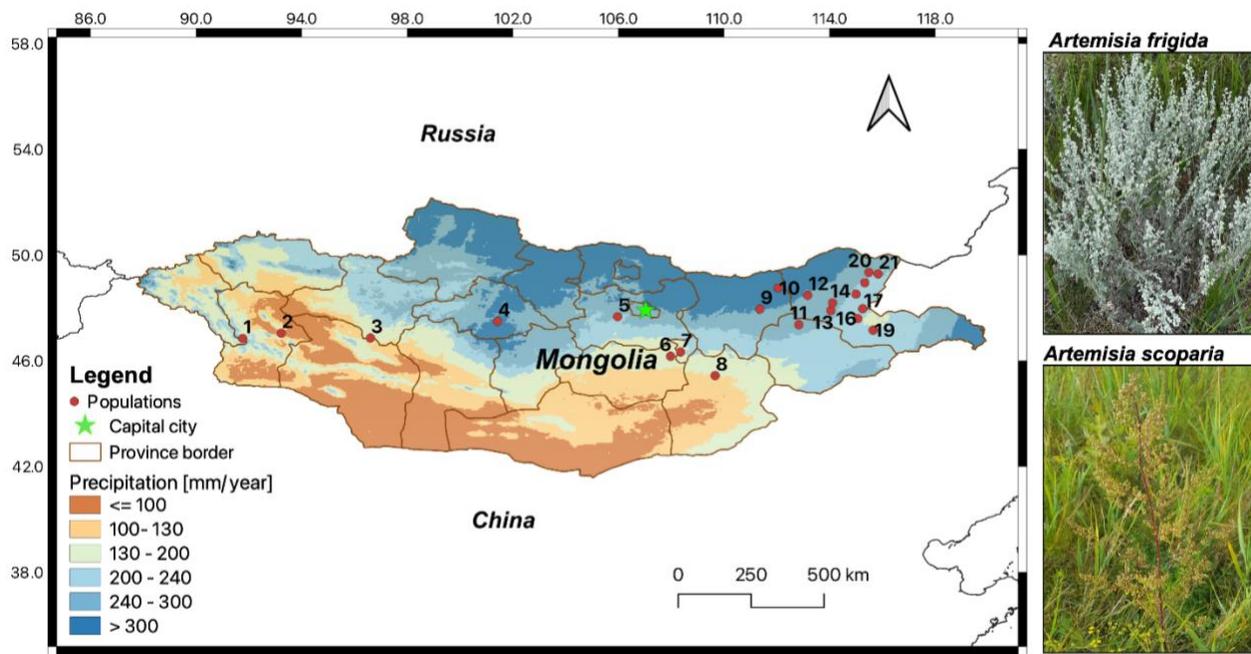
96 **2. Material and methods**

97 **2.1. Study species: *Artemisia frigida* and *A. scoparia***

98 Perennial prairie-sage (*A. frigida*) has the largest natural range within its genus, being
99 distributed across the North American prairie and the Eurasian steppe, whereas *A. scoparia* is a
100 biennial species widely distributed from Central Europe to East Asia. Species' ranges overlap in
101 Inner Asia and specifically in Mongolia, where they are common steppe plants (Hilbig 1995). They
102 share the same breeding system (outbreeding) and dispersal mechanism (wind), yet differ in their
103 life form (biennial herb vs. perennial subshrub). The perennial *A. frigida* grows primarily in
104 mountains, hillsides, and ruderal sites in steppes (Tkach *et al.* 2008). It bears a dense silvery
105 pubescence and has woody ascending stems that are usually strongly branched (Fig. 1). The
106 biennial *A. scoparia* is found in riverbanks, as well as in ruderal sites in steppes and semi-deserts.
107 Its stems are initially pubescent, becoming glabrous and strongly branched in the middle and upper
108 parts (Fig. 1). *Artemisia frigida* and *A. scoparia* are pioneer plants at sites disturbed by grazing
109 and also occur in the early recovery stages of abandoned land that underwent severe soil erosion
110 (Jiao *et al.* 2013; Wang *et al.* 2022). Both species have high seed yields and small seeds (*A. frigida*:
111 0.106 g and *A. scoparia*: 0.047 g) that are easily propagated by wind and are then buried into soils
112 (Yi *et al.* 2019).

113 *Artemisia scoparia* belongs to the subgenus *Dracunculus* Besser representing the most
114 basal lineage of *Artemisia* (clade divergence in 17.6 ± 2.1 Mya), while *A. frigida* is part of the
115 subgenus *Absinthium* DC. (clade node 6.8 ± 0.8 Mya; Sanz *et al.* 2011; Hussain *et al.* 2019).

116 *Artemisia frigida* comprises diploids ($2n = 2x = 16$) as well as tetraploids ($2n = 4x = 36$; Pellicer
 117 *et al.* 2010; Korobkov *et al.* 2014). In *A. scoparia*, mostly diploid cytotypes were observed ($2n =$
 118 $2x = 16$ or 18 ; Pellicer *et al.* 2010); yet $2n = 4x = 32$ or 36 have also been reported from Slovenia,
 119 Siberia, and recently from the Western Himalayas (Kawatani 1964; Amel’chenko 1979; Gupta *et*
 120 *al.* 2014).



121
 122 Figure 1. Study area with locations of 21 populations sampled for *A. frigida* and *A. scoparia* across Mongolia.
 123 Precipitation data were derived from Fick & Hijmans (2017).
 124

125 2.2. Study design and sampling

126 Sampling was carried out along a broad-scale longitudinal precipitation gradient from
 127 western to eastern Mongolia during the summers of 2018 and 2019 (Fig. 1). Fresh leaf materials
 128 were collected from 21 populations where both species co-occurred. For each population,
 129 representative herbarium specimens were deposited at Herbarium Senckenbergianum G rlitz
 130 (GLM). As a result, we sampled thirteen eastern (E) populations and four western (W) and four
 131 central (C) populations across various steppe vegetation types (Table 1).

132 At each site, 15 individuals per species were sampled within a 10 m \times 10 m plot. Within
 133 these plots, plant community composition and total cover (%) of vascular plants were recorded,
 134 and a sample of top soil (1 \pm 5 cm depth) with fine plant roots and the humic layer was collected.
 135 Soil samples were separated from litter, debris, and after shifting through a 2 mm sieve the
 136 following measurements were conducted in the laboratory: pH value, electrical conductivity (EC,
 137 as a proxy for salinity), plant available P, N%, organic C%, and C/N ratio. All results refer to oven-
 138 dried soil (75 $^{\circ}$ C, 18 h). Moreover, plots were classified into different steppe types according to
 139 “The steppe vegetation of Mongolia” (Tuvshintogtokh 2014) based on our sampling location,
 140 which was also validated by our field-based plant community composition data.

141 Three functional traits were measured in the same individuals sampled for molecular data.
142 In the field, ‘height of inflorescence (HI)’ (if plants were flowering), ‘height of vegetative part
143 (HV)’, and leaf area for the trait ‘specific leaf area (SLA)’ were measured. The HI was determined
144 as the height from ground level to the tip of the highest inflorescence, and the HV as height of one
145 randomly selected vegetative branch per plant. In *A. frigida*, vegetative and generative shoots
146 differ in length. Thus, both heights were chosen as traits. *Artemisia scoparia* does not develop
147 sterile shoots, and thus only the HI was applicable. For SLA, two fresh leaves were taken from
148 each individual (30 leaves per site) and scanned using a Conrad P-573 handheld document scanner.
149 Scanned pictures were later analyzed with ImageJ (Abràmoff *et al.* 2004) to determine the leaf
150 area. Leaves were then air-dried for more than a month, and biomass weight was measured with a
151 Mettler Toledo XP6 balance in the laboratory. The SLA was then calculated by dividing leaf area
152 by dry mass (Perez-Harguindeguy *et al.* 2013). Population level trait data and their correlation
153 matrices, indicating their independence are provided in Suppl. Table 1).

154 Meteorological data of 20 years (mean annual temperature (MAT), mean annual
155 precipitation (MAP), and mean spring temperature (March-May), mean summer temperature, and
156 mean summer precipitation (June-August) between 1994 – 2013) were retrieved for each locality
157 from the high-resolution CHELSA_V1 dataset, which has the advantage to capture interannual
158 precipitation variation (Karger *et al.* 2017). The coefficient of interannual variation of annual
159 precipitation (cvP) was estimated based on the retrieved MAP data and was also used as a predictor
160 since cvP is a critical driver of rangeland dynamics (von Wehrden *et al.* 2012).

161

162 **2.3. Molecular analyses and microsatellite marker development**

163 Two randomly selected individuals of each species from two distinct populations were used
164 to develop new SSR markers by applying whole genome sequencing (WGS). A previous study by
165 Oyundelger *et al.* (2021), gives detailed steps for DNA extraction, library preparation, quality
166 control and bioinformatics in SSR development. Raw sequencing data were submitted to the NCBI
167 Sequence Read Archive (SRA) and made publicly accessible under BioProject: PRJNA680535.

168 A total of 20 and 21 SSR markers were then tested for optimization in *A. frigida* and
169 *A. scoparia*, respectively, using randomly selected samples from more than ten populations
170 containing 8–16 samples. Furthermore, cross-checking of markers for both species was performed,
171 and ten SSR markers published for *A. frigida* in the master thesis of Wang (2011) were tested with
172 our samples in parallel. Based on reproducibility and polymorphism, 11 markers were chosen for
173 each species. Detailed information on SSR markers of *A. frigida* can be found from Oyundelger *et*
174 *al.* (2021). Information about species-specific SSR markers for *A. scoparia* developed for this
175 study are presented in Table 2. Amplifications of a total of 22 SSR markers were performed in a
176 volume of 12.5µl, and customized PCR reaction mixtures and cycling programs were used (see
177 PCR details from Suppl. Table 2). Individuals of all 21 populations from both species exhibited a
178 maximum of four alleles per locus, indicating prevailing tetraploidy (see Suppl. Table 3 for ploidy
179 information).

180 Table 1. Characteristics of the study sites: (population code, localities, main climatic variables, steppe vegetation type and region).

Pop code	Locality and province	Longitude	Latitude	Altitude [m]	MAT [°C]	MAP [mm]	Summer temp. [°C]	Summer prec. [mm]	cvP [%]	Steppe type	Region
1	Munkhkhairkhan, Khovd	91.765	46.841	1781	-6.1	147	8.9	87	33	MoS	W
2	Center of Khovd, Khovd	93.228	47.044	1355	2.6	115	19.5	77	42	DrS	W
3	Taishir Soum, Gobi-Altai	96.605	46.860	2009	-1.6	172	14.7	105	29	DrS	W
4	Khotont Soum, Arkhangai	101.421	47.492	1608	-1.3	300	14.3	198	24	MoS	W
5	Hustai National Park, Tuv	105.968	47.666	1264	0.9	167	18.3	118	24	MoS	C
6	Tsagaandelger, Dundgovi	107.975	46.170	1280	2.2	117	19.9	82	42	DrS	C
7	Choir, Dundgovi	108.350	46.331	1270	1.9	135	19.7	92	40	DrS	C
8	Altanshiree, Dundgovi	109.660	45.438	1007	3.8	126	21.9	84	31	DeS	C
9	Batnorov, Khentii	111.357	47.955	1078	0.5	286	18.8	194	25	DrS	E
10	Norovlin, Dornod	112.044	48.751	1020	0.6	277	18.6	192	29	DrS	E
11	Hulunbuir, Khentii	112.831	47.364	1008	0.4	231	18.6	164	37	DrS	E
12	Tsagaan-Ovoo, Dornod	113.167	48.480	1009	1.4	240	19.6	166	34	DrS	E
13	Bulgan, Dornod	114.046	47.896	961	1.5	238	19.9	163	46	DrS	E
14	Bayantumen, Dornod	114.109	48.190	991	1.4	210	19.7	142	44	DrS	E
15	Choibalsan, Dornod	114.997	48.519	847	1.5	209	20.3	141	49	DrS	E
16	Matad, Dornod	115.062	47.598	761	2.1	184	20.5	130	52	DrS	E
17	Matad, Dornod	115.250	47.971	1075	1.4	198	19.9	139	53	DrS	E
18	Choibalsan, Dornod	115.331	48.953	909	1.0	231	19.9	153	46	DrS	E
19	Shar-Khudag, Dornod	115.646	47.157	1011	1.5	199	19.7	144	52	DrS	E
20	64n toochig, Dornod	115.485	49.344	650	1.3	232	20.4	154	45	DrS	E
21	Otor pasture, Dornod	115.837	49.288	821	1.1	239	20.3	159	44	DrS	E

181 MAT – mean annual temperature, MAP – mean annual precipitation, Summer temp. – summer mean annual temperature, Summer prec. – summer
182 mean annual precipitation, cvP – coefficient of variation of interannual precipitation, MoS – mountain steppe, DrS – dry steppe, DeS – desert
183 steppe, W – western, C – central, E – eastern region of Mongolia, coordinates are in WGS84

184 Table 2. Characterization of eleven polymorphic microsatellite markers used in this study for *Artemisia*
 185 *scoparia*. Details on SSR markers for *A. frigida* can be found in Oyundelger *et al.* (2021).
 186

No.	Locus	Repeat motif	Primer sequences (5'-3')	Ta (°C)	Allele size range (bp)	Fluorescent dye	PCR type
1	<i>Arcs2</i>	(GT)9	F: TGTA AACGACGGCCAGTTCTC CTTTCTGATTCATTGG R: CGAGATGAATTTGCGTCAT	55	585-620	6 FAM	Multiplex
2	<i>Arcs12</i>	(TGT)9	F: TGTA AACGACGGCCAGTGGAC ATTTGAATGATGTTTCG R: AAGTCTTCCGCCAGCTATA	55	200-265	6 FAM	
3	<i>Arcs7</i>	(TG)11	F: TGTA AACGACGGCCAGTTGT CCATCAAGATACCTATGC GGTTATCGCCTCTCATTG	55	520-560	VIC	Multiplex
4	<i>Arcs11</i>	(ACA)8	F: TGTA AACGACGGCCAGTGAAC GGGAAGATTACAAGC R: CACCAATATTACCTGGTGTG	55	130-180	VIC	
5	<i>Arcs18</i>	(ATG)8	F: TGTA AACGACGGCCAGTACAC TGGAAAGCTATGTGC R: CGAGTCACAGTCATGGTC	55	610-660	PET	Multiplex
6	<i>Arcs19</i>	(TGA)8	F: TGTA AACGACGGCCAGTCCT CAAACCTTGAAAGATAGC R: CCGTATGAGTTAAGCAATCAG	55	350-400	PET	
7	<i>Arcs17</i>	(TGA)8	F: TGTA AACGACGGCCAGTAATG GATTATGTTGATAGCCA R: CAAGTTCCGTTGACTCG	55	135-160	6 FAM	Singleplex
8	<i>Arcs14</i>	(ATA)8	F: TGTA AACGACGGCCAGTATG CACATAATATCCGAGC R: GTGCTGAGACCGAATGC	55	270-325	VIC	Singleplex
9	<i>Arcs20</i>	(ACA)14	F: TGTA AACGACGGCCAGTGAC ACCCATAGACAGGAGC R: GTCAGCTCGAAGCTTTCC	55	~500	NED	Singleplex
10	<i>Arcs21</i>	(TGT)8	F: TGTA AACGACGGCCAGTTGC CTTTGCAACAATTAAC R: GCTGCAAACATTACGTAAGC	55	110-128	NED	Singleplex
11	<i>Ch468</i>	NA	F: TGTA AACGACGGCCAGTTAG GGTTGCAGAAGATAAAC R: GCTTCTTCACTTCCACTAAAG	55	160-236	PET	Singleplex

187
 188 **2.4. Statical analyses**
 189 ***Analysis of genetic diversity and population structure***
 190 To compare the genetic diversity within each species, we employed two programs, which
 191 allowed handling of microsatellite data for polyploids and species with mixed ploidy: GenoDive
 192 v.3.04 (Meirmans 2020) and the R-package *Polysat* v. 1.7 (Clark & Jasieniuk 2011) in R v.4.0.3
 193 (R Core Team 2020). Estimators of genetic diversity comprised allelic diversity (AD), percentage
 194 of polymorphic loci (PPL), observed heterozygosity (H_O), expected heterozygosity (H_E) and
 195 inbreeding coefficient (G_{IS}), all of which were calculated using GenoDive. Bruvo distances were
 196 computed with the R-package *Polysat* v.1.7 (Bruvo *et al.*, 2004). Using the R-package *vegan*
 197 (Oksanen *et al.*, 2007), we calculated the mean Bruvo distance among individuals for any given

198 population (hereafter ‘Bruvo index’; see detail in Oyundelger *et al.* (2021)), which was then used
199 as a surrogate for genetic diversity (See Suppl. Table 4 for the genetic diversity indices). A paired
200 T-test was used to determine the significance of the difference in genetic diversity indices between
201 two species.

202 Coefficients of genetic differentiation (F_{ST} and G_{ST}) were estimated using *Polysat* (Suppl.
203 Table 5). Population genetic structure was further analyzed with Principal Coordinate Analysis
204 (PCoA) using population-wise F_{ST} distance using the R-package *ape* (Paradis & Schliep 2019). In
205 order to reveal environmental variables that were significantly associated with population genetic
206 structure of the species, environmental and vegetation variables were fitted *post hoc* on the
207 ordination using *vegan*, and plots were visualized with *ggplot2* (Wickham 2011).

208 To examine the partitioning of genetic variation between and within populations, Analysis
209 of Molecular Variance (AMOVA; Excoffier *et al.* 1992) was performed in R-package *poppr*
210 (Kamvar *et al.* 2014) based on the individual level Bruvo distance matrix estimated with *Polysat*.

211

212 ***Relationship between genetic and spatial distances***

213 To assess the overall relationship between genetic and spatial distances, Mantel tests
214 between genetic distance (linearized population level pairwise F_{ST} ($F_{ST}/(1-F_{ST})$)) and geographic
215 distances (Euclidean distances) were computed through 10,000 randomizations using the R-
216 package *vegan* (Oksanen *et al.*, 2007). Further Mantel tests were then conducted between genetic
217 distances and a) climatic differences (Euclidean distance of centred and standardized climatic
218 variables); b) distance of soil indicator variables (Euclidean distance of centred and standardized
219 variables), and c) differences in plant community composition (Bray-Curtis’s distance based on
220 log-transformed species’ cover).

221

222 ***Relationships of functional trait variation with genetic and environmental patterns***

223 We estimated population-level means and coefficients of variation (CV) for trait variables,
224 the latter as the ratio of standard deviation to mean. We checked collinearity among traits (mean
225 and CV) with Pearson’s coefficient (Suppl. Table 1) using the R-package *corrplot* (Wei *et al.*
226 2021). As correlation coefficient values (r) of the mean and CVs were below $\sim |.7|$, we did not
227 exclude particular functional traits.

228 To assess whether functional traits are related to environmental heterogeneity and genetic
229 diversity, we fitted linear models (Dobson & Barnett 2018) with mean and CV of traits as the
230 dependent variables. We again used *corrplot* for an exploratory analysis of associations among
231 measures of genetic diversity. As a result, H_E was chosen as main response variable, as it had the
232 highest correlation and depends less on population history (e.g., bottlenecks) compared to the other
233 indices (Rosenberg 2004; Szczecińska *et al.* 2016). For the predictors, we first checked
234 correlations among environmental variables to select representative variables based on their
235 importance and independencies ($r < |.7|$; See Suppl. Table 6 for the data and their correlations).
236 As a result: MAP, MAT and cvP for climate; altitude for topography, and soil C/N ratio for soil
237 nutrient contents were initially used as predictors for the models.

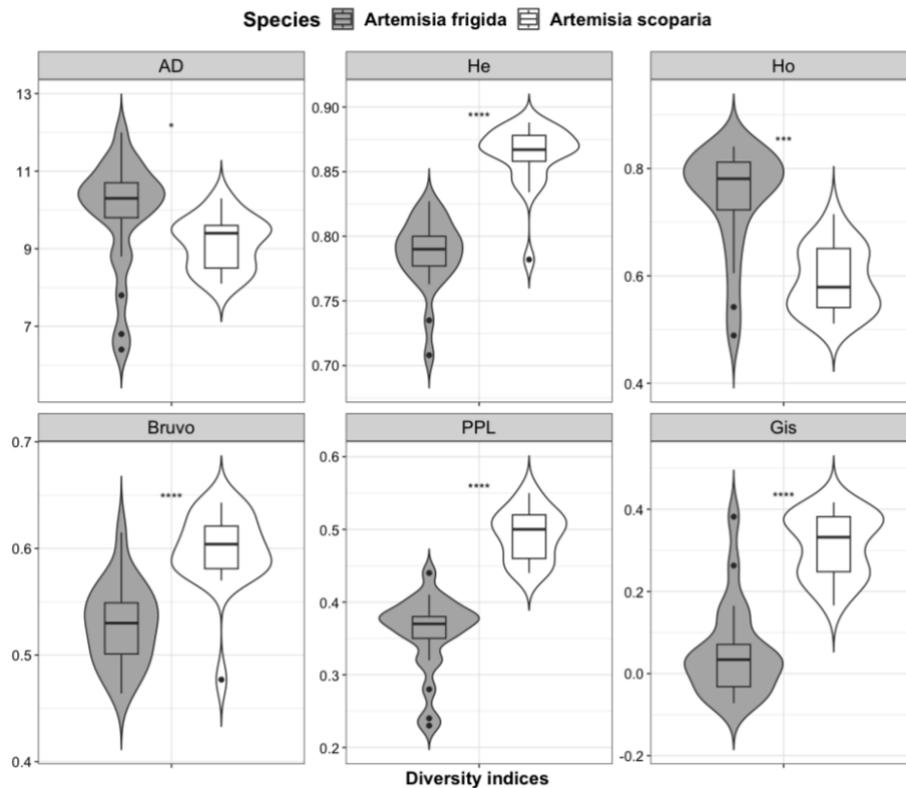
238 All predictors were first scaled to zero mean – unit variance (z-scores) to make effect sizes
 239 comparable. The response variable: cvIH of *A. frigida* was log-transformed due to its non-normal
 240 distribution; other response variables (cv and means) were in normal distribution, and thus no
 241 transformation was done. We then conducted model simplification by dropping the least relevant
 242 variables from linear models until a null model with intercept only. Models were compared using
 243 ANOVA, the summary was used to estimate significances and to choose the most parsimonious
 244 models. Lastly, plotting was used to check residuals of the models for possible deviations from
 245 normality and reasonable distribution of variances.

246

247 3. Results

248 3.1. Comparison of genetic diversity between the perennial and biennial *Artemisia*

249 The overall polymorphic information content (PIC) of newly developed species-specific
 250 SSR markers was high (PIC=0.77 and 0.84) for both *A. frigida* and *A. scoparia*. Paired T-test
 251 revealed that proxies of genetic diversity differed between two the *Artemisia* species (Fig. 2).
 252 Specifically, H_E , Bruvo, PPL and G_{IS} of the biennial *A. scoparia* was significantly higher than in
 253 the perennial *A. frigida*. In contrast, AD and H_o were larger in the perennial than the annual
 254 species, yet with lower significance. Details for estimators of genetic diversity are presented in
 255 Suppl. Table 4.



256

257 Figure 2. Violin boxplots of genetic diversity indices of the perennial *A. frigida* (N= 304) and the biennial *A. scoparia*
 258 (N=303) (AD – Allelic diversity, H_E –expected heterozygosity, H_o – observed heterozygosity, Bruvo – Bruvo index,
 259 PPL – percentage of polymorphic loci and G_{IS} – inbreeding coefficient). Significance codes: $p \leq 0.0001$ ‘****’; $p <$
 260 0.001 ‘***’; $p < 0.05$ ‘*’.

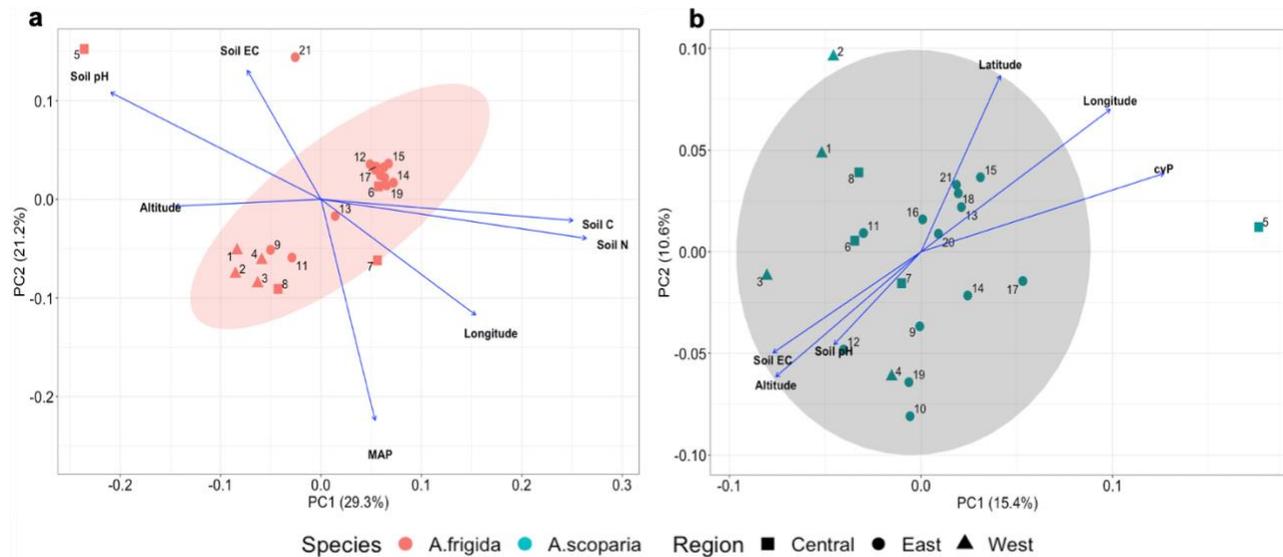
261 **3.2. Population genetic variation and relationship with environmental variables**

262 Coefficients of genetic differentiation of the two species across 21 populations were low overall,
 263 suggesting that isolation is at most moderate over the distances considered here. However,
 264 population differentiation of *A. frigida* was slightly more pronounced (Global F_{ST} = 0.078 and
 265 Global G_{ST} = 0.071) than of *A. scoparia* (Global F_{ST} = 0.064 and Global G_{ST} = 0.055). The most
 266 genetically distant population was population 5 (Hustai National Park) in both species
 267 (dissimilarity data provided in Suppl. Table 5). Analysis of Molecular Variance showed that in
 268 both species, highest genetic variation resided between individuals, while genetic variation
 269 partitioned among regions was slightly higher in *A. frigida* than *A. scoparia* (0.97% and 0.83%,
 270 respectively; Table 3). The ordination plots suggested that there was no pronounced genetic
 271 differentiation among steppe types and regions of Mongolia (individual level PCoA in Suppl.
 272 Table 7), although *A. frigida* exhibited some population level genetic structure (Fig. 3). In the
 273 PCoA ordination of *A. frigida*, the first two axes explained about 50 % of the genetic variation,
 274 and some structuring of eastern vs. western populations mixed with central populations was
 275 discernable. According to *post hoc* fitting of predictor variables, longitude, altitude, mean annual
 276 precipitation (MAP), soil carbon, nitrogen, pH and soil electrical conductivity (EC) showed a
 277 significant association with genetic structure (Fig. 3a). In total 26 % of the total genetic variation
 278 was explained by the first two axes in the populations of *A. scoparia*, representing more continuous
 279 patterns among populations. Main structures along axis 1 and 2 were significantly correlated with
 280 altitude, soil pH and EC together with longitude, latitude and coefficient of variation of interannual
 281 precipitation (cvP), with western populations being in the upper left (Fig. 3b). The ordinations
 282 demonstrated that soil pH and EC, as well as soil C and N, exhibit covariance, as proven by their
 283 high correlations ($r=0.78$ and $r=0.99$; Suppl. Table 6). Results of *post hoc* fitting predictor variables
 284 on the PCoA are provided in the Suppl. Table 8.

285
 286 Table 3. Summary of Analysis of Molecular Variance (AMOVA) of the perennial *Artemisia frigida* and the
 287 biennial *A. scoparia* of 21 populations across Mongolia.

Source of variance	Df	Sum sq	Variance component	% Total		Φ statistic
<i>Artemisia frigida</i>						
Between regions	2	12..56	0.028	0.97	***	0.037
Between populations	18	72.03	0.080	2.69	***	0.027
Within populations	283	806.44	2.850	96.34	***	0.009
Total	303	891.03	2.958			
<i>Artemisia scoparia</i>						
Between regions	2	2.21	0.004	0.83	***	0.035
Between populations	18	11.25	0.013	2.69	***	0.027
Within populations	282	128.68	0.456	96.47	***	0.008
Total	302	142.14	0.473			

288 Df – degrees of freedom, Sum Sq – Sum of square, % total – percentage of variation



289

290 Figure 3. Principal Coordinate Analyses (PCoA) based on F_{ST} distances of the a) perennial *Artemisia frigidida* and
 291 b) the biennial *A. scoparia* among 21 populations across three regions (east, central and west) of Mongolia. Each
 292 symbol represents one population, and 95% confidence intervals are indicated by shaded area. Environmental
 293 predictors were fitted *post hoc* on the ordination plot (only those that passed $p < 0.05$ according to a test with
 294 1,000 permutations are shown). Result of *post hoc* analyses indicating the importance of environmental variables
 295 are provided in Suppl. Table 8.

296

297 The Mantel tests on association between genetic structures (linearized $F_{ST} - F_{ST}/(1-F_{ST})$) with
 298 various environmental variable distances revealed an overall negligible relationship with the
 299 genetic distances in both species (Suppl. Table 9). Geographic distance and the distance of soil
 300 nutrient values in particular showed a significant but weak correlation with the genetic distance of
 301 *A. frigidida* ($r^2 = 0.05^{***}$ and $r^2 = 0.02^*$). In contrast, *A. scoparia* did not exhibit an isolation by
 302 distance effect, while a weak correlation with climatic distance was observed.

303 3.3. Associations of functional traits with genetic and environmental variations

304 Results of linear models showed that means as well as coefficients of variation of functional traits
 305 in *A. frigidida* were associated with climatic and geographic variables, whereas in *A. scoparia*
 306 genetic diversity and soil nutrients had a significant relationship with SLA (Table 4). In the
 307 perennial *A. frigidida*, altitude was positively associated to the physiology related trait (mean SLA),
 308 while variations of morphology related traits, cvHI and cvVH were significantly affected by MAT,
 309 MAP, and cvP. In the biennial *A. scoparia*, genetic diversity showed an association with mean
 310 SLA, and soil nutrient contents with the variation of SLA. With the exception of altitude and cvP,
 311 all significant associations were negative (scatter plots with linear regression line of the significant
 312 models are provided Suppl. Table 10 and 11).

313

314

315 Table 4. Summary of the retained parsimonious and significant linear models assessing the associations of
 316 functional traits of *A. frigidida* and *A. scoparia* with the genetic diversity and environmental variables.

	Functional traits	Predictor	Estimate	Std. Error	Pr(> t)	
<i>A. frigida</i>	Mean SLA	(Intercept)	0.15	0.006	<0.001	***
		Altitude	0.02	0.006	0.012	*
	CV height of inflorescence	(Intercept)	1.29	0.022	<0.001	***
		MAP	-0.09	0.023	0.002	**
		MAT	-0.08	0.025	0.005	**
	CV height of vegetative part	cvP	0.07	0.025	0.01	**
(Intercept)		31.78	1.652	<0.001	***	
<i>A. scoparia</i>	Mean SLA	MAT	-3.86	1.693	0.034	*
		(Intercept)	0.15	0.007	<0.001	***
	CV SLA	H _E	-0.03	0.007	0.001	***
		(Intercept)	1.61	0.037	<0.001	***
		Soil C/N	-9.06	3.602	0.021	*

317 Pr(>|t|) – significance p-value. Significance codes: p ≤ 0.001 ‘***’; p < 0.01 ‘**’; p ≤ 0.05 ‘*’; p ≤ 0.1 ‘.’.

318 4. Discussion

319 4.1. Population genetic diversity and differentiation of *A. frigida* and *A. scoparia*

320

321 Life form and breeding system of plants are known to have a major influence on species'
322 genetic diversity and population genetic structure (see Nybom & Bartish 2000; Reisch &
323 Bernhardt-Römermann 2014; De Kort *et al.* 2021). Our chosen *Artemisia* species both have a wide
324 range of distribution, are wind/water dispersed, outcrossing, and had prevailing tetraploid
325 cytotypes, making a direct comparison of diversity indices possible. Population-level mean values
326 of the genetic diversity in both *Artemisia* species were higher (*A.f.*: H_E = 0.79 and *A.s.*: H_E = 0.86)
327 than in the review of Nybom (2004) for similar life history traits. The genetic diversity was
328 significantly higher in the biennial *A. scoparia* than in the perennial species, according to four of
329 the six diversity indices (H_E, G_{IS}, Bruvo, and PPL; Fig. 2). This is in line with the study of
330 Balfourier *et al.* (1998), who compared outcrossing annual and perennial ryegrass (*Lolium* L.)
331 species. Probably, the effective population size and recombination rate are higher in the biennial
332 than in the perennial. In short-lived species, recombination rate is higher as a result of their shorter
333 life cycles and smaller genome/ lower DNA content (Brazier & Glémin 2022), which may lead to
334 a higher level of genetic diversity. Indeed, Garcia *et al.* (2004) reported that genome size of
335 *A. scoparia* was the smallest (1C = 1.77 pg) within the studied species, while the genome size of
336 *A. frigida* was 2.63 pg. Furthermore, in *A. frigida*, a smaller number of plants may participate in
337 reproduction, as it is often subject to intensive grazing in natural and permanent pastures, and some
338 individuals may survive vegetatively over several seasons. However, this observation is in contrast
339 to some review studies that compared the genetic diversity of different life forms, utilizing
340 allozyme and RAPD markers (see Hamrick & Godt 1990, 1996; Nybom & Bartish 2000; Nybom
341 2004) and AFLP markers (Balfourier *et al.* 1998; Reisch & Bernhardt-Römermann 2014).
342 Nonetheless, individual life history traits, as well as genetic markers and diversity indices utilized
343 affect estimates of population genetic diversity, making the direct comparisons among studies
344 somewhat questionable.

345 Patterns of genetic variation in the two species did not differ much, with spatial differences
346 (among regions) explaining about 1% of the genetic variation, while barely 2-3% variation resided
347 among populations, and the highest variation (more than 95%) was explained by within-population
348 variations (Table 3). Yet, the populations of the perennial *A. frigida* represented some structure
349 illustrated in the PCoA, having fuzzy eastern and western clusters associated with altitude,
350 longitude, amount of precipitation, and soil salinity (Fig. 3a). Patterns in the biennial species were
351 more continuous and impacted by geographical factors, like longitude, latitude, and altitude, as
352 well as the coefficient of interannual precipitation variation (Fig. 3b). Population 5 (Hustai NP) is
353 a geographically central population that, however, represented the greatest genetic distance from
354 others in both species (see PCoA; Fig. 3 and Suppl. Table 5 for differentiation matrices). This
355 pattern has been seen in our former studies (see Oyundelger *et al.* 2021, 2023), and is now
356 supported by the analysis of a second species, indicating this region has a distinct regime of gene
357 flow and/or population connectivity, most likely due to its proximity to the local livestock trade
358 center where animals from all over the country are brought in and may carry seeds.

359 Only few studies have compared the genetic variation of herbaceous species with different
360 life forms (perennial vs. annual) in the same spatial context (Balfourier *et al.* 1998; Zhou *et al.*
361 2008; Heelemann *et al.* 2015), but their findings were contradictory: Zhou *et al.* (2008) found the
362 highest molecular variation among populations in the annual (78%) than the perennial wild rice
363 species (52%). While Balfourier *et al.* (1998) and Heelemann *et al.* (2015) reported that most of the
364 total genetic variation was accounted for within populations in perennial (91%) and annual
365 ryegrass (90%); and wild rosemary species (perennial: 89% and annual: 87%), respectively. Our
366 result was in line with the latter, as within population variations were as high as 96% in both
367 species. Furthermore, genetic variation between populations of the perennial was only marginally
368 higher than that of annual species; yet both were comparably low. The low level of genetic
369 variation between populations and regions, as well as weak correlations between genetic
370 differences with environmental distances, indicate considerable historical and current gene flow
371 between populations, supporting our former studies (Oyundelger *et al.* 2021, 2023).

372

373 4.2. Associations of functional traits with genetic and environmental variations

374

375 Mean values as well as variations of morphology- (IH and VH) and (eco)physiology-
376 (SLA) related traits were predominantly associated with environmental variables rather than with
377 genetic variation (Table 4). This indicates that the traits showed substantial plasticity in response
378 to environmental differences, as demonstrated by a number of other studies (see Gratani 2014;
379 Chevin & Hoffmann 2017; Matesanz & Ramírez-Valiente 2019). Specifically, climate (MAP,
380 MAT, and cvP) was found to be the most important factor influencing the morphological trait
381 variations of the perennial *Artemisia*. This, of course, indicates the importance of climatic
382 conditions for plant growth, as has been previously shown for plant species occurrence and
383 abundance in the Mongolian steppe (von Wehrden & Wesche 2007; von Wehrden *et al.* 2010). In
384 *A. frigida*, morphological differentiation is probably promoted by site-dependent microhabitat

385 differences, primarily in temperature and water availability. Morphological differences become
386 even more pronounced, particularly due to the harsh climate in steppes (MAT: min (-6.1) to max
387 +3.8 C°) with overall limited water availability (MAP: min 117 mm to max 300 mm), as
388 demonstrated by our linear model (Table 4). Phenotypic differences were pronounced between
389 sites/populations, whereas genetic differentiation was less evident (Global $F_{ST} = 0.064$). This is in
390 line with a large body of literature showing plant phenotypic trait responses and genetic
391 differentiation patterns varying highly in abiotic and biotic environmental conditions (Odat *et al.*
392 2004; Bucher *et al.* 2016; König *et al.* 2018), and plant trait differentiations being even enhanced
393 in extreme environments (Chevin & Hoffmann 2017; Karbstein *et al.* 2019).

394 Specific leaf area (SLA) relates to photosynthesis, relative growth rate, and stress tolerance
395 (Perez-Harguindeguy *et al.* 2013), and is known to be subject to substantial plasticity (Pan *et al.*
396 2013; Stotz *et al.* 2022) as well as being partly under genetic control (Knight & Ackerly 2003;
397 Scheepens *et al.* 2010). In our study, mean SLA was significantly associated with altitude in
398 *A. frigida* and with genetic diversity in *A. scoparia*. Soil nutrient availability also had a significant
399 impact on the variation of the SLA in *A. scoparia*, supporting the common observations, as we
400 detected the effect of both environment and genetics on SLA (Table 4). Significant relationships
401 of the mean and cvSLA with environmental variables were observed in other studies. For instance,
402 Woodward (1983) noted a negative association between altitude and SLA in *Festuca* L. and *Carex*
403 L. species, which was explained by an underlying relationship between altitude and temperature.
404 Yulin *et al.* (2005) detected an increasing SLA in habitats with higher amounts of soil nutrients
405 (total nitrogen and organic carbon) in *Artemisia halodendron* Turcz. ex Besser, as soil nutrient
406 stress is a major limiting factor for plant growth. A global study has shown a positive association
407 between soil fertility and SLA, whereas negative relationships exist between soil C/N ratio and
408 SLA (Ordoñez *et al.* 2009), supporting our findings. Furthermore, genetic effects on SLA variance
409 were observed in *Campanula* L. (Scheepens *et al.* 2010), which were attributed to selection-
410 induced adaptations. The same may hold true for our observation that genetically less diverse
411 populations represented a larger mean SLA, as a result of local adaptation. Yet, this negative
412 association might be rather an artifact attributed to the (natural outlier) population 5 (Hustai NP),
413 where the lowest population level diversity ($H_E = 0.78$) and the largest mean specific leaf area
414 (SLA = 0.24 mm/mg) were detected (see relationship in Suppl. Table 10).

415 **Conclusion**

416 Understanding plant adaptation — both in terms of morphological and genetic aspects — to
417 environmental heterogeneity has been a focal point of many studies. However, steppe plants have
418 rarely been investigated, and no comparative studies of species with different life-history traits
419 have been conducted to date. Our findings demonstrated that genetic diversity in both species was
420 relatively high (*A.f.*: $H_E = 0.79$ and *A.s.*: $H_E = 0.86$), and their genetic variation and functional trait
421 characteristics were significantly affected by geographical factors and soil nutrient contents.
422 Surprisingly, climatic factors exhibited a relatively limited impact, and when there was an effect,
423 it was primarily associated with the amount and variation of precipitation. This aligns with the
424 overarching observation in Mongolia that precipitation serves as the primary limiting factor for

425 plant growth, occurrence, and abundance. Thus, plants in these areas require significant
426 adaptations to thrive in the water-limiting habitats while retaining sufficient genetic diversity.
427

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442

443 **Author contributions**

444 All authors contributed to this work, i.e., study conception and design, sample collection and
445 vegetation surveys including species identification were performed by CMR, KW, BO and KO.
446 Library construction and bioinformatics were done by DH and VH. DNA extractions,
447 microsatellite analyses and statistics were done by LG and KO. The first draft of the manuscript
448 was written by KO and all authors commented on previous versions of the manuscript. All authors
449 read and approved the final manuscript.
450

451 **Data accessibility**

452 WGS raw sequencing data is available in the NCBI Sequence Read Archive (SRA) under
453 BioProject PRJNA680535. Further dataset generated and analyzed during the current study are
454 provided in the Supplement material tables.
455

456 **Declaration of competing interest**

457 The authors declare the following financial interests/personal relationships which may be
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463 **References**

464
465 Abràmoff, M.D., Magalhães, P.J. & Ram, S.J. (2004) Image processing with ImageJ. *Biophotonics*

466 *International* 11: 36–42.

467 Al-Ajmi, A.H., AL-Wahibi, M.S., Mustafa, A.E.-Z., Soliman, D.A. & Dewir, Y.H. (2021) Morphological
468 and molecular assessment of genetic diversity of seven species of the genus *Artemisia* L.(Asteraceae).
469 *Arabian Journal for Science and Engineering* 46: 5361–5371.

470 Amelchenko, V.P. (1979) Contribution to study of *Artemisia* from Yenisei group. *New data on the Siberian*
471 *nature*. [Амельченко В.П. 1979. К изучению полыней Приенисейской группы // Новые данные
472 о природе Сибири. Томск. С. 114–118, Tomsk, pp. 114–118].

473 Baasanmunkh, S., Urgamal, M., Oyuntsetseg, B., Sukhorukov, A.P., Tsegmed, Z., Son, D.C., Erst, A.,
474 Oyundelger, K., Kechaykin, A.A. & Norris, J. (2022) Flora of Mongolia: annotated checklist of native
475 vascular plants. *PhytoKeys* 192: 63–169.

476 Balfourier, F., Charmet, G. & Ravel, C. (1998) Genetic differentiation within and between natural
477 populations of perennial and annual ryegrass (*Lolium perenne* and *L. rigidum*). *Heredity* 81: 100–110.

478 Brazier, T. & Glémin, S. (2022) Diversity and determinants of recombination landscapes in flowering
479 plants. *PloS Genetics* 18: e1010141.

480 Bucher, S.F., Auerswald, K., Tautenhahn, S., Geiger, A., Otto, J., Müller, A. & Römermann, C. (2016)
481 Inter- and intraspecific variation in stomatal pore area index along elevational gradients and its relation
482 to leaf functional traits. *Plant Ecology* 217: 229–240.

483 Chevin, L.-M. & Hoffmann, A.A. (2017) Evolution of phenotypic plasticity in extreme environments.
484 *Philosophical Transactions of the Royal Society B: Biological Sciences* 372: 20160138.

485 Clark, L. V & Jasieniuk, M. (2011) POLYSAT: an R package for polyploid microsatellite analysis.
486 *Molecular Ecology Resources* 11: 562–566.

487 Csilléry, K., Ovaskainen, O., Sperisen, C., Buchmann, N., Widmer, A. & Gugerli, F. (2020) Adaptation to
488 local climate in multi-trait space: evidence from silver fir (*Abies alba* Mill.) populations across a
489 heterogeneous environment. *Heredity* 124: 77–92.

490 Dobson, A.J. & Barnett, A.G. (2018) *An introduction to generalized linear models*. C. Chatfield & J. Zidek
491 (eds.). CRC press company, Boca Raton, London, New York, Washington, D.C

492 Excoffier, L., Smouse, P.E. & Quattro, J.M. (1992) Analysis of molecular variance inferred from metric
493 distances among DNA haplotypes: application to human mitochondrial DNA restriction data.
494 *Genetics* 131: 479–491.

495 Fick, S.E. & Hijmans, R.J. (2017) WorldClim 2: new 1-km spatial resolution climate surfaces for global
496 land areas. *International Journal of Climatology* 37: 4302–4315.

497 Garcia, S., Sanz, M., Garnatje, T., Kreitschitz, A., McArthur, E.D. & Vallès, J. (2004) Variation of DNA
498 amount in 47 populations of the subtribe Artemisiinae and related taxa (Asteraceae, Anthemideae):
499 karyological, ecological, and systematic implications. *Genome* 47: 1004–1014.

500 Gratani, L. (2014) Plant phenotypic plasticity in response to environmental factors. *Advances in Botany*
501 2014: 208747.

502 Gupta, R.C., Goyal, H. & Singh, V. (2014) Cytology of the genus *Artemisia* (Anthemidae, Asteraceae) in
503 the Western Himalayas. *Biologia* 69: 1134–1141.

504 Hamrick, J.L. & Godt, M.J. (1990) Allozyme diversity in plant species. In: H. D. A. Brown, T. M. Clegg,
505 L. A. Kahler, & S. B. Weir (eds.) *Plant Population Genetics, Breeding and Genetic Resources*.
506 Sinauer, Sunderland, MA, USA, pp. 43–63.

507 Hamrick, J.L. & Godt, M.J.W. (1996) Effects of life history traits on genetic diversity in plant species.
508 *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences* 351: 1291–
509 1298.

510 Heelemann, S., Bäuerlein, V., Krug, C.B., Esler, K.J., Poschlod, P. & Reisch, C. (2015) Genetic variation
511 of two species with different life-history traits in the endangered renosterveld of South Africa—a
512 comparative analysis of *Eriocephalus africanus* and *Hemimeris racemosa*. *African Journal of Ecology*
513 53: 447–453.

514 Hilbig, W. (1995) *The Vegetation of Mongolia*. SPB Academic Publishing, Ulaanbaatar Mongolia.

515 Hughes, P.W., Soppe, W.J.J. & Albani, M.C. (2019) Seed traits are pleiotropically regulated by the
516 flowering time gene PERPETUAL FLOWERING 1 (PEP1) in the perennial *Arabis alpina*. *Molecular*

- 517 *Ecology* 28: 1183–1201.
- 518 Hussain, A., Potter, D., Kim, S., Hayat, M.Q. & Bokhari, S.A.I. (2019) Molecular phylogeny of *Artemisia*
519 (Asteraceae-Anthemideae) with emphasis on undescribed taxa from Gilgit-Baltistan (Pakistan) based
520 on nrDNA (ITS and ETS) and cpDNA (psbA-trnH) sequences. *Plant Ecology and Evolution* 152:
521 507–520.
- 522 Jiao, J., Han, L., Jia, Y., Lei, D., Wang, N. & Li, L. (2013) Seed morphology characteristics in relation to
523 seed loss by water erosion in the Loess Plateau. *SpringerPlus* 2: S9.
- 524 Kamvar, Z.N., Tabima, J.F. & Grünwald, N.J. (2014) Poppr: an R package for genetic analysis of
525 populations with clonal, partially clonal, and/or sexual reproduction. *PeerJ* 2: e281.
- 526 Karbstein, K., Tomasello, S. & Prinz, K. (2019) Desert-like badlands and surrounding (semi-) dry
527 grasslands of Central Germany promote small-scale phenotypic and genetic differentiation in *Thymus*
528 *praecox*. *Ecology and Evolution* 9: 14066–14084.
- 529 Kawatani, T. (1964) Chromosome numbers in *Artemisia*. *Eisei Shikenjo Hokoku* 82: 183–193.
- 530 Knight, C.A. & Ackerly, D.D. (2003) Evolution and plasticity of photosynthetic thermal tolerance, specific
531 leaf area and leaf size: congeneric species from desert and coastal environments. *New Phytologist* 160:
532 337–347.
- 533 König, P., Tautenhahn, S., Cornelissen, J.H.C., Kattge, J., Bönisch, G. & Römermann, C. (2018) Advances
534 in flowering phenology across the Northern Hemisphere are explained by functional traits. *Global*
535 *Ecology and Biogeography* 27: 310–321.
- 536 Korobkov, A.A., Kotseruba, V. V & Probatova, N.S. (2014) Chromosome numbers of some species of
537 *Artemisia* L. from Altai region, South Siberia. *Botanica Pacifica: a Journal of Plant Science and*
538 *Conservation* 3: 61–66.
- 539 De Kort, H., Prunier, J.G., Ducatez, S., Honnay, O., Baguette, M., Stevens, V.M. & Blanchet, S. (2021)
540 Life history, climate and biogeography interactively affect worldwide genetic diversity of plant and
541 animal populations. *Nature Communications* 12: 516.
- 542 Locascio, A., Lucchin, M. & Varotto, S. (2009) Characterization of a MADS FLOWERING LOCUS C-
543 LIKE (MFL) sequence in *Cichorium intybus*: a comparative study of CiMFL and AtFLC reveals
544 homologies and divergences in gene function. *New Phytologist* 182: 630–643.
- 545 Matesanz, S. & Ramírez-Valiente, J.A. (2019) A review and meta-analysis of intraspecific differences in
546 phenotypic plasticity: implications to forecast plant responses to climate change. *Global Ecology and*
547 *Biogeography* 28: 1682–1694.
- 548 Meirmans, P.G. (2020) Genodive version 3.0: Easy-to-use software for the analysis of genetic data of
549 diploids and polyploids. *Molecular Ecology Resources*.
- 550 Munkhzul, O., Oyundelger, K., Narantuya, N., Tuvshintogtokh, I., Oyuntsetseg, B., Wesche, K. & Jäschke,
551 Y. (2021) Grazing effects on Mongolian steppe vegetation – a systematic review of local literature.
552 *Frontiers in Ecology and Evolution* 9: 703220.
- 553 Nybom, H. (2004) Comparison of different nuclear DNA markers for estimating intraspecific genetic
554 diversity in plants. *Molecular Ecology* 13: 1143–1155.
- 555 Nybom, H. & Bartish, I. V (2000) Effects of life history traits and sampling strategies on genetic diversity
556 estimates obtained with RAPD markers in plants. *Perspectives in Plant Ecology, Evolution and*
557 *Systematics* 3: 93–114.
- 558 Odat, N., Jetschke, G. & Hellwig, F.H. (2004) Genetic diversity of *Ranunculus acris* L. (Ranunculaceae)
559 populations in relation to species diversity and habitat type in grassland communities. *Molecular*
560 *Ecology* 13: 1251–1257.
- 561 Ordoñez, J.C., Van Bodegom, P.M., Witte, J.M., Wright, I.J., Reich, P.B. & Aerts, R. (2009) A global study
562 of relationships between leaf traits, climate and soil measures of nutrient fertility. *Global Ecology and*
563 *Biogeography* 18: 137–149.
- 564 Oyundelger, K., Herklotz, V., Harpke, D., Oyuntsetseg, B., Wesche, K. & Ritz, C. (2021) Contrasting
565 effects of local environment and grazing pressure on genetic diversity and structure of *Artemisia*
566 *frigida*. *Conservation Genetics* 22: 947–962.
- 567 Oyundelger, K., Munkhzul, O., Ritz, C.M. & Wesche, K. (2023) Long-term grazing exclusion effects

568 populations genetics and functional traits of *Artemisia frigida* in Mongolia. *Journal of Arid*
569 *Environments* 209: 104900.

570 Oyundelger, K., Ritz, C.M., Munkhzul, O., Lang, B., Ahlborn, J., Oyuntsetseg, B., Römermann, C. &
571 Wesche, K. (2020) Climate and land use affect genetic structure of *Stipa glareosa* PA Smirn. in
572 Mongolia. *Flora* 266: 151572.

573 Pan, S., Liu, C., Zhang, W., Xu, S., Wang, N., Li, Y., Gao, J., Wang, Y. & Wang, G. (2013) The scaling
574 relationships between leaf mass and leaf area of vascular plant species change with altitude. *PloS one*
575 8: e76872.

576 Paradis, E. & Schliep, K. (2019) ape 5.0: an environment for modern phylogenetics and evolutionary
577 analyses in R. *Bioinformatics* 35: 526–528.

578 Pellicer, J., Garcia, S., Canela, M.A., Garnatje, T., Korobkov, A.A., Twibell, J.D. & Vallès, J. (2010)
579 Genome size dynamics in *Artemisia* L. (Asteraceae): following the track of polyploidy. *Plant Biology*
580 12: 820–830.

581 Perez-Harguindeguy, N., Diaz, S., Garnier, E., Lavorel, S., Poorter, H., Jaureguiberry, P., Bret-Harte, M.S.,
582 Cornwell, W.K., Craine, J.M. & Gurvich, D.E. (2013) New handbook for standardised measurement
583 of plant functional traits worldwide. *Australian Journal of Botany* 61: 167–234.

584 R Core Team (2020) R: A language and environment for statistical computing.

585 Reisch, C. & Bernhardt-Römermann, M. (2014) The impact of study design and life history traits on genetic
586 variation of plants determined with AFLPs. *Plant Ecology* 215: 1493–1511.

587 Riggins, C.W. & Seigler, D.S. (2012) The genus *Artemisia* (Asteraceae: Anthemideae) at a continental
588 crossroads: Molecular insights into migrations, disjunctions, and reticulations among Old and New
589 World species from a Beringian perspective. *Molecular Phylogenetics and Evolution* 64: 471–490.

590 Rosenberg, N.A. (2004) DISTRUCT: a program for the graphical display of population structure.
591 *Molecular Ecology Notes* 4: 137–138.

592 Sanz, M., Schneeweiss, G., Vilatersana Lluç, R. & Vallès Xirau, J. (2011) Temporal origins and
593 diversification of *Artemisia* and allies (Anthemideae, Asteraceae). *Collectanea Botanica* 30: 7–15.

594 Scheepens, J.F., Frei, E.S. & Stöcklin, J. (2010) Genotypic and environmental variation in specific leaf area
595 in a widespread Alpine plant after transplantation to different altitudes. *Oecologia* 164: 141–150.

596 Stotz, G.C., Salgado-Luarte, C., Escobedo, V.M., Valladares, F. & Gianoli, E. (2022) Phenotypic plasticity
597 and the leaf economics spectrum: plasticity is positively associated with specific leaf area. *Oikos* 2022:
598 e09342.

599 Szczecińska, M., Sramko, G., Wołosz, K. & Sawicki, J. (2016) Genetic diversity and population structure
600 of the rare and endangered plant species *Pulsatilla patens* (L.) Mill in East Central Europe. *PloS one*
601 11: e0151730.

602 Tkach, N. V., Hoffmann, M.H., Röser, M., Korobkov, A.A. & Von Hagen, K.B. (2008) Parallel evolutionary
603 patterns in multiple lineages of arctic *Artemisia* L. (Asteraceae). *Evolution* 62: 184–198.

604 Tuvshintogtokh, I. (2014) *The Steppe Vegetation of Mongolia*. C. Sanchir (ed.). Bembisän, Ulaanbaatar.
605 610 pp.

606 Vallès, J., Garcia, S., Hidalgo, O., Martín, J., Pellicer, J., Sanz, M. & Garnatje, T. (2011) Biology, genome
607 evolution, biotechnological issues and research including applied perspectives in *Artemisia*
608 (Asteraceae). In: J.-C. Kader & M. Delseny (eds.) *Advances in Botanical Research*. Academic Press,
609 pp. 349–419.

610 Vallès, J. & McArthur, E.D. (2001) *Artemisia* systematics and phylogeny: cytogenetic and molecular
611 insights. *USDA Forest Service Proceedings* 21: 67–74.

612 Waitt, D.E. & Levin, D.A. (1998) Genetic and phenotypic correlations in plants: a botanical test of
613 Cheverud's conjecture. *Heredity* 80: 310–319.

614 Wang, Z. (2011) Study on genetic diversity of traditional Mongolian medicine *Artemisia frigida*. Minzu
615 University of China, Master Thesis.

616 Wang, Z., Lv, S., Han, G., Wang, Z., Li, Z., Ren, H., Wang, J., Sun, H. & Zhang, G. (2022) Heavy grazing
617 reduced the spatial heterogeneity of *Artemisia frigida* in desert steppe. *BMC Plant Biology* 22: 337.

618 von Wehrden, H., Hanspach, J., Kaczensky, P., Fischer, J., Wesche, K., von Wehrden, H., Hanspach, J.,

619 Kaczensky, P., Fischer, J. & Wesche, K. (2012) Global assessment of the non-equilibrium concept in
620 rangelands. *Ecological Applications* 22: 393–399.

621 von Wehrden, H., Hanspach, J., Ronnenberg, K. & Wesche, K. (2010) Inter-annual rainfall variability in
622 Central Asia—a contribution to the discussion on the importance of environmental stochasticity in
623 drylands. *Journal of Arid Environments* 74: 1212–1215.

624 von Wehrden, H. & Wesche, K. (2007) Relationships between climate, productivity and vegetation in
625 southern Mongolian drylands. *Basic and Applied Dryland Research* 1: 100.

626 Wei, T., Simko, V.R., Levy, M., Xie, Y., Jin, Y. & Zemla, J. (2021) Package “corrplot”: Visualization of a
627 Correlation Matrix. *Version 0.84*.

628 Wickham, H. (2011) ggplot2. *Wiley Interdisciplinary Reviews: Computational Statistics* 3: 180–185.

629 Woodward, F.I. (1983) The significance of interspecific differences in specific leaf area to the growth of
630 selected herbaceous species from different altitudes. *New Phytologist*: 313–323.

631 Yi, F., Wang, Z., Baskin, C.C., Baskin, J.M., Ye, R., Sun, H., Zhang, Y., Ye, X., Liu, G. & Yang, X. (2019)
632 Seed germination responses to seasonal temperature and drought stress are species-specific but not
633 related to seed size in a desert steppe: Implications for effect of climate change on community
634 structure. *Ecology and Evolution* 9: 2149–2159.

635 Yulin, L.I., Johnson, D.A., Yongzhong, S.U., Jianyuan, C.U.I. & Zhang, T. (2005) Specific leaf area and
636 leaf dry matter content of plants growing in sand dunes. *Botanical Bulletin of Academia Sinica* 46.

637 Zhou, H.-F., Zheng, X.-M., Wei, R.-X., Second, G., Vaughan, D.A. & Ge, S. (2008) Contrasting population
638 genetic structure and gene flow between *Oryza rufipogon* and *Oryza nivara*. *Theoretical and Applied*
639 *Genetics* 117: 1181–1189.

640