## Crosstalk Between Sodium-Glucose Cotransporter Inhibitors and Sodium-Hydrogen Exchanger- 1 and 3 in Cardiometabolic Diseases

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#### Abstract

The hallmark of type 2 diabetes mellitus (T2DM) is abnormal glucose homeostasis due to hyperglycaemia or insulin resistance. Metabolic abnormalities in T2DM lead to cellular dysfunction and the development of diabetic cardiomyopathy and heart failure. New antihyperglycemic agents, such as glucagon-like peptide-1 receptor agonists and the sodium-glucose cotransporter-2 inhibitors (SGLT2i) have shown to attenuate endothelial dysfunction at the cellular level. In addition, they showed cardiovascular safety and cardioprotective effects. How these drugs exert their cardioprotective effects is unknown, although recent studies show that cardiovascular homeostasis occurs through the interplay of the sodium hydrogen exchangers (NHE), specifically NHE1 and NHE3 with SGLT2i. Another theoretical explanation for the SGLT2i cardioprotective effects is through natriuresis by the kidney. This theory highlights the possible involvement of renal NHE transporters in the management of heart failure. This review outlines possible mechanisms predisposing to diabetic cardiomyopathy and discusses the interaction between NHE and SGLT2i in cardiovascular disease.







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24 The hallmark of type 2 diabetes mellitus (T2DM) is abnormal glucose homeostasis due to hyperglycaemia or insulin resistance. Metabolic abnormalities in T2DM lead to cellular 25 26 dysfunction and the development of diabetic cardiomyopathy and heart failure. New 27 antihyperglycemic agents, such as glucagon-like peptide-1 receptor agonists and the sodium-28 glucose cotransporter-2 inhibitors (SGLT2i) have shown to attenuate endothelial dysfunction 29 at the cellular level. In addition, they showed cardiovascular safety and cardioprotective 30 effects. How these drugs exert their cardioprotective effects is unknown, although recent 31 studies show that cardiovascular homeostasis occurs through the interplay of the sodium 32 hydrogen exchangers (NHE), specifically NHE1 and NHE3 with SGLT2i. Another 33 theoretical explanation for the SGLT2i cardioprotective effects is through natriuresis by the 34 kidney. This theory highlights the possible involvement of renal NHE transporters in the 35 management of heart failure. This review outlines possible mechanisms predisposing to 36 diabetic cardiomyopathy and discusses the interaction between NHE and SGLT2i in

37 cardiovascular disease.

#### 38 1. Introduction

39 Diabetes Mellitus (DM) is a metabolic disorder where the body either does not produce 40 sufficient amount of insulin, has impaired insulin action or a combination of the two. Type 1 41 (T1DM) which constitutes ~5-10% of diabetes cases has high incidence in children and 42 adolescents and is caused by immune destruction of the  $\beta$ -islets in the pancreas. The majority 43 of cases are Type 2 (T2DM), which results from a combination of beta cell dysfunction and 44 insulin resistance. Insulin resistance and the accompanied hyperinsulinemia are the early 45 detected metabolic abnormality in subjects destined to develop T2DM and precedes the 46 deterioration in glucose homeostasis (American Diabetes, 2017; Kahanovitz et al., 2017). 47 T2DM affects approximately 463 million people worldwide and future estimates suggest that 48 102 of 1000 people will be diagnosed with diabetes by 2030 (Saeedi et al., 2019). Chronic 49 diabetes without appropriate treatment causes microvascular and macrovascular 50 complications like nephropathy, retinopathy, neuropathy, and atherosclerotic cardiovascular 51 diseases (CVDs).

52 CVDs are considered the most common cause of morbidity and mortality in diabetic 53 patients. In the US, CVD death rates are 1.7 times higher among adults with DM than those 54 without, which is attributable to the increased risk of stroke, myocardial infarction (MI), and 55 heart failure (HF) (Leon & Maddox, 2015). Patients with T2DM have a two-to-five-folds 56 increased risk of HF, independent of other risk factors like hypertension, coronary artery 57 disease, and dyslipidaemia (Martín-Timón et al., 2014) (Nichols et al., 2004). A rise in 58 glycated haemoglobin by 1% has been associated with an 8% increase in CVD risk (Stratton 59 et al., 2000). Furthermore, the presence of T2DM worsens the prognosis of heart failure. In 60 addition, T1DM patients have a 30% risk of HF with every 1% increase in glycated 61 haemoglobin (Zhao et al., 2014). 62 Hyperglycaemia and insulin resistance are the major etiological factors promoting

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63 cardiomyopathy and HF in diabetic patients (Jia et al., 2016). Ultimately, the progression of
64 HF in DM is linked to pathological changes to the heart muscle and coronary vasculature,
65 which eventually lead to diabetic cardiomyopathy (DCM) (Jia et al., 2018).

66 Recently discovered antihyperglycemic agents, such as glucagon-like peptide-1 receptor

agonists, and the sodium-glucose cotransporter-2 (SGLT2) inhibitors (SGLT2i) have shown

68 cardioprotective effects. SGLT2i were shown to decrease the rates of HF and hospitalization

69 from HF in several clinical trials (Ali et al., 2019). How these new antihyperglycemic drugs

70 exert cardioprotection is unknown, although some studies show involvement of the sodium

71 hydrogen exchanger (NHE) and sodium-glucose transporter (SGLT) families. This review

72 discusses some of the mechanisms predisposing to diabetic cardiomyopathy and highlights

73 the role of NHE and SGLT transporters in cardiovascular disease.

#### 74 2. Pathophysiology of Diabetic Cardiomyopathy

DCM is recognized by defects in the structure and performance of the myocardium in individuals with diabetes, independent of other cardiac risk factors. The structural abnormalities in DCM progress through three stages, an early stage characterized by diastolic dysfunction, which gradually develops to systolic dysfunction in the advanced stage, and eventually to HF in the late stage. (Jia et al., 2016).

80 Early-stage DCM, mainly caused by hyperglycaemia and insulin resistance, presents with 81 impairment in the left ventricle (LV) diastolic filling, compensated by increased LV and atrial 82 filling pressure and left atrial enlargement (Seferović & Paulus, 2015). Hyperglycaemia leads 83 to downregulation of GLUT4, impaired glycolysis, and increased free FA levels (from 84 impaired FA metabolism). Insulin resistance results in increased lipolysis and elevated 85 plasma FFA concentration as well leading to increased influx to myocytes and development of cardiac steatosis. The events result in high levels of ROS, impaired Ca<sup>2+</sup> homeostasis, 86 mitochondrial dysfunction, endoplasmic reticulum stress, oxidative stress, and activation of 87 88 the sympathetic nervous system, all of which promote cardiac hypertrophy, fibrosis, and 89 cardiomyocyte apoptosis (Jia et al., 2016). The advanced stage comprises continued cardiac 90 injury and further stimulates the Renin-Angiotensin-Aldosterone System (RAAS) and 91 maladaptive immune responses that culminate in impaired autophagy of cells (Jia et al., 92 2016). Advanced stage features include LV hypertrophy and cardiac remodelling, with 93 impaired cardiac diastolic function. Consequently, the individual may develop HF with a 94 normal ejection fraction. In late-stage DCM, neurohumoral activation, impaired metabolism, 95 and myocardial fibrosis weaken coronary microcirculation and the diastolic and systolic 96 functions of the heart (Aronow & Ahn, 1999; Jia et al., 2018). Additionally, impaired insulin 97 signalling and oxidative stress both decrease levels of the vasodilator nitric oxide (NO) (Jia et 98 al., 2016).

#### 99 2.1 The Role of Endothelial Dysfunction in the Development of DCM

100 Studies on DCM development using animal models have implicated multiple 101 pathophysiologic mechanisms, such as mitochondrial dysfunction, RAAS activation, Ca<sup>2+</sup> 102 homeostasis impairment, lipotoxicity, myocardial steatosis, glucose toxicity, and most 103 recently, endothelial dysfunction. Hyperglycaemia is one of the main factors triggering 104 endothelial dysfunction by exerting several biochemical changes that damage cardiac and 105 vascular endothelial cells. Some of these changes trigger ROS production and induce 106 oxidative stress levels that overwhelm cells, enhance non-enzymatic glycation, activate 107 protein kinase-C (PKC), and ameliorate the cells' redox potential (Avogaro et al., 2011). 108 Oxidative stress promotes the formation and deposition of AGE products creating elevated 109 interstitial collagen deposition and increased myocardial wall stiffness. If untreated, all of 110 these DCM-related structural changes would result in HF (Jia et al., 2016). 111 The endothelium is a single-layer cellular lining of the whole vascular system. Endothelial 112 cells have unique functions vital for cardiovascular homeostasis. For example, the 113 endothelium functions as a semi-permeable barrier between blood and body tissues. The 114 endothelium also controls vascular tone by secreting the vasodilators nitric oxide, 115 prostacyclin, and endothelium-derived hyperpolarizing factors, as well as producing 116 vasoconstrictors like endothelin-1 and thromboxane-A<sub>2</sub>. Endothelial dysfunction, 117 characterized by low nitric oxide bioavailability, occurs when endothelial cells lose their 118 barrier property and fail to balance vascular dilatory and constrictive tone, coagulation, and 119 anticoagulation. T1DM and T2DM patients show decreased vasorelaxation by NO (Avogaro 120 et al., 2011; Endemann & Schiffrin, 2004). Reduced NO production is observed in diabetic 121 experimental models (Hink et al., 2001; Shi & Vanhoutte, 2009), and in-vitro studies with 122 endothelial cells have shown that high glucose levels lead to less NO production (De Vriese 123 et al., 2000). Endothelial dysfunction is considered the first step in developing atherosclerotic 124 complications in metabolic conditions such as diabetes, pre-diabetes, and obesity (Avogaro et al., 2006). 125 126 Several mechanisms contribute to lower NO bioavailability during endothelial

dysfunction. Production of ROS through NADPH oxidase, an electron transport chain
 protein, leads to oxidative stress. ROS reacts with NO to produce a cytotoxic oxidant
 compound called peroxynitrite. Peroxynitrite increases oxidative stress even further, which in
 turn lowers NO production through uncoupling of NO synthases (NOS) and mediates low density lipoprotein oxidation. Peroxynitrite also leads to protein dysfunction via nitration of

132 proteins. In insulin resistance, the PI3K/Akt pathway involved in NOS activation is inhibited,

- 133 while endothelin-1 and adhesion molecule production pathways remain intact. Also, the
- 134 presence of AGE products contributes to oxidative stress and leads to endothelial
- 135 dysfunction. Endoplasmic reticulum stress, a pro-apoptotic pathway, is another mechanism
- 136 where the pro-survival unfolded protein response becomes chronically activated (Aviello &
- 137 Knaus, 2018; Avogaro et al., 2011; Endemann & Schiffrin, 2004; Maamoun et al., 2019;
- 138 Muniyappa & Sowers, 2013).

139 Antihyperglycemic medications that target and attenuate endothelial dysfunction such 140 as liraglutide, metformin, pioglitazone, and SGLT2i (empagliflozin; EMPA, canagliflozin, 141 CANA; dapagliflozin, DAPA) are becoming of great interest. (Batzias et al., 2018; Eriksson 142 & Nyström, 2015). In porcine coronary artery cultured endothelial cells, high glucose increased endothelial dysfunction markers, oxidative stress, and VCAM-1, and reduced NOS 143 144 expression. Treatment with SGLTi exerted a protective effect and prevented endothelial 145 dysfunction (Khemais-Benkhiat et al., 2020). Additionally, in the obese ZSF1 rat model, 146 systolic blood pressure (BP) was higher than the lean control group, NOS was 147 downregulated, and expression of the adhesion molecule VCAM-1 was increased. Chronic 148 treatment of T2DM ZDF rats with empagliflozin (EMPA) prevented oxidative stress, 149 signalling and inflammation, AGE products formation, and attenuated endothelial dysfunction (Steven et al., 2017). In Apo-E<sup>-/-</sup> streptozotocin (STZ)-induced diabetic mice, 150 151 treatment with EMPA also attenuated endothelial dysfunction and reduced atherogenesis 152 (Ganbaatar et al., 2019). The EMBLEM trial included 117 patients with T2DM and 153 concurrent CVDs, randomized into a 1:1 ratio to receive either placebo or EMPA over 24 154 weeks (Tanaka et al., 2019). The primary endpoint was the change in reactive hyperaemia 155 index, an endothelial dysfunction marker, from baseline. Per-protocol analysis did not show 156 an improvement in endothelial dysfunction. However, the study was limited by the small 157 number of patients and unrepresentable population. The mean population systolic BP was 158 130 mmHg and the BMI 26.4 kg m<sup>-2</sup>, which are lower than expected values for diabetic 159 patients with concurrent cardiovascular disease. In T2DM mice, treatment with dapagliflozin 160 (DAPA) attenuated endothelial dysfunction, vascular smooth muscle dysfunction, and arterial 161 stiffness (Lee et al., 2018). Gaspari et al. (2018) showed that DAPA attenuated TNFa- and 162 hyperglycaemia-induced endothelial dysfunction in vitro with a human endothelial cell line. While *in-vivo*, both adult and aged ApoE<sup>-/-</sup> mice chronically administered with DAPA 163 164 showed attenuated endothelial dysfunction and less vascular adhesion molecules.

#### 165 **2.2** The Role of Metabolic Disturbances in the Development of DCM

166 Myocardial cells are characterized by their metabolic flexibility, which is the ability to 167 utilize several substrates such as glucose, lactate, and fatty acids (FAs) to generate ATP 168 molecules. In a healthy heart, there is a constant supply of ATP by oxidative phosphorylation 169 of FAs in the mitochondria (60 - 90%), while maintaining a balance in using other substrates 170 like glucose and lactate (Stanley & Chandler, 2002). 171 In diseases such as HF and DM, the metabolic balance is impaired. The failing heart 172 increases the utilization of glucose over FAs to increase energy production. However, in 173 diabetic heart, there may be a metabolic shift toward FA oxidation rather than glucose 174 oxidation. This shift is thought to be due to the chronic hyperglycaemia, insulin deficiency, 175 and insulin resistance. The enhanced FA oxidation observed in a diabetic heart might exceed 176 cardiac utilization capacity and predispose the heart to triacylglycerols (TAGs) and ceramides 177 disposition, which in turn contributes to cardiac hypertrophy and stenosis. Along with the 178 burden created by advanced glycated end products (AGEs), the cardiac metabolic changes 179 promote collagen deposition and induce myocardial fibrosis leading to the damage of the 180 cardiomyocytes present in DCM (Fuentes-Antrás et al., 2015). 181 Furthermore, in cardiac diseases, ischemia and hypoxia promote a shift to anaerobic 182 respiration. The activity of adenosine monophosphate kinase (AMPK), an energy balancing 183 enzyme that promotes anaerobic ATP production, is allosterically regulated by the ratio of 184 AMP to ATP. When ATP is abundant, it binds to AMPK and inactivates it. Therefore, during 185 pathological low energy states when AMP is abundant, AMPK is activated to provide the heart with ATP. In addition to energy production, AMPK activation protects cells against 186 187 myocardial injury during ischemia, reduces reactive oxygen species (ROS), and attenuates 188 endoplasmic reticulum stress Additionally, sodium (Na<sup>+</sup>) overload, a characteristic of HF, increases calcium ( $Ca^{2+}$ ) efflux which interferes with the Krebs cycle, that is adding up to the 189 190 metabolic disturbances (Qi & Young, 2015).

#### 191 **3.** Characteristics of NHE & SGLT membrane transporters

#### **3.1 NHE Overview**

193 The sodium hydrogen exchangers (NHE) family of integral membrane protein antiporters 194 consists of 10 isoforms that function by exchanging sodium cations with protons through cell 195 membranes (Packer, 2017). NHE1 and NHE3 are two well-studied isoforms involved in renal and cardiovascular homeostasis. NHE1, ubiquitous in mammalian cells, is the dominant
isoform in the heart where it regulates intracellular pH, cell volume, and proliferation and
shows the highest expression in the kidney (Das et al., 1987; Packer, 2017; Parker et al.,
2015). NHE3 contributes to regulating extracellular volume and BP by reabsorption of Na<sup>+</sup> in

200 the kidney (Dominguez Rieg et al., 2016; Packer, 2017).

#### 201 **3.1.1** Activity and Regulation of NHE1 and NHE3

202 NHE1 influxes Na<sup>+</sup> in response to intracellular acidification, where the protein exhibits 203 an allosteric binding site for protons. Regulation of NHE1 can also occur in response to 204 different membrane receptors that can exert conformational changes or C-terminal 205 phosphorylation. Extracellular and hormonal pathways, such as angiotensin II (ANG-II), 206 endothelin-I, and thrombin, control the activity of NHE1 regulators. Receptor regulators of 207 NHE1 include protein kinases, G-coupled receptors, and integrin receptors (Vallés et al., 208 2015). Tyrosine kinase activation increases NHE1 activity through the Ras-mediated ERK 209 cascade, including Ras downstream effectors such as MEK1/2, Raf-1, and ERK. However, 210 the serine/threonine kinase ERK downstream effector called p90 ribosomal S6 kinase (p90<sup>RSK</sup>) directly phosphorylates NHE1 instead of ERK (Putney et al., 2002). The RSK 211 family of transporters includes four isoforms (RSK1-4). RSK1, also designated as p90<sup>RSK</sup>, 212 sustains regular cardiac function, making this enzyme essential (Lara et al., 2013). p90<sup>RSK</sup> 213 hyperactivity induces cardiac hypertrophy and HF. In neonates, p90<sup>RSK</sup> activation increases c-214 215 Fos and Egr-1 expression in ventricular myocytes to promote myocytes' development (Takahashi et al., 1999). Furthermore,  $p90^{RSK}$  activation reduces glycogen synthase kinase-3 $\beta$ 216 217 (GSK-3β) activity in mice with defective ryanodine receptor ion channels leading to cardiac hypertrophy progression (Q. He et al., 2010). A study by Takeishi et al. (1999) found aberrant 218 activation of p90<sup>RSK</sup> in guinea pig pressure-overload-induced hypertrophic myocardium. 219 Moreover, patients with dilated cardiomyopathy had higher levels of activated p90<sup>RSK</sup> than 220 their healthy peers (Horie et al., 1992; Javadov et al., 2009; Muthusamy et al., 2013; Takeishi 221 222 et al., 2002; Yamaguchi et al., 2011). These findings highlight the role of p90<sup>RSK</sup> in inducing 223 cardiac dysfunction, remodelling, and its role in NHE1 activation. 224 Akt is another kinase known to regulate NHE1 activity. The duration of Akt activation is the 225 determinant of its effect (Takeishi et al., 2002). Short-term Akt activation promotes 226 physiological hypertrophy during postnatal cardiac development characterized by normal or

227 enhanced contractile function (Walsh, 2006), while contractile dysfunction characterizes

228 long-term Akt activation (Shiojima et al., 2002). A study done on mouse embryo fibroblasts

- showed that Akt inhibition reduced NHE1 activity by blocking the translocation of NHE1 to
- 230 the cell membrane. Furthermore, upstreaming of Akt enhances p90<sup>RSK</sup> activation and thus

231 plays a role in cardiomyopathy (Clement et al., 2013; Kemi et al., 2008).

232 The role of cardiac AMPK, one of the NHE channel regulatory kinases, in cardiac

233 metabolism is not known. However, studies suggest that activating AMPK by

234 phosphorylation triggers the trafficking of glucose transporters (GLUT1 and GLUT4) to the

sarcolemma and increases glucose uptake (Qi & Young, 2015; Rotte et al., 2010).

236 The G protein-coupled receptor subunits Gaq and Ga13 also activate NHE1. Ga13 activates

237 NHE1 through the GTPase RhoA pathway, while Gaq activates NHE1 through the PKC-

238 dependent mechanism. The suppression of PKC in several Ga<sub>q</sub> protein-coupled receptors,

239 namely α1-adrenergic, vasopressin, and endothelin-1, impairs NHE1 activation. However, in

some Ca<sup>2+</sup> mobilizing Ga<sub>q</sub> coupled receptors, NHE1 activation can occur independently of

241 PKC. Moreover, integrin receptors can activate NHE1, which may be due to the shared

signalling pathway with Ga<sub>13</sub> that activates NHE1 (Avkiran & Haworth, 2003; Kitamura et
al., 1995).

Other than receptor-mediated regulation, NHE1 regulation occurs through the direct binding of regulatory proteins to the C-terminal. Accessory proteins, which take part in the regulation of cardiac NHE1 activity were also investigated, such as carbonic anhydrase-II (CAII), Ca<sup>2+</sup>-binding proteins (calmodulin and calcineurin B homologous proteins [CHPs]), and phospholipids. Cellular Na<sup>+</sup> concentration, regulated by NHE1, is instrumental for function, playing roles in Ca<sup>2+</sup> regulation, metabolism, contractility, and heart stability (Lambert et al., 2015).

251 Several physiological and hormonal modulators regulate NHE3 activity. The majority of 252 the NHE3 regulatory hormones are coupled to protein kinases associated with intracellular 253 signalling cascades. Different mechanisms such as direct phosphorylation, protein trafficking, 254 and interaction with accessory proteins modulate NHE3 activity (Dynia et al., 2010; Pedersen 255 & Counillon, 2019). Moreover, in a normal state, the regulation of NHE3 is dependent on its 256 C-terminal phosphorylation. Various kinases, including casein kinase 2 (CK2), serum 257 glucocorticoid-regulated kinase-1 (SGK1), protein kinase A (PKA), Ca<sup>2+</sup>/Calmodulindependent Protein Kinase-II (CaMKII), cGKII, GSK-3, AKT, ERK and p90<sup>RSK</sup> mediate 258 259 NHE3 phosphorylation (Dynia et al., 2010). No et al. (2015) demonstrated that 260 lysophosphatidic acid (LPA) stimulated NHE3 activity by LPA5 receptor and EGF receptor

261 (EGFR) transactivity. This, in turn, activated proline-rich tyrosine kinase 2 (Pyk2) and ERK

- specifically in the apical membrane. The authors hypothesized that RSK could be an
- associated effector of Pyk2 and ERK since RSK is a well-known effector of EGFR and ERK.
- In contrast, the regulation of RSK by Pyk2 is still not known. The study showed that RSK2,
- but not RSK1, regulated direct phosphorylation of NHE3 and concluded that RSK2
- 266 phosphorylation of NHE3 mediates NHE3 regulation by LPA.

#### 267 3.2 SGLT Receptors Overview

Sodium-glucose co-transporters (SGLTs) are active symporters that belong to the solutecarrier family-5 (SLC5) of active glucose transportation, and facilitate glucose homeostasis (Wright et al., 2011). The human SLC5 transporter family contains 12 members, with up to six different SGLT receptors identified in human cells. Functional studies showed that all SLC5 family proteins weigh between 60- to 80-kDa (580–718 amino acids). The moststudied isoforms of this family, SGLT1 and SGLT2, are involved in glucose absorption and glucosuria.

275

#### 3.2.1 Activity and Regulation of SGLT

276 Several studies have focused on the activity and expression of SGLT under different 277 physiological/pathophysiological settings. SGLT1 expression was in the small intestine, 278 kidneys, liver, lungs, cardiac myocytes, and highly expressed in the human heart. SGLT2 279 expression was primarily found in the kidney and pancreatic alpha cells. (Kashiwagi et al., 280 2015). SGLT1 levels are elevated further in cardiac ischemia or hypertrophy disease states. 281 This increase in SGLT1 expression can be linked to the increased phosphorylation of 282 secondary messengers such as ERK 1/2 and the mammalian target of rapamycin (mTOR), 283 involved in the signaling pathways of cardiac ischemia/hypertrophy. However, further studies 284 are required to confirm the proposed mechanism (Di Franco et al., 2017). 285 The kidney plays a vital role in glucose homeostasis by promoting the reabsorption of 286 filtered glucose. The two isoforms carry out reabsorption across the apical cell membranes 287 (Poulsen et al., 2015). SGLT2 is located on the luminal membrane of the proximal 288 convoluted tubule in S1 and S2 segments, whereas SGLT1 is expressed in the S3 segment 289 (Figure 1) (Chao, 2014). A healthy kidney reabsorbs 90% of filtered glucose from the 290 proximal tubule via SGLT2, whereas a diabetic kidney increases its reabsorption of glucose 291 by 20% more than the normal rate through the overexpression of SGLT2. The active

transport of glucose by both isoforms is linked with the transport of Na<sup>+</sup> into the intracellular

- 293 fluid (Chao, 2014; Novikov & Vallon, 2016). Inhibition of this process promotes the
- reduction of intracellular Na<sup>+</sup> levels and excretion of glucose in urine (glucosuria), leading to
- the correction of hyperglycaemia (Chao, 2014).

#### 296 4. Role of SGLT & NHE1 and 3 in Diabetes

297 DM can stimulate the proliferation of vascular smooth muscle cells (VSMCs) to 298 proliferate through insulin and insulin-like growth factor 1 (IGF-1), which is in turn mediated 299 by NHE1. Insulin can stimulate the transcription of NHE1 directly and regulate the activity of 300 NHE1 in nonvascular cells, while IGF-1 regulates NHE1 activity in vascular cells. Moreover, 301 hyperglycaemia affects the activity of NHE1. For example, hyperglycaemia increases the 302 production of diacylglycerol precursors, leading to the PKC activation, consequently 303 activating NHE1. Also, NHE1 in VSMCs can be activated by the oxidized LDL which has 304 been shown to be elevated in DM and hyperlipidaemia. Furthermore, AGEs react with the 305 extracellular matrix, resulting in the thickening of vessel walls. Besides, VSMCs adhesion, 306 which is mediated by cell surface integrins and extracellular matrix proteins, promotes PKC 307 activation and stimulation of NHE1 activity. Interestingly, it was speculated that glycation of 308 the extracellular matrix protein fibronectin inhibited NHE activity and suppressed the growth 309 of VSMCs (Hannan & Little, 1998).

On the other hand, the activity of NHE3 is stimulated as a result of increased levels of insulin, glucose, and specific adipokines in T2DM. The increased activity and upregulation of NHE3 may be instrumental to developing chronic complications in diabetic patients such as: diabetic nephropathy and uric acid nephrolithiasis (Packer, 2017). The early phase of diabetic kidney disease presents changes in eGFR, elevated reabsorption of salt and water, and expanded extracellular volume, all of which advance to hypertension, hyperfiltration, and eventually renal hypertrophy (Girardi & Sole, 2012).

317 The number of main Na<sup>+</sup> and water transporters are hypothesized to increase in diabetic 318 kidneys as a compensatory mechanism due to extensive water and Na<sup>+</sup> loss (Song et al., 319 2003). The study demonstrated that STZ- induced T1DM rats had an increased protein 320 content of Na<sup>+</sup> and water transporters NHE3 (204% of the vehicle mean), thiazide-sensitive 321 Na<sup>+</sup>/Cl<sup>-</sup> co-transporter, and  $\alpha$ ,  $\beta$ , and  $\gamma$  subunits of the epithelial sodium channel. According 322 to another study conducted by Klisic et al. (2006) using a similar animal model, brush-border 323 membrane NHE3 activity was significantly higher by 40% after seven days and 37% after 14 324 days compared to control rats. However, the increased activity of NHE3 was not associated

325 with changes in NHE3 protein or mRNA. Unlike Song et al. (2003), they selectively used 326 cortical brush-border membrane vesicle for analysis to reflect proximal tubule NHE3 and not 327 the analysis of whole kidney homogenates. An STZ-induced diabetic rat study demonstrated 328 that diabetic kidneys were 67% larger in size, had 22% longer proximal tubules, and 20% 329 longer distal tubules compared to normal rat kidneys Rasch (1984). Since the reabsorption of 330 Na<sup>+</sup> occurs mainly in the proximal tubules, its elongation can easily result in increased 331 activity of NHE3 (Girardi & Sole, 2012). Hyperglycaemia also enhances ANG-II production 332 by stimulation of angiotensinogen and RAAS. This further activates NHE3 via the SGK1 333 signalling cascade involving phosphatidylinositol 3-kinase (PI3-kinase) and 3-334 phosphoinositide-dependent protein kinase-1 (PDK1) (Ackermann et al., 2009; Stevens et al., 335 2008). Another signal cascade of ANG-II induced NHE3 stimulation includes the nonreceptor tyrosine kinase (c- Src), PI3-kinase activation, PKC (du Cheyron et al., 2003; 336 Tsuganezawa et al., 1998), and Ca<sup>2+</sup> and CaMKII (P. He et al., 2010). In the proximal tubule, 337 338 the uptake of albumin requires the involvement of the megalin/cubilin complex. In diabetic 339 nephropathy, there is decreased endocytosis of albumin due to decreased megalin expression, 340 characterized by microalbuminuria (Tojo et al., 2001). The decreased albumin uptake leads to 341 elevated intratubular albumin concentration, stimulating NHE3 activity and further worsening 342 kidney damage (Girardi & Sole, 2012). In the Opossum kidney cells, high glucose levels 343 resulted in hypertrophy due to increased osmolality (Drumm et al., 2003). Consequently, albumin uptake increased because of NHE3 overactivity. 344

#### 345 5. Role of SGLT & NHE1 and 3 in Cardiovascular Diseases

#### 346 5.1 Ischemia-Reperfusion Injury, Cardiac Remodelling, and Hypertrophy

347 HF is a syndrome often developed after several remodelling processes in the heart that 348 includes LV hypertrophy, fibrosis, and diastolic dysfunction (Uthman et al., 2018). In 349 diabetes, the heart is in a state of metabolic overload due to cardiac metabolism. Several vital 350 mechanisms were linked to the induction of cardiac impairment and the early development of 351 HF that overlap with other CVDs. NHE and SGLT's potential relevance to the direct effects 352 in the myocardium will be discussed concerning the early stages of HF development. 353 As NHE 1 is the main plasma membrane isoform in the heart, it takes an essential part in 354 cardiac functioning in normal and disease states. Hormones such as endothelin-1, ANG-II, 355 and α-adrenergic stimulators, contribute to NHE1 activity in cardiac remodelling (Odunewu-356 Aderibigbe & Fliegel, 2014; Wakabayashi et al., 2013). Overactivity of NHE1 has been

357 proven to cause several pathological changes in the myocardium, including ischemia-358 reperfusion injury (IRI), cardiac remodelling, hypertrophy, and apoptosis that eventually can 359 progress to HF, The potential mechanisms underlying the role of NHE1 in the remodelling 360 process can be summarized by the role of both Na<sup>+</sup> accumulation and mitochondrial 361 remodelling (Karmazyn et al., 2008). During the disease state, as an adaptive mechanism, 362 NHE1 activity is increased to correct the reduced intracellular pH. Since the  $Na^+/K^+$  ATPase 363 becomes inactive during ischemia, NHE-mediated Na<sup>+</sup> influx leads to the intracellular 364 accumulation of Na<sup>+</sup> (Cingolani Horacio & Ennis Irene, 2007; Imahashi et al., 2007; 365 Karmazyn et al., 2008). This rise in intracellular Na<sup>+</sup> consequently leads to the two-folds elevation in intracellular Ca<sup>2+</sup> by direct reversal of Na<sup>+</sup>/Ca<sup>2+</sup> exchanger (Figure 2) 366 (Wakabayashi et al., 2013), leading to an intracellular Ca<sup>2+</sup> overload which in turn triggers 367 deleterious pathways that lead to myocardial injury, hypertrophy, and subsequent dysfunction 368 369 (Figure 3). Additionally, impairment of mitochondrial function and structure due to swelling, 370 ATP depletion/dysfunction, ROS production, and opening of the mitochondrial permeability 371 transition pore (MPTP) often accompanies cardiac hypertrophy. On the other hand, NHE1 372 inhibition and gene ablation attenuates the opening of MPTP and balances the amounts 373 fission and fusion proteins on the mitochondria. Hence, NHE1 inhibition serves as a cardioprotective mechanism to prevent Na<sup>+</sup> and Ca<sup>2+</sup> accumulation and subsequent activation 374 375 of intracellular pathways, which in turn may improve mitochondrial function and structure 376 integrity, and the cumulative adverse effects on the myocardium (Odunewu-Aderibigbe & 377 Fliegel, 2014). Various studies have shown that inhibition and genetic ablation of NHE1 of 378 in-vivo models protected the myocardium from ischemia-reperfusion injury (Wang et al., 379 2003). In another study, although transgenic mice models overexpressing NHE1 had no 380 significant effect on cardiac function, intracellular pH, intracellular Na<sup>+</sup>, and ischemia-381 reperfusion injury, NHE1 inhibition with cariporide prior to the development of ischemia prevented accumulation of Na<sup>+</sup> and Ca<sup>2+</sup> and decreased ischemia-reperfusion injury, showing 382 383 that baseline NHE1 activity was not the rate-limiting step (Imahashi et al., 2007). 384 The effect of SGLT on diabetic hearts has been well researched within the last few 385 years. Although most of the literature reported on the impact of SGLT2i preventing CVD in

T2DM, SGLT2 receptors were not detected in the heart. New data confirmed that SGLT1
expression and activity are upregulated in ischemia, hypertrophic, failing, and diabetic hearts
in humans with end-stage cardiomyopathy and animal models (García-Ropero et al., 2019;
Uthman et al., 2018). Ramratnam et al. (2014) reported that overexpression of SGLT1 in
transgenic mice was associated with pathologic cardiac hypertrophy and LV dysfunction.

391 During ischemia, glucose uptake and utilization increase along with a 2-to-3-fold

392 upregulation of SGLT1. This upregulation was postulated to be an adaptive response to injury

393 and as a response to AMPK and ERK1/2 activation (García-Ropero et al., 2019). How

394 SGLT1 up-regulation makes an impact is not known, and discrepancies between studies leave

- 395 it uncertain whether SGLT1 receptors exert a protective or deleterious role in cardiac
- 396 physiology.

397 During acute injuries, SGLT1 over-expression facilitates glucose uptake and 398 generates ATP molecules for the heart through anaerobic glycolysis. Kashiwagi et al. (2015) 399 provided evidence of the protective role of SGLT1 against IRI. Using the ex-vivo murine 400 langendorff model, they studied the role of SGLT1 inhibition by phlorizin on cardiac 401 function. During IRI, the use of phlorizin resulted in significant impairment in the recovery of 402 LV contractions and increased infarct size (due to increased CPK activity). There was also a 403 reduction in ATP content associated with a decrease in glucose uptake and glycolysis, 404 showing that SGLT1 inhibition during ischemia-reperfusion impairs cardiac metabolism. 405 On the other hand, several studies showed that SGLT1 inhibition leads to the improvement of 406 cardiomyopathy. This is evidenced by several experimental studies in SGLT1 knockdown 407 models. Ramratnam et al. (2014) reported that double-transgenic mice (SGLT1 knockdown 408 with PRKAG2 mutation) have attenuated cardiac glycogen accumulation, cardiac 409 hypertrophy, and LV dysfunction. Similarly, Z. Li et al. (2019) discovered that 410 pharmacological and genetic inhibition of SGLT1 prevented injuries following ischemia-411 reperfusion (in in-vivo, ex-vivo, and in-vitro models) and reduced ROS, myocardial necrosis, 412 infarct size, along with improved hemodynamic functions. Furthermore, an in-vitro study 413 (Balteau et al., 2011) on adult rat myocytes demonstrated that increased glucose transport 414 through SGLT1 results in NADPH oxidase (NOX2) activation, leading to increased 415 production of ROS and subsequent damage to cardiomyocytes (Figure 3). This effect was 416 counteracted by phlorizin, an SGLT1 inhibitor. 417 In contrast to SGLT1, SGLT2 receptors were not detected in the heart. The

418 mechanism of SGLT2i cardioprotection is still undetermined, but studies have shown that

419 SGLT2i affect cardiomyocytes by directly inhibiting NHE and improving mitochondrial

420 function. Future studies should investigate if there is a link between SGLT2i and SGLT1 in

421 failing hearts and whether dual inhibition may have other beneficial effects on the

422 myocardium. However, Lee et al. (2021) induced MI through left anterior descending artery

- 423 ligation in non-diabetic mice. Three-days after MI induction, there was a transient expression
- 424 of SGLT2 in the site of occlusion in the heart showed by immunofluorescence and western

blot. However, the authors could not conclude if SGLT2 is expressed by cardiomyocytes orby the inflammatory cells migrating to the infarct site.

427 Overall, there is an interplay between the two membrane transporters (SGLT1 and NHE1) 428 in mediating cardiac effects in failing hearts. While the role of NHE1 inhibition is well 429 defined, the cardioprotective mechanism of SGLT2 inhibition is unknown with the exemption 430 of SGLT2i direct NHE1 inhibition. Interestingly, a preclinical in-vitro study in 431 lipopolysaccharide (LPS) stimulated mouse cardio-fibroblasts tested the hypothesis that 432 DAPA can cause NHE1 downregulation AMPK- dependent pathway. Ye et al. (2018) 433 reported that DAPA resulted in elevated levels of the phosphorylated form of AMPK in LPS 434 stimulated cardio-fibroblasts. The results showed that DAPA mitigated the rise in NHE1 435 mRNA and confirmed the relation between NHE1 and Hap70 through the AMPK dependent 436 pathway. Similarly, Uthman et al. (2018), proved that the three SGLT2i available in the

437 market directly suppressed NHE1 activity in-vitro.

#### 438 **5.2 Diabetic Cardiomyopathy**

439 A plethora of evidence suggests that NHE1 is noticeably involved in mediating cardiac 440 hypertrophic responses in DCM, and therefore a potential therapeutic target (Karmazyn, 441 2003). Mraiche et al. (2011) used two transgenic mouse models; one expressing wild type 442 NHE1 and another expressing an activated form to investigate the effect of NHE1 activation 443 on cardiac hypertrophy. NHE1 hyperactivation has been linked to elevated glucose levels in 444 DCM induced by PKC-dependent mechanisms. Additionally, an increase in heart weight to 445 body weight, apoptosis, fibrosis, and a decrease in cardiac functionality was recorded. 446 Studies had shown an enhanced mitochondrial NHE1 activity in the hearts of diabetic rats. 447 Allen and Xiao (2003) have illustrated that the main pathway for Na<sup>+</sup> entry during 448 reperfusion of an ischemic diabetic heart is NHE1. Na<sup>+</sup> concentration changes are linked to 449 altered Ca<sup>2+</sup> influx, production of ROS, and cell damage. Additionally, expression of the 450 activated form of NHE1 increased the sensitivity to neurohormonal stimulation (using 451 phenylephrine). Indeed, patients with DM experience neurohormonal dysregulation. During 452 HF, neurohormonal systems like norepinephrine, ANG-II, aldosterone, and neprilysin are 453 activated, causing impaired insulin sensitivity and microvascular complications (Doliba et al., 454 2018; Packer, 2017). Reduced insulin sensitivity and adipokine abnormalities are 455 characteristic of DM and pathophysiological for HF.

456 Compared to NHE1, the NHE3 isoform distribution is mainly limited to the kidney and 457 gastrointestinal epithelial cells. The main role of NHE3 in DCM is related to its regulation of 458 Na<sup>+</sup> reabsorption in the proximal tubules, which regulates sodium uptake following 459 glomerular filtration. NHE3 activity is enhanced with neurohormonal stimulation by 460 norepinephrine, ANG-II, and aldosterone in HF. Additionally, insulin, glucose, and some 461 adipokines, which are elevated in T2DM, stimulate NHE3. In HF, NHE3 activity is elevated 462 in the kidney mediating Na<sup>+</sup> reabsorption, leading to fluid and Na<sup>+</sup> retention, peripheral 463 oedema, and diuretic resistance. NHE3 hyperactivity in DM also leads to kidney mesangial 464 cell proliferation, hyperfiltration, and diabetic nephropathy, contributing to cardiac overload 465 and further worsening of HF (Packer, 2017; Silva dos Santos et al., 2019). Considering all 466 these pathophysiological changes, concluding that the NHE family could link HF and DM is 467 reasonable.

468 Regarding the SGLT family, evidence shows enhanced SGLT1 expression in end-469 stage cardiomyopathy in obese mice with T2DM. Controversially, reduced expression of 470 SGLT1 is recorded in T1DM. This suggests that the increase in its expression might be 471 attributed to the hyperinsulinemia state found in T2DM, but not T1DM. SGLT1 expression 472 was linked to cardiac fibrosis and collagen deposition in the heart (Zhou et al., 2015). 473 Hypertrophic cardiomyopathy was induced through a transverse aortic constriction in a titin-474 truncated mouse model that increases interstitial fibrosis in wild-type mice without affecting 475 SGLT1 deficient mice. Additionally, SGLT1 contributes to the oxidative stress seen in DCM, 476 as its destruction in mouse atrium cardiomyocytes protects the cells against hypoxia and 477 reoxygenation injury (Kuznetsov et al., 2015). Furthermore, mice with cardiomyocyte-478 specific SGLT1 knockdown were resistant to both in-vivo and ex-vivo myocardial

479 ischemia/reperfusion injury (Yoshii et al., 2019).

480 SGLT2 is an isoform mainly present in the kidneys, while there is a limited-to-no 481 expression in the heart. However, the cardioprotective effects of SGLT2i suggest that SGLT2 482 is involved in DCM by expression in the kidney; there is an increased expression of renal 483 SGLT2 and enhanced glucose reabsorption (Vallon & Sharma, 2010). Studies using knockout 484 mice as a negative control have shown an enhanced SGLT2 expression in T2DM and T1DM 485 mice. However, the biological mechanism for SGLT2 upregulation in DM is not understood. 486 A study with human embryonic cells (HEK-293T) showed that insulin phosphorylated the 487 SGLT2 Ser624 residue, which increased ROS production, further damaging kidney cells 488 (Novikov & Vallon, 2016). Interestingly, using hypoinsulinemic T1DM, there was also 489 enhanced expression of SGLT2, which suggests the involvement of other regulatory proteins.

490 ANG-II can increase SGLT2 expression, and its role in inducing cardiac hypertrophy, heart

491 failure, and DCM is proven. This shows a link between the expression of SGLT2 in the

492 kidneys and DCM (Vallon & Sharma, 2010).

#### 493 **5.3 Hypertension**

494 Hypertension occurs as an autoregulatory response to increased Na<sup>+</sup> concentration due to 495 increased reabsorption. Na<sup>+</sup> reabsorption is mediated by activating the RAAS and the 496 consequent triggering of the ANG-II Type 1 (AT1) receptor, stimulating NHE3-induced Na<sup>+</sup> 497 influx (Dominguez Rieg et al., 2016). Increased Na<sup>+</sup> influx promotes the significant 498 expansion of extracellular volume and cardiac output and mediates a rise in peripheral 499 vascular resistance resulting in elevated BP (Girardi & Sole, 2012). Hypertension signals the 500 body to promote re-establishment of the expanded volume via decreased eGFR followed by 501 pressure natriuresis.

502 Overexpression of NHE3 in proximal tubules was detected in the spontaneously 503 hypertensive rat (SHR) model of human primary hypertension (X. C. Li et al., 2019). 504 Interestingly, ANG-II leads to the overexpression of NHE3 in cultured cells of the proximal 505 tubules as it stimulates the exocytosis of NHE3. In fact, it was found that along with NHE 506 regulatory factor 1, IRBIT protein forms a complex with NHE3 during exocytosis after ANG-507 II stimulation. (He et al., 2016). Other anti-natriuretic peptide hormones such as insulin and 508 glucocorticoid caused the activation of NHE3 in proximal tubules (Fuster et al., 2007; X. C. 509 Li et al., 2019; Pao et al., 2010; Wang et al., 2007). Li et al. (2015) studied the role of NHE3 in hypertension using NHE3<sup>-/-</sup> mice with the transgenic rescue of NHE3 in the small intestine 510 511 and affirmed their hypothesis that NHE3 is essential for ANG-II induced hypertension. In 512 mice with ANG-II-induced hypertension, the selective genetic deletion of NHE3 of the 513 proximal tubule attenuated the condition (X. C. Li et al., 2019). Studies showed that 50% of 514 hypertensive individuals were insulin resistant. Moreover, hypertensive patients are at an 515 high risk of developing CVDs (Lima et al., 2009). NHE3 participates in Na<sup>+</sup> reabsorption in 516 proximal tubules and plays a critical role in the absorption of dietary Na<sup>+</sup> from the gut. Two 517 studies had investigated the role of gut NHE3 using oral NHE3 inhibitor with low systemic 518 absorption on obese SHR. The treatment had significantly reduced the absorption of Na<sup>+</sup> 519 from the gut and reduced BP (Linz et al., 2016; Linz et al., 2012). 520 NHE1 contributes to pH, salt, and volume regulation, linking it to hypertension. Using 521 NHE1-overexpressing transgenic mice, Kuro-o et al. (1995) showed that NHE1

- 522 overexpression caused salt-sensitive BP elevation in mice. Primary hypertensive animal
- 523 models and peripheral cells of primary hypertensive donors also showed increased NHE1
- 524 activation (Orlov et al., 1999). Conversely, NHE1 knockout in mice leads to a reduction in
- 525 BP and artery tension (Boedtkjer et al., 2012). It is suggested that NHE1 overactivity in
- 526 VSMCs increases intracellular Na<sup>+</sup>, reduces Na<sup>+</sup>/Ca<sup>2+</sup> exchangers, and leads to elevated
- 527 intracellular Ca<sup>2+</sup> and increased contraction. With chronic NHE1 overactivation, abnormal
- 528 cell proliferation can occur in VSMCs (Bobulescu et al., 2005). In proximal tubules,
- 529 alterations in Na<sup>+</sup> transporters impact the extracellular volume thus changing BP
- 530 independently from transporters in other renal segments. In hypertension, there is an increase
- 531 in Na<sup>+</sup> reabsorption that mainly occurs in the proximal tubule and loop of Henle. SGLT2,
- 532 which is localized in the proximal tubule, is responsible for 60-90% of renal uptake of Na+
- and glucose (Cianciolo et al., 2020; Thomas & Cherney, 2018).
- 534 The relationship between SGLT2 activity and hypertension is not known yet. When
- 535 Bautista et al. (2004) compared SGLT2 activity in the proximal tubule of renovascular
- 536 hypertensive rats with normotensive rats, they saw that Na<sup>+</sup>-dependent glucose uptake and
- 537 SGLT2 expression were higher in the renovascular hypertensive group. In chronically infused
- 538 ANG-II Wistar rats, the activity and expression of SGLT2 were increased. Using EMPA did
- not affect the BP; however, losartan, a RAAS inhibitor, reduced BP. In this study, Losartan
- 540 prevented renal damage, while EMPA produced a minimal protective effect. Nonetheless,
- 541 EMPA attenuated oxidative stress (Reyes-Pardo et al., 2019). Clinical trials have consistently
- shown that SGLT2i reduces BP (Sanidas et al., 2020). In the EMPA-REG OUTCOME trial,
- 543 EMPA was correlated with minimal BP reduction (Zinman et al., 2015). Similarly, CANVAS
- and CANVAS-R studies showed a reduction in systolic BP by 3.9 mmHg in the canagliflozin
- 545 (CANA) treated group compared to placebo (Neal et al., 2017). In the DECLARE-TIMI 58
- 546 trial, patients treated with DAPA had lower BP by 2.7 mmHg versus placebo (Wiviott et al.,
- 547 2018). A meta-analysis comprised of 27 RCTs with 12,960 participants concluded that
- 548 SGLT2i resulted in lower systolic and diastolic BP by 4 mmHg (95%CI, -4.4 to -3.5), and 1.6
- 549 mmHg (95%Cl, -1.9 to -1.3), respectively from baseline (Baker et al., 2014).
- 550 SGLT2 upregulation could be a partial contributor to hypertension pathogenesis, however
- 551 several hypotheses explain the role of SGLT2 and its inhibition in BP regulation (Sanidas et
- al., 2020). Diuresis associated with SGLT2i may cause reduced BP. However, diuresis is a
- 553 temporary SGLT2i effect, while BP reduction from baseline is a sustained effect (Filippatos
- 554 et al., 2016; Sanidas et al., 2020).

555 A direct relationship between SGLT1 and BP has not been established. SGLT1 deficient 556 (SGLT1<sup>-/-</sup>) mice show glucose-galactose malabsorption; however, the absence of SGLT1 did 557 not affect BP compared to wild-type mice (Gorboulev et al., 2012). BP exhibits a diurnal 558 rhythm and SGLT1 expression exhibits a similar rhythm with the highest expression in the 559 morning (Tavakkolizadeh et al., 2001). Remarkably, a hypertensive animal model showed a 560 downregulation in SGLT1 function and expression (Mate et al., 2006). More research is 561 needed to determine how SGLT1 is involved in hypertension pathophysiology (Poulsen et 562 al., 2015).

563 In summary, SGLT and NHE exhibit different roles in hypertension. Hypertension can 564 worsen the prognosis of DCM where it contributes to the enlargement of the cardiac wall 565 thickness and mass. Increased BP, along with other stimuli, causes vasoconstriction and fluid 566 overload that aggravates cardiac hypertrophy and fibrosis of the myocardium.

#### 567 6. Available Inhibitors and Their Clinical Outcomes

#### 568 6.1 Clinical Evaluation of NHE-1 Inhibitors

569 Substantial evidence supports the protective role of inhibiting NHE1 in reducing IRI 570 development, cardiac hypertrophy, systolic dysfunction, and HF. Several NHE1 inhibitor 571 studies (e.g., cariporide, eniporide, and zoniporide) showed significant protection against CV 572 injuries (Packer, 2017). Despite that, clinical studies in human subjects showed varying 573 results. Therefore, a cardioprotective role of NHE1 inhibition in humans is controversial. 574 The ESCAMI randomized trial investigated eniporide effect on patients (n=1389) with ST-575 elevation MI (Zeymer et al., 2001) for the primary outcome of the change in infarct size with 576 eniporide as add-on therapy to reperfusion in IRI. However, eniporide did not reduce the 577 infarct size nor improve patients' clinical outcomes. However, the protective effect of 578 cariporide in animal models may have been due to the administration of cariporide during 579 ischemia and not during reperfusion (Klein et al., 2000). Rupprecht et al. (2000) tested the 580 effect of cariporide (40 mg) on 100 patients with acute anterior MI getting direct coronary 581 angioplasty. Compared to placebo, patients who received cariporide had higher ejection 582 fraction (50% vs. 40%; P<0.05), lower end-systolic volume (69 vs. 97 mL; P<0.05), 583 significant improvement in wall motion abnormalities, and reduced cumulative release of 584 CK-MB (p= 0.047). Thus, NHE inhibition by cariporide may prevent reperfusion injury and 585 aid in the recovery from ventricular dysfunction. This study contradicts the ESCAMI study's 586 findings concerning the effects of NHE inhibition, as an adjunct to reperfusion therapy, on

587 the myocardium. The GUARDIAN study assessed the safety and efficacy of cariporide (20, 588 80, or 120 mg) in a cohort of patients (n=11,590) at risk for myocardial necrosis (Chaitman, 589 2003). The cardioprotective effect was only evident in patients who underwent coronary 590 artery bypass graft surgery (CABG) and treated with 120 mg cariporide. The EXPEDITION 591 study was the first phase-3 myocardial protection trial to examine cardioprotective effects of 592 cariporide in high-risk patients (n=5,761) undergoing CABG (Mentzer et al., 2008). The drug 593 resulted in increased mortality rates associated with increased cerebrovascular events (2.2% 594 with cariporide vs. 1.5% with placebo; P=0.02). The incidence of death or MI was 595 significantly reduced from 20.3% in the placebo group to 16.6% in the cariporide-group 596 (P=0.0002). However, due to the increased mortality, the study was early terminated. The 597 findings suggested that NHE1 inhibition could significantly reduce ischemia-reperfusion 598 injuries and that cariporide is unlikely to be used clinically. The mixed findings obtained 599 from the clinical research of NHE inhibitors conflict with the highly favourable evidence 600 from experimental studies and emphasize the challenges facing the translation of potential 601 therapies from the laboratory to the clinic.

#### 602 6.2 Clinical Evaluation of SGLT Inhibitors

603 As SGLT1 and SGLT2 are considered the primary transporters involved in glucose 604 homeostasis, several drugs have been developed to inhibit their activity. Inhibiting SGLT1 605 results in better post-meal blood glucose control by blocking glucose uptake in the intestine, 606 which decreases the glycaemic burden. Furthermore, as most glucose reabsorption processes 607 in the proximal convoluted tubule are mediated by SGLT2, inhibition of this transporter 608 reduces the kidney glucose threshold and excretion of glucose lowers glucose plasma levels. 609 This effect is insulin-independent, and therefore, if this class of inhibitor is used alone, the 610 risk of hypoglycaemia is low. These drugs can also increase weight loss by promoting urinary 611 glucose excretion (Raskin, 2013).

The development of SGLT inhibitors started in 1835 with the discovery of phlorizin,
which was speculated to treat malaria and infections until 1886 when it was reported to cause
glucosuria and renal effects (Chasis et al., 1933; Dominguez Rieg & Rieg, 2019).
Administration of subcutaneous phlorizin to diabetic rats with insulin resistance normalized
insulin sensitivity and glucose levels (Rossetti, Shulman, et al., 1987; Rossetti, Smith, et al.,
However, the clinical use of phlorizin was limited due to its poor bioavailability, low

618 solubility, and non-selectivity in SGLT inhibition with increased selectivity to SGLT2

619 compared to SGLT1 (Crespy et al., 2001; Dominguez Rieg & Rieg, 2019).

620 Due to the limitations of phlorizin, other compounds were developed, such as T-1095 and its

621 active form T-1095A which are synthetic compounds derived from phlorizin. Oral T-1095

622 exhibited dose-dependent elevation in urine glucose excretion by inhibiting SGLT2 in the

623 proximal tubule, resulting in reduced blood glucose concentration (Oku et al., 1999).

624 Additionally, T-1095 reduced postprandial blood glucose levels in STZ-induced diabetic rats

via inhibition of SGLT1 in the intestine. However, the clinical use of T-1095 was limited dueto its non-selectivity.

627 Currently, several SGLT2 inhibitors are approved for clinical use in the US and 628 worldwide, and others are under development. Sotagliflozin is an example of a dual SGLT1/2 629 inhibitor with only ~30-folds higher selectivity for SGLT2 over SGLT1, seeking approval by 630 the FDA (Dominguez Rieg & Rieg, 2019). Two randomized controlled trials, SOLOIST-631 WHF and SCORED, randomized T2DM patients with CKD or recent HF hospitalizations, 632 respectively, to receive either sotagliflozin or placebo and found a statistically significant 633 reduction in death from cardiovascular causes, HF hospitalizations, urgent visits for HF, and 634 all-cause mortality (Bhatt, Szarek, Pitt, et al., 2020; Bhatt, Szarek, Steg, et al., 2020). 635 Other SGLT inhibitors are still under investigation, such as Mizagliflozin, a selective SGLT1 636 inhibitor; and licogliflozin, a dual SGLT1/2 inhibitor (Dominguez Rieg & Rieg, 2019). 637 Recently, several SGLT2i were developed and approved to be used in T2DM patients. In 638 addition to their glucose-lowering effects, CANA DAPA and EMPA showed clinical 639 evidence of improved clinical outcome of HF, chronic kidney disease, and CVD in patients 640 with adequate eGFR.(García-Ropero et al., 2019). The CANVAS program joined the analysis 641 of CANVAS and CANVAS-R, which included patients with T2DM and increased CV risk to 642 assess CANA use compared to placebo (Neal et al., 2017). The CANVAS trial assessed CV 643 risk and major adverse cardiac events, while the CANVAS-R trial assessed the progression of 644 albuminuria in patients using CANA versus placebo. The combined analysis showed CANA 645 lowers CV events and probably attenuates albuminuria progression. However, it increases the 646 risk of metatarsal amputation compared to placebo. EMPA is another example of an SGLT2 647 inhibitor with cardioprotective evidence. In the EMPA-REG OUTCOME trial (Zinman et al., 648 2015), EMPA was reported to reduce CV death by 38%, HF hospitalization by 35%, and 649 death from any cause by 32% in T2DM patients at high CV risk. Additionally, the 650 DECLARE-TIMI trial evaluated the effect of DAPA in patients with T2DM and established 651 CVD or CV risk factors (Wiviott et al., 2018). Although DAPA was associated with lower

652 rates of HF hospitalization or CV death than placebo, there was no difference in major 653 adverse cardiac events between placebo and DAPA. Furthermore, the DAPA-HF and 654 EMPEROR-REDUCED trials found a protective effect of DAPA and EMPA, respectively, 655 against CV death plus HF hospitalizations in HF patients regardless of the presence of 656 diabetes (McMurray et al., 2019; Packer et al., 2020). As per these findings, the American 657 Diabetes Association recommends a combination therapy of metformin and SGLT2 inhibitor 658 for established ASCVD, HF, or chronic kidney disease (ADA, 2021). Other clinical studies pointed to the natriuretic effects of SGLT2i, which impact CV benefits through a reduction in 659 660 fluid retention and the risk of developing HF. Using immunofluorescence, Pessoa et al. (2014) reported that NHE3 co-localizes with SGLT2, not SGLT1, concluding that SGLT2i 661 662 causes diuresis via NHE3 inhibition. A recent randomized placebo-controlled crossover study in 20 patients with T2DM and HF treated with EMPA monotherapy showed a significant 663 664 increase in fractional excretion of Na+ (FENa) compared to placebo (P=0.001). A synergistic 665 effect on the FENa was reported when combined with bumetanide (P=0.001). Moreover, after 666 14 days of SGLT2 inhibition by EMPA and its persistent natriuretic effect, there was a 667 reduction in blood volume (P=0.035) and plasma volume (P=0.04) without inducing 668 neurohormonal activation, off-target electrolyte wasting, and renal dysfunction. Thus, the benefits of long-term use of EMPA in HF patients may be volume management attributed to 669 670 the natriuretic effects (Griffin et al., 2020).

#### 671 **7. Conclusion**

672 Diabetes mellitus is highly associated with cardiovascular disease, as hyperglycaemia triggers cardiac metabolic imbalances, endothelial dysfunction, ROS production, RAAS 673 activation, and impaired Ca<sup>2+</sup> homeostasis, leading to heart failure. There is an increasing 674 675 evidence supporting the cardioprotective role of SGLT2i. Overall, there is an interplay 676 between SGLT and NHE in mediating cardiac effects seen in the failing hearts. NHE1 and 677 NHE3 are two well-studied isoforms involved in renal and cardiovascular homeostasis. In the 678 heart, NHE1 regulates intracellular pH, cell volume, proliferation, and Na<sup>+</sup> concentration, which in turn plays a role in  $Ca^{2+}$  regulation, metabolism, contractility, and stability of the 679 680 heart. On the other hand, renal NHE3 contributes to the regulation of extracellular volume 681 and BP. While the role of NHE1 inhibition is well defined, the exact cardioprotective 682 mechanism of SGLT2 inhibition has not been determined, with the exemption of SGLT2i 683 directed NHE1 inhibition. Further studies are needed to investigate the interaction between 684 NHE3 and SGLT2.

#### 685 Figure Captions

- 686 **Figure 1.** Glucose reabsorption through SGLT1 & SGLT2 in the normal kidney
- 687 **Figure 2.** Potential Pathways Underlying the Hypertrophic Effect of NHE1
- 688 (A) During non-ischemic events (normal conditions) NHE is relatively quiescent. The Na+
- 689 K+ ATPase (Na<sup>+</sup> pump) utilizes ATP to extrude Na<sup>+</sup>, and the bidirectional Na<sup>+</sup>/Ca<sup>2+</sup>
- 690 exchanger works predominantly in the forward ( $Ca^{2+}$  efflux) mode. (B) During ischemic
- 691 events [Na<sup>+</sup>]<sub>i</sub> rises during ischemia concomitant with a fall in pH. NHE becomes activated in
- 692 response to intracellular acidosis and other hypertrophic stimulatory factors. Since the  $Na^+/K^+$
- 693 ATPase becomes inactive during ischemia, NHE-mediated Na<sup>+</sup> influx leads to the
- 694 intracellular accumulation of Na<sup>+</sup>. Increased Na<sup>+</sup> elevates intracellular Ca<sup>2+</sup> by altering the
- reversal potential of  $Na^+/Ca^{2+}$  exchangers. Elevated  $Ca^{2+}$  activates various pro-hypertrophic
- 696 factors, including CaN and CaMKII, and increases MPTP, contributing to mitochondrial
- 697 remodelling. Mitochondrial remodelling results in increased ROS production, which in
- 698 combination with other factors contributes to activating transcriptional factors resulting in
- 699 cardiac hypertrophy.
- 700 Figure 3. The Role of SGLT, NHE, and their inhibitors, in Diabetes and Cardiovascular
- 701 **Diseases.** Increased SGLT activity in the proximal tubules leads to decreased natriuresis and
- 702 increased reabsorption of glucose, worsening heart failure and diabetes, respectively. In the
- 703 heart, hypertrophic signals such as endothelin-1, ANG-II, thrombin, and norepinephrine
- 704 increase NHE1 activity, leading to Na<sup>+</sup> accumulation and mitochondrial dysfunction which
- activates pro-hypertrophic transcription factor. Hyperglycaemia leads to increased glucose
- transport through SGLT1, leading to increased NOX2 activity, and subsequent damage to the
- 707 cardiomyocytes through ROS.
- 708

### 709 **References**

710	Ackermann, T. F., Boini, K. M., Völkl, H., Bhandaru, M., Bareiss, P. M., Just, L., Vallon, V.,
711	Amann, K., Kuhl, D., Feng, Y., Hammes, HP., & Lang, F. (2000). SGK1-sensitive renal
712	tubular glucose reabsorption in diabetes American Journal of Physiology-Renal
713	Physiology 206(4) E850-E866 https://doi.org/10.1152/aiprenal.00228.2008
714	$1 \text{ mystology}, 290(4), 1059 1000. \underline{\text{mtps://doi.org/10.1152/djptenat.90250.2000}}$
715	ADA. (2021, Jan). 9. Pharmacologic Approaches to Glycemic Treatment: Standards of Medical
716	$C_{are in Diabetes 2021}$ Diabetes Care $(4(Suppl 1) Superior Standards of Medical$
717	https://doi.org/10.2227/doi.Sooo
719	<u>Inttps://doi.org/10.233//dc2i-5009</u>
710	Ali A Dain C Histo D Newland Janes D Datal D C Evens M Formanda K James J
719	All, A., Baln, S., HICKS, D., Newland Jones, P., Patel, D. C., Evans, M., Fernando, K., James, J.,
720	Milline, N., Viljoen, A., & Wilding, J. (2019, Oct). SGL12 Inhibitors: Cardiovascular
/21	Benefits Beyond HbAic-Translating Evidence into Practice. Diabetes Ther, 10(5), 1595-
722	1622. <u>https://doi.org/10.1007/\$13300-019-0657-8</u>
723	
724	Allen, D. G., & Xiao, X. H. (2003, Mar 15). Role of the cardiac Na+/H+ exchanger during
725	ischemia and reperfusion. <i>Cardiovasc Res, 57</i> (4), 934-941.
726	<u>https://doi.org/10.1016/s0008-6363(02)00836-2</u>
727	
728	American Diabetes, A. (2017, Jan). 2. Classification and Diagnosis of Diabetes. 40(Suppl 1), S11-
729	S24. https://doi.org/10.2337/dc17-S005
730	
731	Aronow, W. S., & Ahn, C. (1999, Mar). Incidence of heart failure in 2,737 older persons with
732	and without diabetes mellitus. <i>Chest.</i> 115(3), 867-868.
733	https://doi.org/10.1378/chest.115.3.867
734	
735	Aviello, G., & Knaus, U. G. (2018, 2018/07/01). NADPH oxidases and ROS signaling in the
736	gastrointestinal tract. Mucosal Immunology, 11(4), 1011-1023.
737	https://doi.org/10.1028/s41285-018-0021-8
738	<u>meps, / uonorg/10.1030/341303 010 0021 0</u>
739	Avkiran M & Haworth R S (2002 Mar 15) Regulatory effects of G protein-coupled receptors
740	on cardiac sarcolemmal Nat /Ht exchanger activity: signalling and significance
741	Cardiovase Pag. $r_{\pi}(A) = A_{\pi} + C_{\pi}(A)$ of a regulation of base of the second s
742	Curatovasc Res, 57(4), 942-952. https://doi.org/10.1010/s0008-0303(02)00782-4
742	Avegare A. Albiere M. Menegazze I. de Vreutzenberg S. & Edini C. D. (2011)
743	Find the liel Durfumetion in Disketon. The web of venerations much misme
/44	Endotnelial Dystunction in Diabetes. The role of reparatory mechanisms,
745	34(Supplement 2), 5285-5290. <u>https://doi.org/10.2337/dc11-s239</u>
/40	
/4/	Avogaro, A., Fadini, G. P., Gallo, A., Pagnin, E., & de Kreutzenberg, S. (2006, 2006/03/01/).
748	Endothelial dysfunction in type 2 diabetes mellitus. <i>Nutrition, Metabolism and</i>
749	Cardiovascular Diseases, 16, S39-S45.
750	https://doi.org/https://doi.org/10.1016/j.numecd.2005.10.015
751	
752	Baker, W. L., Smyth, L. R., Riche, D. M., Bourret, E. M., Chamberlin, K. W., & White, W. B.
753	(2014, Apr). Effects of sodium-glucose co-transporter 2 inhibitors on blood pressure: a
754	systematic review and meta-analysis. <i>J Am Soc Hypertens, 8</i> (4), 262-275.e269.
755	https://doi.org/10.1016/j.jash.2014.01.007
756	
757	Balteau, M., Tajeddine, N., de Meester, C., Ginion, A., Des Rosiers, C., Brady, N. R.,
758	Sommereyns, C., Horman, S., Vanoverschelde, J. L., Gailly, P., Hue, L., Bertrand, L., &
759	Beauloye, C. (2011, Nov 1). NADPH oxidase activation by hyperglycaemia in

760	cardiomyocytes is independent of glucose metabolism but requires SGLT1. Cardiovasc
761	<i>Res</i> , 92(2), 237-246. <u>https://doi.org/10.1093/cvr/cvr230</u>
762	
763	Batzias, K., Antonopoulos, A. S., Oikonomou, E., Siasos, G., Bletsa, E., Stampouloglou, P. K.,
764	Mistakidi, CV., Noutsou, M., Katsiki, N., Karopoulos, P., Charalambous, G.,
765	Thanopoulou, A., Tentolouris, N., & Tousoulis, D. (2018, 2018/12/04). Effects of Newer
766	Antidiabetic Drugs on Endothelial Function and Arterial Stiffness: A Systematic
767	Review and Meta-Analysis. Journal of Diabetes Research, 2018, 1232583.
768	https://doi.org/10.1155/2018/1232583
769	
770	Bautista, R., Manning, R., Martinez, F., Avila-Casado Mdel, C., Soto, V., Medina, A., &
771	Escalante, B. (2004, Jan). Angiotensin II-dependent increased expression of Na+-
772	glucose cotransporter in hypertension. Am J Physiol Renal Physiol, 286(1), F127-133.
773	https://doi.org/10.1152/ajprenal.00113.2003
774	
775	Bhatt, D. L., Szarek, M., Pitt, B., Cannon, C. P., Leiter, L. A., McGuire, D. K., Lewis, J. B.,
776	Riddle, M. C., Inzucchi, S. E., Kosiborod, M. N., Cherney, D. Z. I., Dwyer, J. P., Scirica,
777	B. M., Bailey, C. J., Díaz, R., Ray, K. K., Udell, J. A., Lopes, R. D., Lapuerta, P., & Steg, P.
778	G. (2020, 2021/01/14). Sotagliflozin in Patients with Diabetes and Chronic Kidney
779	Disease. New England Journal of Medicine, 384(2), 129-139.
780	https://doi.org/10.1056/NEJM0a2030186
781	
782	Bhatt, D. L., Szarek, M., Steg, P. G., Cannon, C. P., Leiter, L. A., McGuire, D. K., Lewis, J. B.,
783	Riddle, M. C., Voors, A. A., Metra, M., Lund, L. H., Komajda, M., Testani, J. M., Wilcox,
784	C. S., Ponikowski, P., Lopes, R. D., Verma, S., Lapuerta, P., & Pitt, B. (2020, 2021/01/14).
785	Sotagliflozin in Patients with Diabetes and Recent Worsening Heart Failure. New
786	England Journal of Medicine, 384(2), 117-128. https://doi.org/10.1056/NEJM0a2030183
787	
788	Bobulescu, I. A., Di Sole, F., & Moe, O. W. (2005). Na+/H+ exchangers: physiology and link to
789	hypertension and organ ischemia. <i>Current opinion in nephrology and hypertension</i> ,
790	14(5), 485-494. https://doi.org/10.1097/01.mnh.0000174146.52915.5d
791	
792	Boedtkjer, E., Damkier, H. H., & Aalkjaer, C. (2012, Apr 15). NHE1 knockout reduces blood
793	pressure and arterial media/lumen ratio with no effect on resting pH(i) in the vascular
794	wall. J Physiol, 590(8), 1895-1906. https://doi.org/10.1113/jphysiol.2011.227132
795	
796	Chaitman, B. R. (2003, 2003/01/01). A Review of the GUARDIAN Trial Results: Clinical
797	Implications and the Significance of Elevated Perioperative CK-MB on 6-Month
798	Survival, Journal of Cardiac Surgery, 18(s1), 13-20, https://doi.org/10.1046/j.1540-
799	8191.18.51.3.X
800	
801	Chao, E. C. (2014). SGLT-2 Inhibitors: A New Mechanism for Glycemic Control. <i>Clinical</i>
802	diabetes : a publication of the American Diabetes Association, $32(1)$ , 4-11.
803	https://doi.org/10.2337/diaclin.32.1.4
804	<u>-</u>
805	Chasis, H., Jolliffe, N., & Smith, H. W. (1933). THE ACTION OF PHLORIZIN ON THE
806	EXCRETION OF GLUCOSE. XYLOSE. SUCROSE. CREATININE AND UREA BY MAN
807	The Journal of clinical investigation, 12(6), 1082-1000, https://doi.org/10.1172/JC100550
808	,

809	Cianciolo, G., De Pascalis, A., Gasperoni, L., Tondolo, F., Zappulo, F., Capelli, I., Cappuccilli,
810	M., & La Manna, G. (2020, Jun 15). The Off-Target Effects, Electrolyte and Mineral
811	Disorders of SGLT2i. <i>Molecules</i> , 25(12). <u>https://doi.org/10.3390/molecules25122757</u>
812	
813	Cingolani Horacio, E., & Ennis Irene, L. (2007, 2007/03/06). Sodium-Hydrogen Exchanger,
814	Cardiac Overload, and Myocardial Hypertrophy. <i>Circulation</i> , 115(9), 1090-1100.
815	https://doi.org/10.1161/CIRCULATIONAHA.106.626929
816	
817	Clement, D. L., Mally, S., Stock, C., Lethan, M., Satir, P., Schwab, A., Pedersen, S. F., &
818	Christensen, S. T. (2013). PDGFR $\alpha$ signaling in the primary cilium regulates NHE1-
819	dependent fibroblast migration via coordinated differential activity of MEK1/2-ERK1/2-
820	pooRSK and AKT signaling pathways <i>Journal of cell science</i> , $126(Pt A)$ , 052-065
821	https://doi.org/10.1242/ics.116426
822	<u>Ittp://doi.org/10.1242/jcs.110420</u>
823	Crespy V Aprilian O Morand C Besson C Manach C Demigné C & Rémésy C (2001
823	Dec) Bioavailability of phloretin and phloridzin in rate <i>LNutr</i> 12(12) 2227 2220
02 <del>4</del> 825	bttps://doi.org/10.1002/in/12.12.227
02 <i>3</i> 876	<u>11(1ps.//doi.org/10.1093/j11/131.12.3227</u>
020 927	Das A. K. Das I. D. & Chandraselear S. (108-) Specific heart muscle disease in dishetes
021	Das, A. K., Das, J. F., & Chandrasekar, S. (1907). Specific field infuscie disease in underes
020 920	$\frac{1}{2}$
029	17(3), 299-302. <u>https://doi.org/10.1016/0107-5273(87/90080-5</u>
830	
831	De vriese, A. S., verbeuren, T. J., van de voorde, J., Lameire, N. H., & vannoutte, P. M. (2000,
832	2000/07/01). Endothelial dysfunction in diabetes
833	[ <u>https://doi.org/10.1038/sj.bjp.0703393</u> ]. British Journal of Pharmacology, 130(5), 963-
834	974. <u>https://doi.org/https://doi.org/10.1038/sj.bjp.0703393</u>
835	
836	Di Franco, A., Cantini, G., Tani, A., Coppini, R., Zecchi-Orlandini, S., Raimondi, L., Luconi, M.,
83/	& Mannucci, E. (2017, 2017/09/15/). Sodium-dependent glucose transporters (SGLT) in
838	human ischemic heart: A new potential pharmacological target. International Journal
839	of Cardiology, 243, 86-90. <u>https://doi.org/https://doi.org/10.1016/j.ijcard.2017.05.032</u>
840	
841	Doliba, N. M., Babsky, A. M., & Osbakken, M. D. (2018, 2018-October-24). The Role of Sodium
842	in Diabetic Cardiomyopathy [Review]. Front Physiol, 9(1473).
843	https://doi.org/10.3389/fphys.2018.01473
844	
845	Dominguez Rieg, J. A., de la Mora Chavez, S., & Rieg, T. (2016). Novel developments in
846	differentiating the role of renal and intestinal sodium hydrogen exchanger 3. American
847	journal of physiology. Regulatory, integrative and comparative physiology, 311(6), R1186-
848	R1191. <u>https://doi.org/10.1152/ajpregu.00372.2016</u>
849	
850	Dominguez Rieg, J. A., & Rieg, T. (2019). What does sodium-glucose co-transporter 1 inhibition
851	add: Prospects for dual inhibition. <i>Diabetes</i> , obesity & metabolism, 21 Suppl 2(Suppl 2),
852	43-52. <u>https://doi.org/10.1111/dom.13630</u>
853	
854	Drumm, K., Lee, E., Stanners, S., Gassner, B., Gekle, M., Poronnik, P., & Pollock, C. A. (2003).
855	Albumin and Glucose Effects On Cell Growth Parameters, Albumin Uptake and
856	Na <sup>+</sup> /H <sup>+</sup> -Exchanger Isoform 3 in OK Cells. <i>Cellular</i>
857	Physiology and Biochemistry, 13(4), 199-206. <u>https://doi.org/10.1159/000072422</u>
858	

859	du Cheyron, D., Chalumeau, C., Defontaine, N., Klein, C., Kellermann, O., Paillard, M., &
860	Poggioli, J. (2003). Angiotensin II stimulates NHE3 activity by exocytic insertion of the
861	transporter: Role of PI 3-kinase. <i>Kidney International, 64</i> (3), 939-949.
862	<u>https://doi.org/10.1046/j.1523-1755.2003.00189.x</u>
863	
864	Dynia, D. W., Steinmetz, A. G., & Kocinsky, H. S. (2010). NHE3 function and phosphorylation
865	are regulated by a calyculin A-sensitive phosphatase. American journal of physiology.
866	Renal physiology, 298(3), F745-F753. <u>https://doi.org/10.1152/ajprenal.00182.2009</u>
867	
868	Endemann, D. H., & Schiffrin, E. L. (2004). Endothelial Dysfunction. Journal of the American
869	Society of Nephrology, 15(8), 1983-1992.
870	https://doi.org/10.1097/01.Asn.0000132474.50966.Da
871	
872	Eriksson, L., & Nyström, T. (2015, Jul). Antidiabetic agents and endothelial dysfunction -
873	beyond glucose control. <i>Basic Clin Pharmacol Toxicol, 117</i> (1), 15-25.
874	https://doi.org/10.1111/bcpt.12402
875	
876	Filippatos, T. D., Tsimihodimos, V., & Elisaf, M. S. (2016, 2016/08/12). Mechanisms of blood
877	pressure reduction with sodium-glucose co-transporter 2 (SGLT2) inhibitors. <i>Expert</i>
878	Opinion on Pharmacotherapy, 17(12), 1581-1583.
8/9	https://doi.org/10.1080/14656566.2016.1201073
880	
881	Fuentes-Antras, J., Picatoste, B., Ramirez, E., Egido, J., Tunon, J., & Lorenzo, O. (2015).
882	largeting metabolic disturbance in the diabetic heart. Cardiovascular diabetology, 14,
883	17-17. <u>https://doi.org/10.1186/s12933-015-0173-8</u>
884 885	Eventer D.C. Dehulanny, I.A. Zhang, I. Wada, I. & Mag, O. W. (appendix). Characterization
003	Fuster, D. G., Bobulescu, I. A., Zhang, J., Wade, J., & Moe, O. W. (2007, Feb). Characterization
000	of the regulation of renal Na+/H+ exchanger NHE3 by insum. American journal of
00/ 888	https://doi.org/10.1152/2ipropal.002.40.2006
880	<u>intips://doi.org/10.1152/ajprenai.00240.2000</u>
890	Canbaatar B. Fukuda D. Vagi S. Kusunose K. Vamada H. Soeki T. & Sata M. (2010)
891	PourFmpagliflozin a SGLT2 inhibitor attenuates endothelial dysfunction and
892	atherogenesis by inhibiting inflammatory responses in the vasculature and adipose
893	tissue in diabetic apolipoprotein F-deficient <i>Furopean Heart Journal</i>
894	(Supplement 1) https://doi.org/10.1003/eurhearti/ehz745.0186
895	40(0upplement_1). <u>https://doi.org/10.1095/cumeart//cm2/45.0100</u>
896	García-Ropero, Á., Vargas-Delgado, A. P., Santos-Gallego, C. G., & Badimon, I. I. (2010, Jul 4).
897	Inhibition of Sodium Glucose Cotransporters Improves Cardiac Performance.
898	International journal of molecular sciences, 20(13), https://doi.org/10.3300/jims20133280
899	
900	Gaspari, T., Spizzo, I., Liu, H., Hu, Y., Simpson, R. W., Widdop, R. E., & Dear, A. E. (2018, Jan).
901	Dapagliflozin attenuates human vascular endothelial cell activation and induces
902	vasorelaxation: A potential mechanism for inhibition of atherogenesis. <i>Diab Vasc Dis</i>
903	Res, 15(1), 64-73. https://doi.org/10.1177/1479164117733626
904	
905	Girardi, A. C. C., & Sole, F. D. (2012). Deciphering the mechanisms of the Na+/H+ exchanger-3
906	regulation in organ dysfunction. American Journal of Physiology-Cell Physiology,
907	302(11), C1569-C1587. https://doi.org/10.1152/ajpcell.00017.2012
908	

909 910 911 912 913 914 915	<ul> <li>Gorboulev, V., Schürmann, A., Vallon, V., Kipp, H., Jaschke, A., Klessen, D., Friedrich, A., Scherneck, S., Rieg, T., Cunard, R., Veyhl-Wichmann, M., Srinivasan, A., Balen, D., Breljak, D., Rexhepaj, R., Parker, H. E., Gribble, F. M., Reimann, F., Lang, F., Wiese, S., Sabolic, I., Sendtner, M., &amp; Koepsell, H. (2012). Na(+)-D-glucose cotransporter SGLT1 is pivotal for intestinal glucose absorption and glucose-dependent incretin secretion. <i>Diabetes</i>, <i>61</i>(1), 187-196. <u>https://doi.org/10.2337/db11-1029</u></li> </ul>
916 917 918 919 920 921	<ul> <li>Griffin, M., Rao, V. S., Ivey-Miranda, J., Fleming, J., Mahoney, D., Maulion, C., Suda, N.,</li> <li>Siwakoti, K., Ahmad, T., Jacoby, D., Riello, R., Bellumkonda, L., Cox, Z., Collins, S.,</li> <li>Jeon, S., Turner, J. M., Wilson, F. P., Butler, J., Inzucchi, S. E., &amp; Testani, J. M. (2020,</li> <li>Sep 15). Empagliflozin in Heart Failure: Diuretic and Cardiorenal Effects. <i>Circulation</i>,</li> <li>142(11), 1028-1039. <u>https://doi.org/10.1161/circulationaha.120.045691</u></li> </ul>
922 923 924 925	Hannan, K. M., & Little, P. J. (1998, 1998/10/01). Mechanisms regulating the vascular smooth muscle Na/H exchanger (NHE-1) in diabetes. <i>Biochemistry and Cell Biology, 76</i> (5), 751-759. <u>https://doi.org/10.1139/098-093</u>
926 927 928 929 930	He, P., Klein, J., & Yun, C. C. (2010). Activation of Na+/H+ exchanger NHE3 by angiotensin II is mediated by inositol 1,4,5-triphosphate (IP3) receptor-binding protein released with IP3 (IRBIT) and Ca2+/calmodulin-dependent protein kinase II. <i>J Biol Chem</i> , 285(36), 27869-27878. <u>https://doi.org/10.1074/jbc.M110.133066</u>
931 932 933 934 935	He, P., Zhao, L., No, Y. R., Karvar, S., & Yun, C. C. (2016). The NHERF1 PDZ1 domain and IRBIT interact and mediate the activation of Na+/H+ exchanger 3 by ANG II. American Journal of Physiology-Renal Physiology, 311(2), F343-F351. <u>https://doi.org/10.1152/ajprenal.00247.2016</u>
935 936 937 938 939	He, Q., Harding, P., & LaPointe, M. C. (2010, Jan). PKA, Rap1, ERK1/2, and p90RSK mediate PGE2 and EP4 signaling in neonatal ventricular myocytes. <i>Am J Physiol Heart Circ</i> <i>Physiol, 298</i> (1), H136-143. <u>https://doi.org/10.1152/ajpheart.00251.2009</u>
940 941 942 943 944	<ul> <li>Hink, U., Li, H., Mollnau, H., Oelze, M., Matheis, E., Hartmann, M., Skatchkov, M., Thaiss, F., Stahl, R. A., Warnholtz, A., Meinertz, T., Griendling, K., Harrison, D. G., Forstermann, U., &amp; Munzel, T. (2001, Feb 2). Mechanisms underlying endothelial dysfunction in diabetes mellitus. <i>Circ Res</i>, 88(2), E14-22. <u>https://doi.org/10.1161/01.res.88.2.e14</u></li> </ul>
945 946 947 948 949	Horie, S., Moe, O., Yamaji, Y., Cano, A., Miller, R. T., & Alpern, R. J. (1992, Jun 15). Role of protein kinase C and transcription factor AP-1 in the acid-induced increase in Na/H antiporter activity. <i>Proc Natl Acad Sci U S A</i> , <i>89</i> (12), 5236-5240. <u>https://doi.org/10.1073/pnas.89.12.5236</u>
950 951 952 953 954	<ul> <li>Imahashi, K., Mraiche, F., Steenbergen, C., Murphy, E., &amp; Fliegel, L. (2007, 2007/05/01).</li> <li>Overexpression of the Na+/H+ exchanger and ischemia-reperfusion injury in the myocardium. <i>American Journal of Physiology-Heart and Circulatory Physiology</i>, 292(5), H2237-H2247. <a href="https://doi.org/10.1152/ajpheart.00855.2006">https://doi.org/10.1152/ajpheart.00855.2006</a></li> </ul>
955 956 957 958 959	Javadov, S., Rajapurohitam, V., Kilić, A., Zeidan, A., Choi, A., & Karmazyn, M. (2009, Jun). Anti-hypertrophic effect of NHE-1 inhibition involves GSK-3beta-dependent attenuation of mitochondrial dysfunction. <i>J Mol Cell Cardiol, 46</i> (6), 998-1007. <u>https://doi.org/10.1016/j.yjmcc.2008.12.023</u>

960 961	Jia, G., DeMarco, V. G., & Sowers, J. R. (2016). Insulin resistance and hyperinsulinaemia in diabetic cardiomyopathy. <i>Nature reviews. Endocrinology, 12</i> (3), 144-153.
962 963	https://doi.org/10.1038/nrendo.2015.216
964 965 966	Jia, G., Hill Michael, A., & Sowers James, R. (2018, 2018/02/16). Diabetic Cardiomyopathy. <i>Circulation Research</i> , 122(4), 624-638. <u>https://doi.org/10.1161/CIRCRESAHA.117.311586</u>
967 968 969	Kahanovitz, L., Sluss, P. M., & Russell, S. J. (2017). Type 1 Diabetes - A Clinical Perspective. Point of care, 16(1), 37-40. <u>https://doi.org/10.1097/POC.000000000000125</u>
970 971 972	Karmazyn, M. (2003). Role of NHE-1 in Cardiac Hypertrophy and Heart Failure. In M. Karmazyn, M. Avkiran, & L. Fliegel (Eds.), <i>The Sodium-Hydrogen Exchanger: From</i> <i>Molecule to its Role in Disease</i> (pp. 211-219). Springer US. <u>https://doi.org/10.1007/978-1-</u>
973 974	<u>4615-0427-6 14</u>
975 976 977 978	Karmazyn, M., Kilić, A., & Javadov, S. (2008). The role of NHE-1 in myocardial hypertrophy and remodelling. <i>J Mol Cell Cardiol, 44</i> (4), 647-653. <u>https://doi.org/10.1016/j.yjmcc.2008.01.005</u>
979 980 981 982	Kashiwagi, Y., Nagoshi, T., Yoshino, T., Tanaka, T. D., Ito, K., Harada, T., Takahashi, H., Ikegami, M., Anzawa, R., & Yoshimura, M. (2015). Expression of SGLT1 in Human Hearts and Impairment of Cardiac Glucose Uptake by Phlorizin during Ischemia- Reperfusion Injury in Mice. <i>PLoS One, 10</i> (6), e0130605.
983 984	https://doi.org/10.1371/journal.pone.0130605
985 986 987 988 989	Kemi, O. J., Ceci, M., Wisloff, U., Grimaldi, S., Gallo, P., Smith, G. L., Condorelli, G., & Ellingsen, O. (2008, Feb). Activation or inactivation of cardiac Akt/mTOR signaling diverges physiological from pathological hypertrophy. <i>J Cell Physiol</i> , 214(2), 316-321. <u>https://doi.org/10.1002/jcp.21197</u>
990 991 992 993 994	<ul> <li>Khemais-Benkhiat, S., Belcastro, E., Idris-Khodja, N., Park, S. H., Amoura, L., Abbas, M., Auger, C., Kessler, L., Mayoux, E., Toti, F., &amp; Schini-Kerth, V. B. (2020, Feb).</li> <li>Angiotensin II-induced redox-sensitive SGLT1 and 2 expression promotes high glucose-induced endothelial cell senescence. <i>J Cell Mol Med</i>, 24(3), 2109-2122.</li> <li><u>https://doi.org/10.1111/jcmm.14233</u></li> </ul>
995 996 997 998 998	Kitamura, K., Singer, W. D., Cano, A., & Miller, R. T. (1995, Jan). G alpha q and G alpha 13 regulate NHE-1 and intracellular calcium in epithelial cells. <i>Am J Physiol, 268</i> (1 Pt 1), C101-110. <u>https://doi.org/10.1152/ajpcell.1995.268.1.C101</u>
1000 1001 1002 1003	<ul> <li>Klein, H. H., Pich, S., Bohle, R. M., Lindert-Heimberg, S., &amp; Nebendahl, K. (2000, Oct 17). Na(+)/H(+) exchange inhibitor cariporide attenuates cell injury predominantly during ischemia and not at onset of reperfusion in porcine hearts with low residual blood flow. <i>Circulation</i>, 102(16), 1977-1982. <u>https://doi.org/10.1161/01.cir.102.16.1977</u></li> </ul>
1004 1005 1006 1007	Klisic, J., Nief, V., Reyes, L., & Ambuhl, P. M. (2006). Acute and chronic regulation of the renal Na/H+ exchanger NHE3 in rats with STZ-induced diabetes mellitus. <i>Nephron Physiol</i> , 102(2), p27-35. <u>https://doi.org/10.1159/000089091</u>
1008 1009 1010	Kuro-o, M., Hanaoka, K., Hiroi, Y., Noguchi, T., Fujimori, Y., Takewaki, S., Hayasaka, M., Katoh, H., Miyagishi, A., Nagai, R., & et al. (1995, Jan). Salt-sensitive hypertension in

1011	transgenic mice overexpressing Na(+)-proton exchanger. <i>Circ Res, 76</i> (1), 148-153.
1012	<u>https://doi.org/10.1101/01.res.76.1.148</u>
1013	Kurnsteen A.V. Javaden C. Cislinger C. Froteshnig C. & Crimm M. (2017) Hors and H.
1014	Kuzhelsov, A. V., Javadov, S., Sickinger, S., Froischnig, S., & Grinnin, M. (2015). H9C2 and HL-1
1015	cens demonstrate distinct features of energy metabolism, introchondrial function and
1010	sensitivity to hypoxia-reoxygenation. <i>Biochimica et biophysica acta</i> , 1853(2), 2/0-284.
1017	<u>https://doi.org/10.1016/J.bbamcr.2014.11.015</u>
1010	Lembert D. Gradulati C. Dang V. Margulias K. D. Dang F. & Dang G. (across Augus)
1019	Lambert, K., Stoduiski, S., Peng, A., Margunes, K. D., Despa, F., & Despa, S. (2015, Aug 27).
1020	Enhanced Na Chicese Cetransport I Am Heart Assoc (a) 2022
1021	https://doi.org/10.1161/jaba.uz.002182
1022	<u>11(1ps.//doi.org/10.1101/jana.115.002103</u>
1023	Lara P. Sock! M. L. & Pardo, O. F. (2012, Son 1) The pool PSK family members: common
1024	functions and isoform specificity. <i>Cancer Pas.</i> 72(17), 5201, 5208
1025	https://doi.org/10.1158/0008 $= 472$ Cap 12, 4448
1020	<u>inteps.//doi.org/10.1150/0008-54/2.Can-12-4446</u>
1027	Lee D.M. Battson M.I. Jarrell D.K. Hou S. Ecton K.F. Weir T.I. & Centile C.I. (2018)
1020	2018/04/27) SGLT2 inhibition via danagliflozin improves generalized vascular
1029	dysfunction and alters the gut microbiota in type 2 diabetic mice. Cardiovascular
1031	Diabetology $17(1)$ 62 https://doi.org/10.1186/s12022-018-0708-x
1032	$Diabetology, 17(1), 02. \underline{meps.//doi.org/10.1100/012933/010/0700 x}{}$
1032	Lee S.Y. Lee T.W. Park G.T. Kim I.H. Lee HC. Han IH. Yoon A. Yoon D. Kim S.
1034	Iung S M Choi I H Chon M K Lee S H Hwang K W Kim I Park Y H Kim
1035	I H. Chun K L & Hur I (2021 3/) Sodium/glucose Co-Transporter 2 Inhibitor
1036	Empagliflozin, Alleviated Transient Expression of SGLT2 after Myocardial Infarction.
1037	Korean Circ J. $51(3)$ , $251-262$ , https://doi.org/10.4070/kci.2020.0303
1038	
1039	Leon, B. M., & Maddox, T. M. (2015, Oct 10). Diabetes and cardiovascular disease:
1040	Epidemiology, biological mechanisms, treatment recommendations and future
1041	research. World J Diabetes, 6(13), 1246-1258. <u>https://doi.org/10.4239/wjd.v6.i13.1246</u>
1042	
1043	Li, X. C., Shull, G. E., Miguel-Qin, E., Chen, F., & Zhuo, J. L. (2015, Nov). Role of the Na+/H+
1044	exchanger 3 in angiotensin II-induced hypertension in NHE3-deficient mice with
1045	transgenic rescue of NHE3 in small intestines. <i>Physiol Rep</i> , 3(11).
1046	https://doi.org/10.14814/phy2.12605
1047	
1048	Li, X. C., Zheng, X., Chen, X., Zhao, C., Zhu, D., Zhang, J., & Zhuo, J. L. (2019, Apr 1). Genetic
1049	and genomic evidence for an important role of the $Na(+)/H(+)$ exchanger 3 in blood
1050	pressure regulation and angiotensin II-induced hypertension. <i>Physiol Genomics</i> , 51(4),
1051	97-108. <u>https://doi.org/10.1152/physiolgenomics.00122.2018</u>
1052	
1053	Li, Z., Agrawal, V., Ramratnam, M., Sharma, R. K., D'Auria, S., Sincoular, A., Jakubiak, M.,
1054	Music, M. L., Kutschke, W. J., Huang, X. N., Gifford, L., & Ahmad, F. (2019, Sep 1).
1055	Cardiac sodium-dependent glucose cotransporter 1 is a novel mediator of
1056	ischaemia/reperfusion injury. <i>Cardiovasc Res, 115</i> (11), 1646-1658.
1057	https://doi.org/10.1093/cvr/cvz037
1058	
1059	Lima, N. K. C., Abbasi, F., Lamendola, C., & Reaven, G. M. (2009). Prevalence of Insulin
1060	Resistance and Related Risk Factors for Cardiovascular Disease in Patients With

1061	Essential Hypertension. American Journal of Hypertension, 22(1), 106-111.
1062	https://doi.org/10.1038/ajh.2008.263
1063	
1064	Linz, B., Hohl, M., Reil, J. C., Böhm, M., & Linz, D. (2016, Mar). Inhibition of NHE3-mediated
1065	Sodium Absorption in the Gut Reduced Cardiac End-organ Damage Without
1066	Deteriorating Renal Function in Obese Spontaneously Hypertensive Rats. J Cardiovasc
1067	Pharmacol, 67(3), 225-231. https://doi.org/10.1097/fjc.00000000000336
1068	
1069	Linz, D., Wirth, K., Linz, W., Heuer, H. O., Frick, W., Hofmeister, A., Heinelt, U., Arndt, P.,
1070	Schwahn, U., Böhm, M., & Ruetten, H. (2012, Dec). Antihypertensive and laxative
1071	effects by pharmacological inhibition of sodium-proton-exchanger subtype 3-mediated
1072	sodium absorption in the gut. <i>Hypertension</i> , 60(6), 1560-1567.
1073	https://doi.org/10.1161/hypertensionaha.112.201590
1074	
1075	Maamoun, H., Abdelsalam, S. S., Zeidan, A., Korashy, H. M., & Agouni, A. (2019). Endoplasmic
1076	Reticulum Stress: A Critical Molecular Driver of Endothelial Dysfunction and
1077	Cardiovascular Disturbances Associated with Diabetes. International journal of
1078	molecular sciences, 20(7), 1658. <u>https://doi.org/10.3390/ijms20071658</u>
1079	
1080	Martín-Timón, I., Sevillano-Collantes, C., Segura-Galindo, A., & Del Cañizo-Gómez, F. J.
1081	(2014). Type 2 diabetes and cardiovascular disease: Have all risk factors the same
1082	strength? World J Diabetes, 5(4), 444-470. <u>https://doi.org/10.4239/wjd.v5.i4.444</u>
1083	
1084	Mate, A., Barfull, A., Hermosa, A. M., Gómez-Amores, L., Vázquez, C. M., & Planas, J. M.
1085	(2006, Sep). Regulation of sodium-glucose cotransporter SGLT1 in the intestine of
1086	hypertensive rats. Am J Physiol Regul Integr Comp Physiol, 291(3), R760-767.
1087	<u>https://doi.org/10.1152/ajpregu.00524.2005</u>
1088	
1089	McMurray, J. J. V., Solomon, S. D., Inzucchi, S. E., Køber, L., Kosiborod, M. N., Martinez, F. A.,
1090	Ponikowski, P., Sabatine, M. S., Anand, I. S., Bělohlávek, J., Böhm, M., Chiang, CE.,
1091	Chopra, V. K., de Boer, R. A., Desai, A. S., Diez, M., Drozdz, J., Dukát, A., Ge, J.,
1092	Howlett, J. G., Katova, T., Kitakaze, M., Ljungman, C. E. A., Merkely, B., Nicolau, J. C.,
1093	O'Meara, E., Petrie, M. C., Vinh, P. N., Schou, M., Tereshchenko, S., Verma, S., Held,
1094	C., DeMets, D. L., Docherty, K. F., Jhund, P. S., Bengtsson, O., Sjöstrand, M., &
1095	Langkilde, AM. (2019). Dapagliflozin in Patients with Heart Failure and Reduced
1096	Ejection Fraction. <i>New England Journal of Medicine</i> , 381(21), 1995-2008.
1097	https://doi.org/10.1056/NEJM0a1911303
1098	
1099	Mentzer, R. M., Jr., Bartels, C., Bolli, R., Boyce, S., Buckberg, G. D., Chaitman, B., Haverich, A.,
1100	Knight, J., Menasché, P., Myers, M. L., Nicolau, J., Simoons, M., Thulin, L., & Weisel, R.
1101	D. (2008). Sodium-Hydrogen Exchange Inhibition by Cariporide to Reduce the Risk of
1102	Ischemic Cardiac Events in Patients Undergoing Coronary Artery Bypass Grafting:
1103	Results of the EXPEDITION Study. <i>The Annals of Thoracic Surgery</i> , 85(4), 1261-1270.
1104	https://doi.org/10.1016/j.athoracsur.2007.10.054
1105	
1106	Mraiche, F., Oka, T., Gan, X. T., Karmazyn, M., & Fliegel, L. (2011, Jun). Activated NHE1 is
1107	required to induce early cardiac hypertrophy in mice. <i>Basic Res Cardiol</i> , 106(4), 603-
1108	616. <u>https://doi.org/10.1007/s00395-011-0161-4</u>
1109	

1110	Muniyappa, R., & Sowers, J. R. (2013). Role of insulin resistance in endothelial dysfunction.
1111	Reviews in endocrine & metabolic disorders, 14(1), 5-12. <u>https://doi.org/10.1007/s11154-</u>
1112	<u>012-9229-1</u>
1113	
1114 1115	Muthusamy, S., Cheng, M., Jeong, J. J., Kumar, A., Dudeja, P. K., & Malakooti, J. (2013). Extracellular acidosis stimulates NHE2 expression through activation of transcription
1116	factor Egr-1 in the intestinal epithelial cells. PLoS One, 8(12), e82023.
1117	https://doi.org/10.1371/journal.pone.0082023
1118	<u> </u>
1119	Neal, B., Perkovic, V., Mahaffey, K. W., de Zeeuw, D., Fulcher, G., Erondu, N., Shaw, W., Law,
1120	G., Desai, M., & Matthews, D. R. (2017, Aug 17). Canagliflozin and Cardiovascular and Bonal Events in Type 2 Diabeter. <i>N Engl L Med</i> , 277(7), 644, 657
1121	https://doi.org/10.1056/NEIM021611025
1122	<u>Inteps.//doi.org/10.1050/101j10001011925</u>
1123	Nichola C. A. Cullian C. M. Kara C. E. Enhross S. A. & Brown J. D. (2004 Aug.) The
1124	Nichols, G. A., Guinon, C. M., Koro, C. E., Ephross, S. A., & Brown, J. D. (2004, Aug). The
1123	incidence of congestive neart failure in type 2 diabetes: an update. <i>Diabetes Care, 27</i> (8),
1126	1879-1884. <u>https://doi.org/10.2337/diacare.27.8.1879</u>
112/	
1128	No, Y. R., He, P., Yoo, B. K., & Yun, C. C. (2015, Jul 1). Regulation of NHE3 by lysophosphatidic
1129	acid is mediated by phosphorylation of NHE3 by RSK2. Am J Physiol Cell Physiol,
1130	309(1), C14-21. <u>https://doi.org/10.1152/ajpcell.00067.2015</u>
1131	
1132	Novikov, A., & Vallon, V. (2016). Sodium glucose cotransporter 2 inhibition in the diabetic
1133	kidney: an update. <i>Current opinion in nephrology and hypertension, 25</i> (1), 50-58.
1134	<u>https://doi.org/10.1097/MNH.000000000000187</u>
1135	
1136	Odunewu-Aderibigbe, A., & Fliegel, L. (2014, 2014/10/01). The Na+/H+ exchanger and pH
1137	regulation in the heart. <i>IUBMB Life, 66</i> (10), 679-685. <u>https://doi.org/10.1002/iub.1323</u>
1138	
1139	Oku, A., Ueta, K., Arakawa, K., Ishihara, T., Nawano, M., Kuronuma, Y., Matsumoto, M., Saito,
1140	A., Tsujihara, K., Anai, M., Asano, T., Kanai, Y., & Endou, H. (1999). T-1095, an
1141	inhibitor of renal Na+-glucose cotransporters, may provide a novel approach to
1142	treating diabetes. <i>Diabetes, 48</i> (9), 1794. <u>https://doi.org/10.2337/diabetes.48.9.1794</u>
1143	
1144	Orlov, S. N., Adragna, N. C., Adarichev, V. A., & Hamet, P. (1999, Mar). Genetic and
1145	biochemical determinants of abnormal monovalent ion transport in primary
1146	hypertension. Am J Physiol, 276(3), C511-536.
1147	https://doi.org/10.1152/ajpcell.1999.276.3.C511
1148	
1149	Packer, M. (2017). Activation and Inhibition of Sodium-Hydrogen Exchanger Is a Mechanism
1150	That Links the Pathophysiology and Treatment of Diabetes Mellitus With That of
1151	Heart Failure Circulation 126(16) 1548-1550
1152	https://doi.org/doi:10.1161/CIRCUI ATIONAHA 117.020418
1152	<u>meps, aonorg/</u>
1155	Packer M. Anker S. D. Butler I. Filippatos C. Pocock S. I. Carson P. Januzzi I. Verma
1155	S Teuteui H Brueckmann M Jamal W Kimura K Schnee I Zeller C Cotton
1155	D. Bocchi F. Böhm M. Choi D. I. Chopra V. Chuquiura F. Ciappotti N. Jangoong
1150	S. Zhang I. Congaleg Juanatov I. P. Kaul S. Drunner La Dassa, H. D. Markala, P.
115/	S., Zhang, J., Gonzalez Juanaley, J. K., Naul, S., Drunner-La Kocca, HP., Werkely, B., Nicholla S. L. Dorrono, S. Dina, L. Donikovyski, D. Sattar, N. Sanni, M. Saranda, M. F.
1150	INICIIOIIS, S. J., PEITOIIE, S., PIIIA, I., POIIIKOWSKI, P., Sattar, N., Senni, M., Seronde, MF.,
1139	spinar, J., squire, I., Tauuei, S., Wanner, C., & Zannad, F. (2020). Cardiovascular and

1160	Renal Outcomes with Empagliflozin in Heart Failure. <i>New England Journal of Medicine</i> ,
1101	303(15), 1413-1424. <u>11(1ps://doi.org/10.1050/INEJM0d2022190</u>
1162 1163	Pao, A. C., Bhargava, A., Di Sole, F., Quigley, R., Shao, X., Wang, J., Thomas, S., Zhang, J., Shi,
1164 1165	and glucocorticoid-regulated kinase 2 in the regulation of Na+/H+ exchanger 3 in the
1166 1167	mammalian Kidney. <i>Am J Physiol Renal Physiol, 299</i> (6), F1496-1506. https://doi.org/10.1152/ajprenal.00075.2010
1168	Derly M.D. Marrie E.L. & Coholling I.D. (2007) Nov. II. auchanger (NUE) regulation in
1109 1170 1171	kidney proximal tubule. <i>Cellular and molecular life sciences : CMLS</i> , 72(11), 2061-2074.
1172	<u>11(1p3.//doi.org/10.100//300010-013-1040-0</u>
1173 1174	Pedersen, S. F., & Counillon, L. (2019, Oct 1). The SLC9A-C Mammalian Na(+)/H(+) Exchanger Family: Molecules, Mechanisms, and Physiology, <i>Physiol Rev.</i> 09(4), 2015-2113.
1175 1176	https://doi.org/10.1152/physrev.00028.2018
1177	Pessoa T. D. Campos I. C. G. Carraro-Lacroix I. Girardi A. C. C. & Malnic G. (2014)
1178 1179	Functional Role of Glucose Metabolism, Osmotic Stress, and Sodium-Glucose Cotransporter Isoform-Mediated Transport on
1180	Na<:sup>:+<:/sup>:/H<:sup>:+<:/sup>: Exchanger Isoform 3 Activity
1181	in the Renal Proximal Tubule. <i>Journal of the American Society of Nephroloay</i> , 25(9).
1182 1183	2028. <u>https://doi.org/10.1681/ASN.2013060588</u>
1184	Poulsen, S. B., Fenton, R. A., & Rieg, T. (2015). Sodium-glucose cotransport. <i>Current opinion in</i>
1185	nephrology and hypertension, 24(5), 463-469.
1186 1187	https://doi.org/10.1097/MNH.000000000000152
1188	Putney, L. K., Denker, S. P., & Barber, D. L. (2002). The changing face of the Na+/H+
1189	exchanger, NHE1: structure, regulation, and cellular actions. Annu Rev Pharmacol
1190 1191	<i>Toxicol,</i> 42, 527-552. <u>https://doi.org/10.1146/annurev.pharmtox.42.092001.143801</u>
1192	Qi, D., & Young, L. H. (2015). AMPK: energy sensor and survival mechanism in the ischemic
1195 1194	https://doi.org/10.1016/j.tem.2015.05.010
1195	Ramratnam, M., Sharma, R. K., D'Auria, S., Lee, S. L. Wang, D., Huang, X. Y., & Ahmad, F.
1197	(2014, Aug 4). Transgenic knockdown of cardiac sodium/glucose cotransporter 1
1198	(SGLT1) attenuates PRKAG2 cardiomyopathy, whereas transgenic overexpression of
1199	cardiac SGLT1 causes pathologic hypertrophy and dysfunction in mice. <i>I Am Heart</i>
1200 1201	Assoc, 3(4). https://doi.org/10.1161/jaha.114.000899
1202	Rasch, R. (1984, Jul). Tubular lesions in streptozotocin-diabetic rats. <i>Diabetologia</i> , 27(1), 32-37.
1203 1204	https://doi.org/10.1007/BF00253498
1205	Raskin, P. (2013, Jul). Sodium-glucose cotransporter inhibition: therapeutic potential for the
1206	treatment of type 2 diabetes mellitus. <i>Diabetes Metab Res Rev.</i> 20(5), 347-356.
1207	https://doi.org/10.1002/dmrr.2403
1208	
1209 1210	Reyes-Pardo, H., Bautista, R., Vargas-Robles, H., Rios, A., Sánchez, D., & Escalante, B. (2019). Role of sodium/glucose cotransporter inhibition on a rat model of angiotensin II-

1211	dependent kidney damage. <i>BMC nephrology</i> , 20(1), 292. Retrieved 2019/08//, from
1212	http://europepmc.org/abstract/MED/31375080
1213	https://doi.org/10.1186/s12882-019-1490-z
1214	https://europepmc.org/articles/PMC6679465
1215	https://europepmc.org/articles/PMC6679465?pdf=render
1216	
1217	Rossetti, L., Shulman, G. I., Zawalich, W., & DeFronzo, R. A. (1987). Effect of chronic
1218	hyperglycemia on in vivo insulin secretion in partially pancreatectomized rats. The
1219	Journal of clinical investigation, 80(4), 1037-1044. <u>https://doi.org/10.1172/JCl113157</u>
1220	
1221	Rossetti, L., Smith, D., Shulman, G. I., Papachristou, D., & DeFronzo, R. A. (1987, May).
1222	Correction of hyperglycemia with phlorizin normalizes tissue sensitivity to insulin in
1223	diabetic rats. The Journal of clinical investigation, 79(5), 1510-1515.
1224	https://doi.org/10.1172/jci112981
1225	
1226	Rotte, A., Pasham, V., Eichenmüller, M., Bhandaru, M., Föller, M., & Lang, F. (2010,
1227	2010/08/06/). Upregulation of Na+/H+ exchanger by the AMP-activated protein
1228	kinase. Biochemical and Biophysical Research Communications, 398(4), 677-682.
1229	https://doi.org/https://doi.org/10.1016/j.bbrc.2010.06.135
1230	
1231	Rupprecht, HJ., Dahl Jürgen, v., Terres, W., Seyfarth Karl, M., Richardt, G., Schultheiß, HP.,
1232	Buerke, M., Sheehan Florence, H., & Drexler, H. (2000, 2000/06/27). Cardioprotective
1233	Effects of the Na+/H+ Exchange Inhibitor Cariporide in Patients With Acute Anterior
1234	Myocardial Infarction Undergoing Direct PTCA. Circulation, 101(25), 2902-2908.
1235	https://doi.org/10.1161/01.CIR.101.25.2902
1236	
1237	Saeedi, P., Petersohn, I., Salpea, P., Malanda, B., Karuranga, S., Unwin, N., Colagiuri, S.,
1238	Guariguata, L., Motala, A. A., Ogurtsova, K., Shaw, J. E., Bright, D., & Williams, R.
1239	(2019, Nov). Global and regional diabetes prevalence estimates for 2019 and projections
1240	for 2030 and 2045: Results from the International Diabetes Federation Diabetes Atlas,
1241	9(th) edition. Diabetes Res Clin Pract, 157, 107843.
1242	https://doi.org/10.1016/j.diabres.2019.107843
1243	
1244	Sanidas, E. A., Papadopoulos, D. P., Hatziagelaki, E., Grassos, C., Velliou, M., & Barbetseas, J.
1245	(2020, Mar 13). Sodium Glucose Cotransporter 2 (SGLT2) Inhibitors Across the
1246	Spectrum of Hypertension. Am J Hypertens, 33(3), 207-213.
1247	https://doi.org/10.1093/ajh/hpz157
1248	
1249	Seferović, P. M., & Paulus, W. J. (2015). Clinical diabetic cardiomyopathy: a two-faced disease
1250	with restrictive and dilated phenotypes. European Heart Journal, 36(27), 1718-1727.
1251	https://doi.org/10.1093/eurheartj/ehv134
1252	
1253	Shi, Y., & Vanhoutte, P. M. (2009, Sep). Reactive oxygen-derived free radicals are key to the
1254	endothelial dysfunction of diabetes. <i>I Diabetes</i> , 1(3), 151-162.
1255	https://doi.org/10.1111/i.1753-0407.2009.00030.x
1256	
1257	Shiqiima, L. Yefremashvili, M., Luo, Z., Kureishi, Y., Takahashi, A., Tao, J., Rosenzweig, A
1258	Kahn, C. R., Abel, F. D., & Walsh, K. (2002, Oct 4) Akt signaling mediates postnatal
1259	heart growth in response to insulin and nutritional status. I Biol Chem. $277(AO)$ , $27670$ -
1260	37677. https://doi.org/10.1074/ibc.M204572200
1261	);-;;;-; <del><u>-</u></del>

1262	Silva dos Santos, D., Polidoro, J. Z., Borges-Júnior, F. A., & Girardi, A. C. C. (2019, 2020/02/01).
1263	Cardioprotection conferred by sodium-glucose cotransporter 2 inhibitors: a renal
1264	proximal tubule perspective. American Journal of Physiology-Cell Physiology, 318(2),
1265	C328-C336. <u>https://doi.org/10.1152/ajpcell.00275.2019</u>
1266	
1267	Song, J., Knepper, M. A., Verbalis, J. G., & Ecelbarger, C. A. (2003, Dec). Increased renal ENaC
1268	subunit and sodium transporter abundances in streptozotocin-induced type 1 diabetes.
1269	Am I Physiol Renal Physiol. 285(6). F1125-1137.
1270	https://doi.org/10.1152/aiprenal.001/3.2003
1271	
1272	Stapley W C & Chandler M P (2002 Apr) Energy metabolism in the normal and failing
1272	heart: notential for therapeutic interventions. <i>Heart Fail Rev.</i> 7(2) 115-120
1273	https://doi.org/10.1022/a:1015220422577
1274	$\frac{111293.77401.012774.1017520425777}{1010257425777}$
1275	Stoven S. Oolze M. Hanf A. Kröller Schön S. Kashani F. Poohani S. Welschof D. Konn
1270	M. Cädtal Armshmutt II. Via N. Li II. Cabula E. Lashman K. L. Mainanulti I.
1270	M., Goutel-Armbrust, U., Ala, N., LI, $\Pi$ ., Schulz, E., Lackner, K. J., Wojnowski, L.,
12/8	Bottari, S. P., Wenzel, P., Mayoux, E., Munzel, T., & Daiber, A. (2017, 2017/10/01/). The
12/9	SGLT2 inhibitor empagliflozin improves the primary diabetic complications in ZDF
1280	rats. Redox Biology, 13, 370-385.
1281	https://doi.org/https://doi.org/10.1016/j.redox.2017.06.009
1282	
1283	Stevens, V. A., Saad, S., Poronnik, P., Fenton-Lee, C. A., Polhill, T. S., & Pollock, C. A. (2008).
1284	The role of SGK-1 in angiotensin II-mediated sodium reabsorption in human proximal
1285	tubular cells. <i>Nephrology Dialysis Transplantation</i> , 23(6), 1834-1843.
1286	https://doi.org/10.1093/ndt/gfm923
1287	
1288	Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden,
1288 1289	Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., & Holman, R. R. (2000). Association of glycaemia with macrovascular
1288 1289 1290	Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., & Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective
1288 1289 1290 1291	Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., & Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i> , 321(7258), 405-412. https://doi.org/10.1136/bmj.321.7258.405
1288 1289 1290 1291 1292	Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., & Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i> , 321(7258), 405-412. <u>https://doi.org/10.1136/bmj.321.7258.405</u>
1288 1289 1290 1291 1292 1293	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i>, <i>321</i>(7258), 405-412. <u>https://doi.org/10.1136/bmj.321.7258.405</u></li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999,</li> </ul>
1288 1289 1290 1291 1292 1293 1294	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i>, <i>321</i>(7258), 405-412. https://doi.org/10.1136/bmj.321.7258.405</li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999, Jul 16). p90(RSK) is a serum-stimulated Na+/H+ exchanger isoform-1 kinase.</li> </ul>
1288 1289 1290 1291 1292 1293 1294 1295	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i>, 321(7258), 405-412. https://doi.org/10.1136/bmj.321.7258.405</li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999, Jul 16). p90(RSK) is a serum-stimulated Na+/H+ exchanger isoform-1 kinase. Regulatory phosphorylation of serine 703 of Na+/H+ exchanger isoform-1. <i>J Biol Chem</i>,</li> </ul>
1288 1289 1290 1291 1292 1293 1294 1295 1296	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i>, <i>321</i>(7258), 405-412. <u>https://doi.org/10.1136/bmj.321.7258.405</u></li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999, Jul 16). p90(RSK) is a serum-stimulated Na+/H+ exchanger isoform-1 kinase. Regulatory phosphorylation of serine 703 of Na+/H+ exchanger isoform-1. <i>J Biol Chem</i>, <i>274</i>(29). 20206-20214. https://doi.org/10.1074/ibc.274.20.20206</li> </ul>
1288 1289 1290 1291 1292 1293 1294 1295 1296 1297	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i>, <i>321</i>(7258), 405-412. <u>https://doi.org/10.1136/bmj.321.7258.405</u></li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999, Jul 16). p90(RSK) is a serum-stimulated Na+/H+ exchanger isoform-1 kinase. Regulatory phosphorylation of serine 703 of Na+/H+ exchanger isoform-1. <i>J Biol Chem</i>, <i>274</i>(29), 20206-20214. <u>https://doi.org/10.1074/jbc.274.29.20206</u></li> </ul>
1288 1289 1290 1291 1292 1293 1294 1295 1296 1297 1298	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i>, <i>321</i>(7258), 405-412. https://doi.org/10.1136/bmj.321.7258.405</li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999, Jul 16). p90(RSK) is a serum-stimulated Na+/H+ exchanger isoform-1 kinase. Regulatory phosphorylation of serine 703 of Na+/H+ exchanger isoform-1. <i>J Biol Chem</i>, <i>274</i>(29), 20206-20214. https://doi.org/10.1074/jbc.274.29.20206</li> <li>Takeishi, Y., Abe, L. Lee, I. D., Kawakatsu, H., Walsh, R. A., &amp; Berk, B. C. (1999, Dec 3-17).</li> </ul>
1288 1289 1290 1291 1292 1293 1294 1295 1296 1297 1298 1299	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i>, <i>321</i>(7258), 405-412. https://doi.org/10.1136/bmj.321.7258.405</li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999, Jul 16). p90(RSK) is a serum-stimulated Na+/H+ exchanger isoform-1 kinase. Regulatory phosphorylation of serine 703 of Na+/H+ exchanger isoform-1. <i>J Biol Chem</i>, <i>274</i>(29), 20206-20214. https://doi.org/10.1074/jbc.274.29.20206</li> <li>Takeishi, Y., Abe, J., Lee, J. D., Kawakatsu, H., Walsh, R. A., &amp; Berk, B. C. (1999, Dec 3-17). Differential regulation of poo ribosomal S6 kinase and big mitogen-activated protein</li> </ul>
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1288 1289 1290 1291 1292 1293 1294 1295 1296 1297 1298 1299 1300 1301	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i>, <i>321</i>(7258), 405-412. https://doi.org/10.1136/bmj.321.7258.405</li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999, Jul 16). p90(RSK) is a serum-stimulated Na+/H+ exchanger isoform-1 kinase. Regulatory phosphorylation of serine 703 of Na+/H+ exchanger isoform-1. <i>J Biol Chem</i>, <i>274</i>(29), 20206-20214. https://doi.org/10.1074/jbc.274.29.20206</li> <li>Takeishi, Y., Abe, J., Lee, J. D., Kawakatsu, H., Walsh, R. A., &amp; Berk, B. C. (1999, Dec 3-17). Differential regulation of p90 ribosomal S6 kinase and big mitogen-activated protein kinase 1 by ischemia/reperfusion and oxidative stress in perfused guinea pig hearts. <i>Circ Res</i>. <i>85</i>(12), u64-1172.</li> </ul>
1288 1289 1290 1291 1292 1293 1294 1295 1296 1297 1298 1299 1300 1301 1302	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i>, <i>321</i>(7258), 405-412. https://doi.org/10.1136/bmj.321.7258.405</li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999, Jul 16). p90(RSK) is a serum-stimulated Na+/H+ exchanger isoform-1 kinase. Regulatory phosphorylation of serine 703 of Na+/H+ exchanger isoform-1. <i>J Biol Chem</i>, <i>274</i>(29), 20206-20214. https://doi.org/10.1074/jbc.274.29.20206</li> <li>Takeishi, Y., Abe, J., Lee, J. D., Kawakatsu, H., Walsh, R. A., &amp; Berk, B. C. (1999, Dec 3-17). Differential regulation of p90 ribosomal S6 kinase and big mitogen-activated protein kinase 1 by ischemia/reperfusion and oxidative stress in perfused guinea pig hearts. <i>Circ Res</i>, <i>85</i>(12), 1164-1172. https://doi.org/10.1161/01.res.85.12.1164</li> </ul>
1288 1289 1290 1291 1292 1293 1294 1295 1296 1297 1298 1299 1300 1301 1302 1303	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ, 321</i>(7258), 405-412. https://doi.org/10.1136/bmj.321.7258.405</li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999, Jul 16). p90(RSK) is a serum-stimulated Na+/H+ exchanger isoform-1 kinase. Regulatory phosphorylation of serine 703 of Na+/H+ exchanger isoform-1. <i>J Biol Chem</i>, 274(29), 20206-20214. https://doi.org/10.1074/jbc.274.29.20206</li> <li>Takeishi, Y., Abe, J., Lee, J. D., Kawakatsu, H., Walsh, R. A., &amp; Berk, B. C. (1999, Dec 3-17). Differential regulation of p90 ribosomal S6 kinase and big mitogen-activated protein kinase 1 by ischemia/reperfusion and oxidative stress in perfused guinea pig hearts. <i>Circ Res</i>, 85(12), 1164-1172. https://doi.org/10.1161/01.res.85.12.1164</li> <li>Takeishi, Y. Huang, O. Abo, J. Cho, W. Lee, J. D. Kawakatsu, H. Hoit, B. D. Berk, B. C. *</li> </ul>
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1288 1289 1290 1291 1292 1293 1294 1295 1296 1297 1298 1299 1300 1301 1302 1303 1304 1305 1306	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i>, <i>321</i>(7258), 405-412. https://doi.org/10.1136/bmj.321.7258.405</li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999, Jul 16). p90(RSK) is a serum-stimulated Na+/H+ exchanger isoform-1 kinase. Regulatory phosphorylation of serine 703 of Na+/H+ exchanger isoform-1. <i>J Biol Chem</i>, <i>274</i>(29), 20206-20214. https://doi.org/10.1074/jbc.274.29.20206</li> <li>Takeishi, Y., Abe, J., Lee, J. D., Kawakatsu, H., Walsh, R. A., &amp; Berk, B. C. (1999, Dec 3-17). Differential regulation of p90 ribosomal S6 kinase and big mitogen-activated protein kinase 1 by ischemia/reperfusion and oxidative stress in perfused guinea pig hearts. <i>Circ Res</i>, <i>8</i>5(12), 1164-1172. https://doi.org/10.101/01.res.85.12.1164</li> <li>Takeishi, Y., Huang, Q., Abe, J., Che, W., Lee, J. D., Kawakatsu, H., Hoit, B. D., Berk, B. C., &amp; Walsh, R. A. (2002, Jan). Activation of mitogen-activated protein kinases and p90 ribosomal S6 kinase in failing human hearts with dilated cardiomyopathy. <i>Cardiovasc Res</i>, <i>53</i>(1), 131-137. https://doi.org/10.106/s0008-6363(01)00438-2</li> </ul>
1288 1289 1290 1291 1292 1293 1294 1295 1296 1297 1298 1299 1300 1301 1302 1303 1304 1305 1306 1307	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i>, <i>321</i>(7258), 405-412. https://doi.org/10.1136/bmj.321.7258.405</li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999, Jul 16). p90(RSK) is a serum-stimulated Na+/H+ exchanger isoform-1 kinase. Regulatory phosphorylation of serine 703 of Na+/H+ exchanger isoform-1. <i>J Biol Chem</i>, <i>274</i>(29), 20206-20214. https://doi.org/10.1074/jbc.274.29.20206</li> <li>Takeishi, Y., Abe, J., Lee, J. D., Kawakatsu, H., Walsh, R. A., &amp; Berk, B. C. (1999, Dec 3-17). Differential regulation of p90 ribosomal S6 kinase and big mitogen-activated protein kinase 1 by ischemia/reperfusion and oxidative stress in perfused guinea pig hearts. <i>Circ Res</i>, <i>85</i>(12), 1164-1172. https://doi.org/10.1161/01.res.85,12.1164</li> <li>Takeishi, Y., Huang, Q., Abe, J., Che, W., Lee, J. D., Kawakatsu, H., Hoit, B. D., Berk, B. C., &amp; Walsh, R. A. (2002, Jan). Activation of mitogen-activated protein kinases and p90 ribosomal S6 kinase in failing human hearts with dilated cardiomyopathy. <i>Cardiovasc Res</i>, <i>53</i>(1), 131-137. https://doi.org/10.106/s008-6363(01)00438-2</li> </ul>
1288 1289 1290 1291 1292 1293 1294 1295 1296 1297 1298 1299 1300 1301 1302 1303 1304 1305 1306 1307 1308	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i>, <i>321</i>(7258), 405-412. https://doi.org/10.1136/bmj.321.7258.405</li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999, Jul 16). p90(RSK) is a serum-stimulated Na+/H+ exchanger isoform-1 kinase. Regulatory phosphorylation of serine 703 of Na+/H+ exchanger isoform-1. <i>J Biol Chem</i>, <i>274</i>(29), 20206-20214. https://doi.org/10.1074/jbc.274.20.20206</li> <li>Takeishi, Y., Abe, J., Lee, J. D., Kawakatsu, H., Walsh, R. A., &amp; Berk, B. C. (1999, Dec 3-17). Differential regulation of p90 ribosomal S6 kinase and big mitogen-activated protein kinase 1 by ischemia/reperfusion and oxidative stress in perfused guinea pig hearts. <i>Circ Res</i>, <i>85</i>(12), 1164-1172. https://doi.org/10.1161/01.res.85.12.1164</li> <li>Takeishi, Y., Huang, Q., Abe, J., Che, W., Lee, J. D., Kawakatsu, H., Hoit, B. D., Berk, B. C., &amp; Walsh, R. A. (2002, Jan). Activation of mitogen-activated protein kinases and p90 ribosomal S6 kinase in failing human hearts with dilated cardiomyopathy. <i>Cardiovasc Res</i>, <i>53</i>(1), 131-137. https://doi.org/10.106/s0008-6363(01)00438-2</li> <li>Tanaka, A., Shimabukuro, M., Machii, N., Teragawa, H., Okada, Y., Shima, K. R., Takamura, T.,</li> </ul>
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1288 1289 1290 1291 1292 1293 1294 1295 1296 1297 1298 1299 1300 1301 1302 1303 1304 1305 1306 1307 1308 1309 1310	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ</i>, <i>321</i>(7258), 405-412. https://doi.org/10.1136/bmj.321.7258.405</li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999, Jul 16). p90(RSK) is a serum-stimulated Na+/H+ exchanger isoform-1 kinase. Regulatory phosphorylation of serine 703 of Na+/H+ exchanger isoform-1. <i>J Biol Chem</i>, <i>274</i>(29), 20206-20214. https://doi.org/10.1074/jbc.274.29.20206</li> <li>Takeishi, Y., Abe, J., Lee, J. D., Kawakatsu, H., Walsh, R. A., &amp; Berk, B. C. (1999, Dec 3-17). Differential regulation of p90 ribosomal S6 kinase and big mitogen-activated protein kinase 1 by ischemia/reperfusion and oxidative stress in perfused guinea pig hearts. <i>Circ Res</i>, <i>85</i>(12), 1164-1172. https://doi.org/10.1161/01.res.85,12.1164</li> <li>Takeishi, Y., Huang, Q., Abe, J., Che, W., Lee, J. D., Kawakatsu, H., Hoit, B. D., Berk, B. C., &amp; Walsh, R. A. (2002, Jan). Activation of mitogen-activated protein kinases and p90 ribosomal S6 kinase in failing human hearts with dilated cardiomyopathy. <i>Cardiovasc Res</i>, <i>53</i>(1), 131-137. https://doi.org/10.106/s0008-6363(01)00438-2</li> <li>Tanaka, A., Shimabukuro, M., Machii, N., Teragawa, H., Okada, Y., Shima, K. R., Takamura, T., Taguchi, I., Hisauchi, I., Toyoda, S., Matsuzawa, Y., Tomiyama, H., Yamaoka-Tojo, M., Yoshida, H., Sato, Y., Ikehara, Y., Ueda, S., Higashi, Y., &amp; Node, K. (2019). Effect of</li> </ul>
1288 1289 1290 1291 1292 1293 1294 1295 1296 1297 1298 1299 1300 1301 1302 1303 1304 1305 1306 1307 1308 1309 1310	<ul> <li>Stratton, I. M., Adler, A. I., Neil, H. A. W., Matthews, D. R., Manley, S. E., Cull, C. A., Hadden, D., Turner, R. C., &amp; Holman, R. R. (2000). Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. <i>BMJ, 321</i>(7258), 405-412. https://doi.org/10.1136/bmj.321.7258.405</li> <li>Takahashi, E., Abe, J., Gallis, B., Aebersold, R., Spring, D. J., Krebs, E. G., &amp; Berk, B. C. (1999, Jul 16). p90(RSK) is a serum-stimulated Na+/H+ exchanger isoform-1 kinase. Regulatory phosphorylation of serine 703 of Na+/H+ exchanger isoform-1. <i>J Biol Chem, 274</i>(29), 20206-20214. https://doi.org/10.1074/jbc.274.29.20206</li> <li>Takeishi, Y., Abe, J., Lee, J. D., Kawakatsu, H., Walsh, R. A., &amp; Berk, B. C. (1999, Dec 3-17). Differential regulation of p90 ribosomal S6 kinase and big mitogen-activated protein kinase 1 by ischemia/reperfusion and oxidative stress in perfused guinea pig hearts. <i>Circ Res, 85</i>(12), 1164-1172. https://doi.org/10.101/01.res.85.12.1164</li> <li>Takeishi, Y., Huang, Q., Abe, J., Che, W., Lee, J. D., Kawakatsu, H., Hoit, B. D., Berk, B. C., &amp; Walsh, R. A. (2002, Jan). Activation of mitogen-activated protein kinases and p90 ribosomal S6 kinase in failing human hearts with dilated cardiomyopathy. <i>Cardiovasc Res, 53</i>(1), 131-137. https://doi.org/10.1016/S0008-6363(01)00438-2</li> <li>Tanaka, A., Shimabukuro, M., Machii, N., Teragawa, H., Okada, Y., Shima, K. R., Takamura, T., Taguchi, I., Hisauchi, I., Toyoda, S., Matsuzawa, Y., Tomiyama, H., Yamaoka-Tojo, M., Yoshida, H., Sato, Y., Ikehara, Y., Ueda, S., Higashi, Y., &amp; Node, K. (2019). Effect of Empagliflozin on Endothelial Function in Patients With Type 2 Diabetes and</li> </ul>

1313	Controlled, Double-Blind EMBLEM Trial. <i>Diabetes Care, 42</i> (10), e159-e161.
1314	<u>https://doi.org/10.2337/dc19-1177</u>
1315	
1316	Tavakkolizadeh, A., Berger, U. V., Shen, K. R., Levitsky, L. L., Zinner, M. J., Hediger, M. A.,
1317	Ashley, S. W., Whang, E. E., & Rhoads, D. B. (2001, Feb). Diurnal rhythmicity in
1318	intestinal SGLT-1 function, V(max), and mRNA expression topography. <i>Am J Physiol</i>
1319	Gastrointest Liver Physiol, 280(2), G209-215.
1320	<u>https://doi.org/10.1152/ajpgi.2001.280.2.G209</u>
1321	
1322	Thomas, M. C., & Cherney, D. Z. I. (2018, Oct). The actions of SGLT2 inhibitors on
1323	metabolism, renal function and blood pressure. <i>Diabetologia</i> , 61(10), 2098-2107.
1324	<u>https://doi.org/10.1007/s00125-018-4669-0</u>
1325	
1326	Tojo, A., Onozato, M., Ha, H., Kurihara, H., Sakai, T., Goto, A., Fujita, T., & Endou, H. (2001,
1327	September 01). Reduced albumin reabsorption in the proximal tubule of early-stage
1328	diabetic rats [journal article]. <i>Histochemistry and Cell Biology, 116</i> (3), 269-276.
1329	https://doi.org/10.1007/s004180100317
1330	
1331	Tsuganezawa, H., Preisig, P. A., & Alpern, R. J. (1998). Dominant negative c-Src inhibits
1332	angiotensin II induced activation of NHE3 in OKP cells. <i>Kidney International</i> , 54(2),
1333	394-398. <u>https://doi.org/10.1046/j.1523-1755.1998.00029.x</u>
1334	
1335	Uthman, L., Baartscheer, A., Schumacher, C. A., Fiolet, J. W. T., Kuschma, M. C., Hollmann,
1336	M. W., Coronel, R., Weber, N. C., & Zuurbier, C. J. (2018). Direct Cardiac Actions of
1337	Sodium Glucose Cotransporter 2 Inhibitors Target Pathogenic Mechanisms Underlying
1338	Heart Failure in Diabetic Patients. Front Physiol, 9, 1575.
1339	https://doi.org/10.3389/fphys.2018.01575
1340	
1341	Vallés, P. G., Bocanegra, V., Gil Lorenzo, A., & Costantino, V. V. (2015). Physiological Functions
1342	and Regulation of the Na+/H+ Exchanger [NHE1] in Renal Tubule Epithelial Cells.
1343	Kidney Blood Press Res, 40(5), 452-466. https://doi.org/10.1159/000368521
1344	
1345	Vallon, V., & Sharma, K. (2010). Sodium-glucose transport: role in diabetes mellitus and
1346	potential clinical implications. <i>Current opinion in nephrology and hypertension</i> , 19(5).
1347	425-431, https://doi.org/10.1097/MNH.ob013e32833beco6
1348	
1349	Wakabavashi, S., Hisamitsu, T., & Nakamura, T. Y. (2013). Regulation of the cardiac Na+/H+
1350	exchanger in health and disease. I Mol Cell Cardiol. 61, 68-76.
1351	https://doi.org/10.1016/i.vimcc.2013.02.007
1352	
1353	Walsh K (2006 May 2) Akt signaling and growth of the heart Circulation, $u_2(17)$ , 2022-2024
1354	https://doi.org/10.1161/circulationaha.106.615138
1355	<u>Intepsi//donoig/iomoi/enconditionalanoolorjijo</u>
1356	Wang D. Zhang H. Lang F. & Yun C. C. (2007 Jan). Acute activation of NHF2 by
1357	devamethasone correlates with activation of SGK1 and requires a functional
1358	ducocorticoid receptor Am I Physiol Cell Physiol 202(1) C206-404
1359	$\frac{1}{1000} = \frac{1}{1000} = 1$
1360	<u>1102001/10015/10011/2/0/PCC11.00343.2000</u>
1361	Wang Y Meyer I W Ashraf M & Shull G F (2002 Oct 17) Mice with a null mutation in
1362	the NHF1 Na+-H+ exchanger are resistant to cardiac ischemia-renerfusion injury Circ
1363	Res $\alpha_2(8)$ 776-782 https://doi.org/10.1161/01.Res.0000004746.24774.Dc
1000	10, 7)(0), 7/0 702. <u>http://tonoig/iointoi/ointes/oo00094/40.24/74.De</u>

1364	
1365	Wiviott, S. D., Raz, I., Bonaca, M. P., Mosenzon, O., Kato, E. T., Cahn, A., Silverman, M. G.,
1366	Zelniker, T. A., Kuder, J. F., Murphy, S. A., Bhatt, D. L., Leiter, L. A., McGuire, D. K.,
1367	Wilding, J. P. H., Ruff, C. T., Gause-Nilsson, I. A. M., Fredriksson, M., Johansson, P. A.,
1368	Langkilde, AM., & Sabatine, M. S. (2018). Dapagliflozin and Cardiovascular Outcomes
1369	in Type 2 Diabetes. New England Journal of Medicine, 380(4), 347-357.
1370	https://doi.org/10.1056/NEIM0a1812389
1371	
1372	Wright, E. M., Loo, D. D., & Hiravama, B. A. (2011, Apr). Biology of human sodium glucose
1373	transporters. <i>Physiol Rev, 91</i> (2), 733-794. <u>https://doi.org/10.1152/physrev.00055.2009</u>
1374	
1375	Yamaguchi, N., Chakraborty, A., Pasek, D. A., Molkentin, J. D., & Meissner, G. (2011, Jun).
1376	Dysfunctional ryanodine receptor and cardiac hypertrophy: role of signaling
1377	molecules. Am J Physiol Heart Circ Physiol, 300(6), H2187-2195.
1378	<u>https://doi.org/10.1152/ajpheart.00719.2010</u>
1379	
1380	Ye, Y., Jia, X., Bajaj, M., & Birnbaum, Y. (2018, Dec). Dapagliflozin Attenuates Na(+)/H(+)
1381	Exchanger-1 in Cardiofibroblasts via AMPK Activation. Cardiovasc Drugs Ther, 32(6),
1382	553-558. <u>https://doi.org/10.1007/s10557-018-6837-3</u>
1383	
1384	Yoshii, A., Nagoshi, T., Kashiwagi, Y., Kimura, H., Tanaka, Y., Oi, Y., Ito, K., Yoshino, T.,
1385	Tanaka, T. D., & Yoshimura, M. (2019). Cardiac ischemia-reperfusion injury under
1386	insulin-resistant conditions: SGLT1 but not SGLT2 plays a compensatory protective
1387	role in diet-induced obesity. <i>Cardiovascular diabetoloay</i> , 18(1), 85-85.
1388	https://doi.org/10.1186/s12933-019-0889-v
1389	
1390	Zeymer, U., Survapranata, H., Monassier, J. P., Opolski, G., Davies, J., Rasmanis, G., Linssen,
1391	G., Tebbe, U., Schröder, R., Tiemann, R., Machnig, T., & Neuhaus, K. L. (2001, Nov 15).
1392	The Na(+)/H(+) exchange inhibitor eniporide as an adjunct to early reperfusion
1393	therapy for acute myocardial infarction. Results of the evaluation of the safety and
1394	cardioprotective effects of eniporide in acute myocardial infarction (ESCAMI) trial <i>I</i>
1395	Am Coll Cardiol. 38(6) 1644-1650 https://doi.org/10.1016/s0735-1007(01)01608-4
1396	1111 con cululo, jo(0), 1044 1030. <u>https://doi.org/101010/30/31 103/(0/010100 4</u>
1397	7hao W. Katzmarzyk P.T. Horswell R. Wang V. Johnson J. & Hu. G. (2014). HbAic and
1398	beart failure risk among diabetic patients. The Journal of clinical endocrinology and
1300	metabolism oo(2) E262-E267 https://doi.org/10.1210/ic.2012-2225
1400	metubolism, 99(2), 1203 1207. <u>https://doi.org/10.1210/je.2013-3325</u>
1400	Zhou O Kesteven S Wu I Aidem P Cowar M Cramlich M Feneley M P & Harvey
1/02	R. D. (2015) Pressure Overload by Transverse Aartic Constriction Induces Maladaptive
1402	Hypertrophy in a Titin Truncated Mouse Model Piemed Pag Int. 2015, 16254
1403	https://doi.org/10.1155/2015/16256.t
1404	<u>11(1ps://doi.org/10.1155/2015/103504</u>
1405	Zinman B. Wanner C. Lachin I.M. Fitchett D. Bluhmki F. Hantel S. Mattheus M.
1400	Devine T. Johanson O. F. Woorlo H. J. Broadl H. C. & Ingueshi S. F. (2015)
1/00/	Empagliflorin Cardiovacular Outcomes and Mortality in Type a Diabates New
1400	Empagimozin, Carulovascular Outcomes, and Mortality in Type 2 Diabetes. New
1409	England Journal of Medicine, 3/3(22), 2117-2128. <u>https://doi.org/10.1056/INEJM0a1504720</u>
1410	