Meta-analysis of transcriptome reveals key genes relating to oil quality in olive

AliAkbar Asadi¹, Vahid Shariati¹, Soraya Mousavi¹, Roberto Mariotti¹, and Mehdi Hosseini Mazinani¹

¹Affiliation not available

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Abstract

Background: Olive oil contains monounsaturated oleic acid up to 83% and phenolic compounds, making it an excellent source of fat. Due to its economic importance, the quantity and quality of olive oil should be improved in parallel with the international standards. In this study, we analyzed the raw RNA-seq data with a meta-analysis approach to identify important genes and their metabolic pathways involved in olive oil quality.

Results: A deep search of RNA-seq published data shed light on thirty experiments associated with the olive transcriptome, four of these proved to be ideal for meta-analysis. Meta-analysis confirmed the genes identified in previous studies and released new genes, which were not identified before. According to the IDR index, the meta-analysis had good power to identify new differentially expressed genes. The key genes were investigated in the metabolic pathways and were grouped into four classes based on the biosynthetic cycle of fatty acids and factors that affect oil quality. Galactose metabolism, glycolysis pathway, pyruvate metabolism, fatty acid biosynthesis, glycerolipid metabolism, and terpenoid backbone biosynthesis were the main pathways in olive oil quality. In galactose metabolism, raffinose is a suitable source of carbon along with other available sources for carbon in fruit development. The results showed that the biosynthesis of acetyl-CoA in glycolysis and pyruvate metabolism is a stable pathway to begin the biosynthesis of fatty acids. Key genes in oleic acid production as an indicator of oil quality and critical genes that played an important role in production of triacylglycerols were identified in different developmental stages. In the minor compound, the terpenoid backbone biosynthesis was investigated and important enzymes were identified as an interconnected network that produces important precursors for the synthesis of a monoterpene, diterpene, triterpene, and sesquiterpene biosynthesis.

Conclusions: The results of the current investigation can produce functional data related to the quality of olive oil and would be a useful step in reducing the time of cultivar screening by developing gene specific markers in olive breeding programs, releasing also new genes that could be applied in the genome editing approach.

Introduction

Olive oil is a particularly important product because of its fatty acids and phenolic compounds, which are mainly responsible for the beneficial health aspects. Among these compound, high oleic acid content and the presence of minor bioactive compounds are the reason for its attribution as the healthiest among all vegetable oils (Guclu et al., 2020; Lozano-Castellón et al., 2020; Yubero-Serrano et al., 2019). The International Olive Council (IOC) has also stated that oils with the highest levels of oleic acid are the most valuable nutritional products (IOC, 2015). Currently, the cultivation areas and oil production increased in the world but only a few cultivars can yield consistently in the new environmental conditions and often changing negatively their quality profiles (Borges et al., 2017; García-Inza et al., 2016; Torres et al., 2017). Therefore, according to the economic importance of olive oil, the quantity and quality of oil should be improved in parallel, based on international standards.

Several studies have noted that the main factors that influence olive oil quality are genotype, climatic and agronomic conditions, edaphic factors, and the technological method applied for oil extraction. Among these factors, genotype has a preponderant influence (Ambra et al., 2017; Baiano et al., 2013; Beltrán et al., 2016; de la Rosa et al., 2016; Mele et al., 2018; Miho et al., 2021; Mikrou et al., 2020; Pérez et al., 2018; Rugini et al., 2016). Besides studies show that 70% of the observed diversity in terms of fatty acid composition, phenolic compounds, bitterness or taste, and oil stability is genetically influenced (Mousavi et al., 2019, 2022; Parvini et al., 2015; Riachy et al., 2019).

One of the most important goals of RNA-seq experiments is to investigate changes in gene expression profiles under two or more different experimental conditions. The most of the RNA-seq studies performed in olives were related to the study of biotic and abiotic stresses (Grasso et al., 2017; Nissim, Shlosberg, et al., 2020), micro-RNA identification (Yanik et al., 2013), fruit developmentalstages (Alagna et al., 2009, 2016; Galla et al., 2009; Guodong et al., 2019), and cold acclimation (De La O Leyva-Pérez et al., 2015; Guerra et al., 2015). Recently, some studies have focused on the role of environmental stresses such as high temperature and the altitude of cultivated areas on oil content and its quality using RNA-seq technique, but a limited number of these studies were directly related to the evaluation of oil quality (Bruno et al., 2019; Nissim, Shloberg, et al., 2020; Nissim, Shlosberg, et al., 2020). In a study performed by Galla et al. (2009), suppression subtractive hybridization (SSH) was used to isolate and identify a large set of genes that were differentially expressed at three different stages along olive fruit development. In another study conducted by Alagna et al., 2009, differentially expressed genes involved in the metabolism of phenol and fatty acids at different stages of olive fruit development were identified. Moreover, in 2013, the olive cultivars' transcripts were used for de novo assembly and functional annotation (Rodríguez, 2013). In a study conducted by Parra et al. (2013), the transcriptional regulation of the ripening process and activation of abscission zone were detected by RNA-seq. In 2016, the genome of Farga was sequenced and its annotation was identified by RNA-seq of leaf, root, and fruit samples (Cruz et al., 2016). Considering the importance of phenols in olive, in 2016, de novo transcriptome assembly was reported in olive fruit at different development stages and transcripts involved in flavonoid and anthocyanin pathways were identified (Iaria et al., 2016). In another study (Unver et al., 2017), the wild olive genome was sequenced and transcriptome analysis was performed to identify genes involved in oil biosynthesis. Recently, transcripts of all the enzymes in the biosynthetic pathway of tyrosol, hydroxytyrosol and secologanin, oleuropein's precursor, were identified by the RNA-seq method in Koroneiki cultivar (Mougiou et al., 2018). Furthermore, in 2019, targeted metabolome, Pachio ISOseq transcriptome, and Illumina RNA-seq transcriptome were combined to investigate the relationship between phenols biosynthesis and differentially expressed genes during olive fruit development (Guodong et al., 2019).

Technical variation in different experiments could affect the reproducibility of the research. Moreover, due to the cost of sequencing, RNA-seq experiments are mainly performed in a limited number of biological replicates, reducing the statistical power and the ability to detect and validate differences in gene expression. Accordingly, one of the most effective ways to improve reproducibility is to use multiple datasets through meta-analysis(Keel et al., 2018). Therefore, re-analyzing existing data derived from several independent experiments can reveal new information and evaluate the most reliable key genes in a certain biosynthetic pathway. Meta-analysis of RNA-seq data can increase the speed of production of functional data related to the quality of olive oil and produce useful information. This study was conducted to analyze RNA-seq data obtained from multiple studies by meta-analysis approach to validate and identify key genes involved in the main metabolic pathways of oil quality.

Results

The SRA and literature searches results showed that there were thirty experiments associated with the olive transcriptome. By applying the filtration and quality control reported in the methods section, four experiments had the ideal conditions to enter in our meta-analysis. The meta-analysis was performed to compare growth stages in pairs (C1, C2, and C3) and the results of each comparison were shown as an independent Venn diagram (Figure 1). The meta-analysis individuates 1472 differential expressed genes in C1 comparison from which, 155 differential expressed genes were identified for the first time in the present study (Figure 1A). The C2 comparison has identified 5175 differential expressed genes (Figure 1B), among them, 473 differential expressed genes have never been reported in the previous studies. The PRJNA260808 had only two developmental stages, S2 and S3, so only the C3 comparison was considered in this experiment. The results of C3 comparison identified 1034 differential expressed genes (Figure 1C), and in addition, 241 of them were identified for the first time.

The ratio of the identified differentially expressed genes in the meta-analysis to the total identified differential expressed genes were calculated as Integration-driven Discovery Rate (IDR) for each comparison. The IDR for C1 was 10.53 and in C2 and C3 were 9.14 and 23.13, respectively.



Figure 1: The Venn diagrams of growth stages comparison in the meta-analysis

In order to identify essential and critical metabolic pathways in oil biosynthesis and quality compounds, two strategies were performed according to literature review (first step) and metabolic pathways (second step) described in Unver et al., (2017). In the first strategy, all the identified differential expressed genes in the meta-analysis were used for pathway enrichment and then important pathways selected. In the second strategy, to select critical metabolic pathways in oil quality, differential expression genes obtained from the meta-analysis were blasted against 2327 effective genes in oil biosynthesis (Unver et al., 2017). The two steps led to enriching the pathways identified in each strategy. These steps were done separately for each comparison and selected common pathways between overall comparisons, pair comparisons, and specific to each comparison for the up and down-regulated genes (Table 1 and 2). Sixteen metabolic pathways were common between all growth stages for the up-regulated genes (Table 1). The pathways of fatty acids biosynthesis and their elongation were in common between C1 and C2. Furthermore, the most important individuated metabolic pathways were pentose phosphate, pyruvate metabolism, glycolysis/gluconeogenesis, fatty acid biosynthesis, fatty acid elongation, and biotin metabolism. The comparisons' results also showed that the terpenoid compounds biosynthesis pathways were specific to C2 including limonene and pinene degradation, diterpenoid biosynthesis, and monoterpenoid biosynthesis (Table 1). In the metabolic pathways containing down-regulated genes, 23 metabolic pathways were common between all developmental stages (Table 2).

The identified metabolic pathways were grouped according to the biosynthetic cycle of fatty acids and factors affecting oil quality, to provide more details about the most important identified pathways. In the study of oil biosynthesis, attention should be paid to the three key stages of carbon supply in the skeleton of triacylglycerols including the production of fatty acids, the assembly of fatty acids, and finally the formation of complex lipids (Salas et al., 2013).

- 1. Pathways related to carbon source: In the first group, there are metabolic pathways that supply carbon for production of fatty acids, and the pathway of galactose metabolism has been identified in this group (Additional file 1). There are several sources of carbon in olive tree (Salas et al., 2013), which are A) Leaves photosynthesis (transfer of photosynthetic products from leaves to other organs) B) Raffinose family oligosaccharides (making of sugar alcohols and oligosaccharides such as mannitol, raffinose, and stachyose along with other photosynthetic materials in separate reactions), and C) Fruit photosynthesis. In olive, biosynthesis of mannitol and sorbitol, which are sugar alcohol, is present, and this pathway was also observed in the *Apiaceae* family (Sánchez & Harwood, 2002). In addition, the biosynthesis of oligosaccharides such as stachyose and raffinose, which are found in the legumes family, was also observed in olives (Salas et al., 2013). Therefore, it is possible to confirm that making sugar alcohols and oligosaccharides, as observed in other plant families, can be a reliable source of carbon also in olives.
- 2. Pathways related to Acetyl-CoA: In the second group, the presence of acetyl-CoA along with two enzymes acetyl-CoA carboxylase (ACCase) and fatty acid synthase (FAS) is necessary to start the biosynthesis of fatty acids (Salas et al., 2013). It was stated that a stable and rapid pathway for the supply of acetyl-CoA is glycolysis and pentose phosphate pathways using pyruvate by the activity of the enzyme pyruvate dehydrogenase (AID in 2012). In the present study, the key metabolic pathways identified for acetyl-CoA biosynthesis were glycolysis (Additional file 2) and pyruvate metabolism (Additional file 3).
- 3. Pathways related to fatty acids: In the third group, biosynthesis of the carbon chain of fatty acids, the key metabolic pathways of fatty acid biosynthesis, fatty acid elongation, and biotin metabolism were individuated in the present study. Metabolic pathway glycerolipid metabolism was identified as a key metabolic pathway in the assembling of fatty acids and the formation of complex lipids. In addition, two metabolic pathways alpha-linolenic acid metabolism and linoleic acid metabolism were identified as important pathways in the production of by-products of fatty acid biosynthesis.
- 4. Pathways related to non-fatty acids components (minor compound): The fourth group was related to metabolic pathways that have a direct impact on the quality of the oil and its taste. In this group, six key metabolic pathways were observed: terpenoid backbone biosynthesis, phenylpropanoid biosynthesis, flavonoid biosynthesis, limonene and pinene degradation, monoterpenoid biosynthesis, and carotenoid biosynthesis.

The detected metabolic pathways affecting the quality of oil in this study have been investigated independently, and the genes affecting each metabolic pathway have been identified and introduced (Additional file

4).

In olive, breeding programs last about thirty years on average (Lavee et al., 2014; Rallo et al., 2018), while the timing for the selection of new cultivars in other fruit crops has been greatly reduced, also by the application of new efficient genomic tools (Biscarini et al., 2017; Cai et al., 2019; Laurens et al., 2018). In cultivated olives (*Olea europaea* subsp. *europaea* var. *europaea*), the crossbreeding activities have been delayed by the particularly long generation time (Santos-Antunes et al., 2005), the extended juvenile phase and the high demanding nursery practices. This work provides the best candidate genes to construct markers for a fast and reliable genotyping of olive cultivars for their oil quality, offering great opportunities to rapidly screen and planning inter-varietal crosses, and reducing the time for seedling selection. Furthermore, the newly detected candidate genes could be applied as a source for interested traits in genome editing approaches.

The IDR index demonstrated that meta-analysis has good power to identify new differentially expressed genes. Therefore, the low value of IDR indicates a subtly increase in the power of meta-analysis compared to the results obtained in the previously published independent studies (Hong & Breitling, 2008). In addition, the decrease in the number of new differentially expressed genes in the meta-analysis compared to the total genes that were significantly different in comparison between the two developmental stages indicated that the meta-analysis was more efficient than independent studies and provided solid results.

Conclusions

According to the economic importance of olive oil, the quantity and quality of oil should be improved in parallel based on international standards in commercial cultivars. Technical variation in different experiments and limited number of biological replicates in RNA-seq researches could affect the reproducibility and the statistical power of the experiments. Meta-analysis of RNA-seq data can increase the speed of production of functional data related to the quality of olive oil and produce useful information. The present work would be an effective and useful step in reducing the time of cultivar screening by developing key genes relating to oil quality in olive breeding programs. The IDR index indicated the meta-analysis had good power to identify new differentially expressed genes and identified 155, 473, and 241 differential expressed genes for the first time respectively in the C1, C2, and C3. The identified key genes were investigated in the metabolic pathways related to oil quality and the identified metabolic pathways were also grouped into four groups according to the pathway of oil biosynthesis and the factors affecting its quality. The metabolic pathways that affecting the quality of oil have been examined independently confirming the earlier published results through meta-analysis and contributing to the reproducibility of the results. The galactose metabolism pathway can be further investigated as a source of carbon supply in olive fruit, and due to the increase in the expression of its genes in the S3, it can help to supply carbon in the final stage of growth when the fruit photosynthesis is reduced. Moreover, the production of acetyl-CoA in various ways in different growth stage shows the high efficiency of olive in the production of fatty acids. In the fatty acid biosynthesis pathway FabF and SAD have a critical role in oil quality and second identified pathway is more suitable for production of high oleic acid. Furthermore, the production of TAG by the PDAT probably occurred in the mesocarp and played an important role in the production of TAG and oil in the mesocarp. Also, the results of present study showed biosynthesis of phenolic compounds in olives in terms of growth stages might be in the opposite of the biosynthesis of fatty acids and there are the highest amount of phenolic compounds after fruit formation, while the biosynthesis of oil is at its lowest. The identified and confirmed genes by metaanalysis approach provided a suitable dataset to draw on for future studies in different scientific sectors. The present research can also play a role in cultivar selection by applying potential molecular markers developed in the here individuated key genes, reducing the duration of breeding programs, and helping the genome editing by introducing key and essential genes for the development of healthy compounds.

Authors' contributions

AA.A. collected data, performed all steps of the meta-analysis of RNA-seq, wrote the main draft, and done

final revision on the manuscript. V.S. was conceived and designed the project, guide and confirmed the data analysis, and revised and approved the final manuscript. S.M and R.M provided critical advices on the manuscript and revised the manuscript. M.HM revised the final manuscript. The research was co-supervised by M.HM. All authors have read and approved the final manuscript.

References

Aharoni, A., Jongsma, M. A., Kim, T. Y., Ri, M. B., Giri, A. P., Verstappen, F. W. A., Schwab, W., & Bouwmeester, H. J. (2006). Metabolic engineering of terpenoid biosynthesis in plants. *Phytochemistry Reviews*, 5(1), 49–58. https://doi.org/10.1007/s11101-005-3747-3

AID, F. (2012). Plant Lipid Metabolism. *Intech*, 13. http://dx.doi.org/10.1039/C7RA00172J%0Ahttps://www.intechopen.co/biometric-technologies/liveness-detection-in-biometrics%0Ahttp://dx.doi.org/10.1016/j.colsurfa.2011.12.014

Alagna, F., Cirilli, M., Galla, G., Carbone, F., Daddiego, L., Facella, P., Lopez, L., Colao, C., Mariotti, R., Cultrera, N., Rossi, M., Barcaccia, G., Baldoni, L., Muleo, R., & Perrotta, G. (2016). Transcript analysis and regulative events during flower development in olive (Olea europaea L.). *PLoS ONE*, 11(4), 1–32. https://doi.org/10.1371/journal.pone.0152943

Alagna, F., D'Agostino, N., Torchia, L., Servili, M., Rao, R., Pietrella, M., Giuliano, G., Chiusano, M. L., Baldoni, L., & Perrotta, G. (2009). Comparative 454 pyrosequencing of transcripts from two olive genotypes during fruit development. *BMC Genomics*, 10, 399. https://doi.org/10.1186/1471-2164-10-399

Alagna, F., Mariotti, R., Panara, F., Caporali, S., Urbani, S., Veneziani, G., Esposto, S., Taticchi, A., Rosati, A., Rao, R., Perrotta, G., Servili, M., & Baldoni, L. (2012). Olive phenolic compounds: metabolic and transcriptional profiling during fruit development. *BMC Plant Biology*, 12(1), 162. https://doi.org/10.1186/1471-2229-12-162

Ambra, R., Natella, F., Lucchetti, S., Forte, V., & Pastore, G. (2017). α-Tocopherol, β-carotene, lutein, squalene and secoiridoids in seven monocultivar Italian extra-virgin olive oils. *International Journal of Food Sciences and Nutrition*, 68(5), 538–545. https://doi.org/10.1080/09637486.2016.1265099

Amiripour, M., Sadat Noori, S. A., Shariati, V., & Soltani Howyzeh, M. (2019). Transcriptome analysis of Ajowan (Trachyspermum ammi L.) inflorescence. *Journal of Plant Biochemistry and Biotechnology*, 28(4), 496–508. https://doi.org/10.1007/s13562-019-00504-4

Baiano, A., Terracone, C., Viggiani, I., & Nobile, M. A. Del. (2013). Effects of Cultivars and Location on Quality, Phenolic Content and Antioxidant Activity of Extra-Virgin Olive Oils. *Journal of the American Oil Chemists' Society*, 90(1), 103–111. https://doi.org/10.1007/s11746-012-2141-8

Bates, P. D., & Browse, J. (2012). The Significance of Different Diacylgycerol Synthesis Pathways on Plant Oil Composition and Bioengineering. *Frontiers in Plant Science*, 3. https://doi.org/10.3389/fpls.2012.00147

Beltrán, G., Bucheli, M. E., Aguilera, M. P., Belaj, A., & Jimenez, A. (2016). Squalene in virgin olive oil: Screening of variability in olive cultivars. *European Journal of Lipid Science and Technology*, 118(8), 1250–1253. https://doi.org/10.1002/ejlt.201500295

Biscarini, F., Nazzicari, N., Bink, M., Arús, P., Aranzana, M. J., Verde, I., Micali, S., Pascal, T., Quilot-Turion, B., Lambert, P., da Silva Linge, C., Pacheco, I., Bassi, D., Stella, A., & Rossini, L. (2017). Genomeenabled predictions for fruit weight and quality from repeated records in European peach progenies. *BMC Genomics*, 18(1), 432. https://doi.org/10.1186/s12864-017-3781-8

Borges, T. H., Pereira, J. A., Cabrera-Vique, C., Lara, L., Oliveira, A. F., & Seiquer, I. (2017). Characterization of Arbequina virgin olive oils produced in different regions of Brazil and Spain: Phy-

sicochemical properties, oxidative stability and fatty acid profile. *Food Chemistry*, 215, 454–462. htt-ps://doi.org/10.1016/j.foodchem.2016.07.162

Bruno, L., Picardi, E., Pacenza, M., Chiappetta, A., Muto, A., Gagliardi, O., Muzzalupo, I., Pesole, G., & Bitonti, M. B. (2019). Changes in gene expression and metabolic profile of drupes of Olea europaea L. cv Carolea in relation to maturation stage and cultivation area. 1–17.

Cai, L., Quero-García, J., Barreneche, T., Dirlewanger, E., Saski, C., & Iezzoni, A. (2019). A fruit firmness QTL identified on linkage group 4 in sweet cherry (Prunus avium L.) is associated with domesticated and bred germplasm. *Scientific Reports*, 9(1), 5008. https://doi.org/10.1038/s41598-019-41484-8

Castillo, E. M., de Lumen, B. O., Reyes, P. S., & de Lumen, H. Z. (1990). Raffinose Synthase and Galactinol Synthase in Developing Seeds and Leaves of Legumes. *Journal of Agricultural and Food Chemistry*, 38(2), 351–355. https://doi.org/10.1021/jf00092a003

Conde, C., Delrot, S., & Gerós, H. (2008). Physiological, biochemical and molecular changes occurring during olive development and ripening. *Journal of Plant Physiology*, 165(15), 1545–1562. https://doi.org/10.1016/j.jplph.2008.04.018

Cruz, F., Julca, I., Gómez-Garrido, J., Loska, D., Marcet-Houben, M., Cano, E., Galán, B., Frias, L., Ribeca, P., Derdak, S., Gut, M., Sánchez-Fernández, M., García, J. L., Gut, I. G., Vargas, P., Alioto, T. S., & Gabaldón, T. (2016). Genome sequence of the olive tree, Olea europaea. *GigaScience*, 5(1). https://doi.org/10.1186/s13742-016-0134-5

Dar, A. A., Choudhury, A. R., Kancharla, P. K., & Arumugam, N. (2017). The FAD2 gene in plants: Occurrence, regulation, and role. *Frontiers in Plant Science*, 8(October), 1–16. htt-ps://doi.org/10.3389/fpls.2017.01789

de Jaeger, L., Springer, J., Wolbert, E. J. H., Martens, D. E., Eggink, G., & Wijffels, R. H. (2017). Gene silencing of stearoyl-ACP desaturase enhances the stearic acid content in Chlamydomonas reinhardtii. *Bioresource Technology*, 245, 1616–1626. https://doi.org/10.1016/j.biortech.2017.06.128

De La O Leyva-Pérez, M., Valverde-Corredor, A., Valderrama, R., Jiménez-Ruiz, J., Muñoz-Merida, A., Trelles, O., Barroso, J. B., Mercado-Blanco, J., & Luque, F. (2015). Early and delayed long-term transcriptional changes and short-term transient responses during cold acclimation in olive leaves. *DNA Research*, 22(1), 1–11. https://doi.org/10.1093/dnares/dsu033

de la Rosa, R., Arias-Calderon, R., Velasco, L., & Leon, L. (2016). Early selection for oil quality components in olive breeding progenies. *European Journal of Lipid Science and Technology*, 118(8), 1160–1167. https://doi.org/10.1002/ejlt.201500425

Foka, I. C. K., Ketehouli, T., Zhou, Y., Li, X. W., Wang, F. W., & Li, H. (2020). The emerging roles of diacylglycerol kinase (DGK) in plant stress tolerance, growth, and development. *Agronomy*, 10(9), 1–26. https://doi.org/10.3390/agronomy10091375

Galla, G., Barcaccia, G., Ramina, A., Collani, S., Alagna, F., Baldoni, L., Cultrera, N. G. M., Martinelli, F., Sebastiani, L., & Tonutti, P. (2009). Computational annotation of genes differentially expressed along olive fruit development. *BMC Plant Biology*, 9, 1–17. https://doi.org/10.1186/1471-2229-9-128

Garcia-Inza, G. P., Castro, D. N., Hall, A. J., & Rousseaux, M. C. (2016). Opposite oleic acid responses to temperature in oils from the seed and mesocarp of the olive fruit. *European Journal of Agronomy*, 76, 138–147. https://doi.org/10.1016/j.eja.2016.03.003

Grasso, F., Coppola, M., Carbone, F., Baldoni, L., Alagna, F., Perrotta, G., Perez-Pulido, A. J., Garonna, A., Facella, P., Daddiego, L., Lopez, L., Vitiello, A., Rao, R., & Corrado, G. (2017). The transcriptional response to the olive fruit fly (Bactrocera oleae) reveals extended differences between tolerant and susceptible olive (Olea europaea L.) varieties. *PLoS ONE*, 12(8), 1–19. https://doi.org/10.1371/journal.pone.0183050

Guclu, G., Kelebek, H., & Selli, S. (2020). Antioxidant activity in olive oils. In V. Preedy & R. Watson (Eds.), *Olives and Olive Oil in Health and Disease Prevention* (Second, pp. 313–325). Academic Press. https://doi.org/10.1016/B978-0-12-819528-4.00031-6

Guerin, C., Joet, T., Serret, J., Lashermes, P., Vaissayre, V., Agbessi, M. D. T., Beule, T., Severac, D., Amblard, P., Tregear, J., Durand-Gasselin, T., Morcillo, F., & Dussert, S. (2016). Gene coexpression network analysis of oil biosynthesis in an interspecific backcross of oil palm. *The Plant Journal*, 87(5), 423–441. https://doi.org/10.1111/tpj.13208

Guerra, D., Lamontanara, A., Bagnaresi, P., Orru, L., Rizza, F., Zelasco, S., Beghe, D., Ganino, T., Pagani, D., Cattivelli, L., & Mazzucotelli, E. (2015). Transcriptome changes associated with cold acclimation in leaves of olive tree (Olea europaea L.). *Tree Genetics and Genomes*, 11(6). https://doi.org/10.1007/s11295-015-0939-x

Guodong, R., Jianguo, Z., Xiaoxia, L., & Ying, L. (2019). Identification of putative genes for polyphenol biosynthesis in olive fruits and leaves using full-length transcriptome sequencing. *Food Chemistry*, 300. https://doi.org/10.1016/j.foodchem.2019.125246

Han, L., Zhai, Y., Wang, Y., Shi, X., Xu, Y., Gao, S., Zhang, M., Luo, J., & Zhang, Q. (2022). Diacylglycerol Acyltransferase 3(DGAT3) Is Responsible for the Biosynthesis of Unsaturated Fatty Acids in Vegetative Organs of Paeonia rockii. *International Journal of Molecular Sciences*, 23(22), 1–19. https://doi.org/10.3390/ijms232214390

Harwood, J., & Aparicio, R. (2000). Handbook of Olive Oil. https://doi.org/10.1007/978-1-4757-5371-4

Hong, F., & Breitling, R. (2008). A comparison of meta-analysis methods for detecting differentially expressed genes in microarray experiments. *Bioinformatics*, 24(3), 374–382. https://doi.org/10.1093/bioinformatics/btm620

Iaria, D. L., Chiappetta, A., & Muzzalupo, I. (2016). A De novo Transcriptomic Approach to Identify Flavonoids and Anthocyanins "Switch-Off" in Olive (Olea europaea L.) Drupes at Different Stages of Maturation. *Frontiers in Plant Science*, 6 (December). https://doi.org/10.3389/fpls.2015.01246

IOC. (2015). International Trade Standard Applying To Olive Oils and Olive-Pomace Oils. 3, 1–17.

Keel, B. N., Zarek, C. M., Keele, J. W., Kuehn, L. A., Snelling, W. M., Oliver, W. T., Freetly, H. C., & Lindholm-Perry, A. K. (2018). RNA-Seq Meta-analysis identifies genes in skeletal muscle associated with gain and intake across a multi-season study of crossbred beef steers. *BMC Genomics*, 19(1), 1–11. https://doi.org/10.1186/s12864-018-4769-8

Laurens, F., Aranzana, M. J., Arus, P., Bassi, D., Bink, M., Bonany, J., Caprera, A., Corelli-Grappadelli, L., Costes, E., Durel, C.-E., Mauroux, J.-B., Muranty, H., Nazzicari, N., Pascal, T., Patocchi, A., Peil, A., Quilot-Turion, B., Rossini, L., Stella, A., ... van de Weg, E. (2018). An integrated approach for increasing breeding efficiency in apple and peach in Europe. *Horticulture Research*, 5, 11. https://doi.org/10.1038/s41438-018-0016-3

Lavee, S., Avidan, B., & Ben-Ari, G. (2014). Trends in Breeding New Olive Varieties in Israel for Quality and Economic Management. Agricultural Sciences, 05(08), 701–709. https://doi.org/10.4236/as.2014.58073

Le, H., Nguyen, N. H., Ta, D. T., Le, T. N. T., Bui, T. P., Le, N. T., Nguyen, C. X., Rolletschek, H., Stacey, G., Stacey, M. G., Pham, N. B., Do, P. T., & Chu, H. H. (2020). CRISPR/Cas9-Mediated Knockout of Galactinol Synthase-Encoding Genes Reduces Raffinose Family Oligosaccharide Levels in Soybean Seeds. *Frontiers in Plant Science*, 11(December), 1–13. https://doi.org/10.3389/fpls.2020.612942

Lozano-Castellon, J., Lopez-Yerena, A., Rinaldi de Alvarenga, J. F., Romero Del Castillo-Alba, J., Vallverdu-Queralt, A., Escribano-Ferrer, E., & Lamuela-Raventos, R. M. (2020). Health-promoting properties of oleocanthal and oleacein: Two secoiridoids from extra-virgin olive oil. *Critical Reviews in Food Science and Nutrition*, 60(15), 2532–2548. https://doi.org/10.1080/10408398.2019.1650715 Manan, S., Chen, B., She, G., Wan, X., & Zhao, J. (2017). Critical Reviews in Biotechnology Transport and transcriptional regulation of oil production in plants. 8551. https://doi.org/10.1080/07388551.2016.1212185

Mele, M. A., Islam, M. Z., Kang, H. M., & Giuffre, A. M. (2018). Pre-and post-harvest factors and their impact on oil composition and quality of olive fruit. *Emirates Journal of Food and Agriculture*, 30(7), 592–603. https://doi.org/10.9755/ejfa.2018.v30.i7.1742

Miho, H., Moral, J., Barranco, D., Ledesma-Escobar, C. A., Priego-Capote, F., & Diez, C. M. (2021). Influence of genetic and interannual factors on the phenolic profiles of virgin olive oils. *Food Chemistry*, 342, 128357. https://doi.org/10.1016/j.foodchem.2020.128357

Mikrou, T., Pantelidou, E., Parasyri, N., Papaioannou, A., Kapsokefalou, M., Gardeli, C., & Mallouchos, A. (2020). Varietal and Geographical Discrimination of Greek Monovarietal Extra Virgin Olive Oils Based on Squalene, Tocopherol, and Fatty Acid Composition. *Molecules*, 25(17), 3818. https://doi.org/10.3390/molecules25173818

Mougiou, N., Trikka, F., Trantas, E., Ververidis, F., Makris, A., Argiriou, A., & Vlachonasios, K. E. (2018). Expression of hydroxytyrosol and oleuropein biosynthetic genes are correlated with metabolite accumulation during fruit development in olive, Olea europaea, cv. Koroneiki. *Plant Physiology and Biochemistry*, 128 (April), 41–49. https://doi.org/10.1016/j.plaphy.2018.05.004

Mousavi, S., de la Rosa, R., Moukhli, A., El Riachy, M., Mariotti, R., Torres, M., Pierantozzi, P., Stanzione, V., Mastio, V., Zaher, H., El Antari, A., Ayoub, S., Dandachi, F., Youssef, H., Aggelou, N., Contreras, C., Maestri, D., Belaj, A., Bufacchi, M., ... Leon, L. (2019). Plasticity of fruit and oil traits in olive among different environments. *Scientific Reports*, 9(1), 1–13. https://doi.org/10.1038/s41598-019-53169-3

Mousavi, S., Stanzione, V., Mariotti, R., Mastio, V., Azariadis, A., Passeri, V., Valeri, M. C., Baldoni, L., & Bufacchi, M. (2022). Bioactive Compound Profiling of Olive Fruit: The Contribution of Genotype. *Antioxidants*, 11(4), 672. https://doi.org/10.3390/antiox11040672

Nissim, Y., Shloberg, M., Biton, I., Many, Y., Doron-Faigenboim, A., Zemach, H., Hovav, R., Kerem, Z., Avidan, B., & Ben-Ari, G. (2020). High temperature environment reduces olive oil yield and quality. *PLOS ONE*, 15(4), e0231956. https://doi.org/10.1371/journal.pone.0231956

Nissim, Y., Shlosberg, M., Biton, I., Many, Y., Doron-Faigenboim, A., Hovav, R., Kerem, Z., Avidan, B., & Ben-Ari, G. (2020). A High Temperature Environment Regulates the Olive Oil Biosynthesis Network. *Plants*, 9(9), 1135. https://doi.org/10.3390/plants9091135

Parra, R., Paredes, M. A., Sanchez-Calle, I. M., & Gomez-Jimenez, M. C. (2013). Comparative transcriptional profiling analysis of olive ripe-fruit pericarp and abscission zone tissues shows expression differences and distinct patterns of transcriptional regulation. *BMC Genomics*, 14(1), 1–20. https://doi.org/10.1186/1471-2164-14-866

Parvini, F., Zeinanloo, A. A., Ebrahimie, E., Tahmasebi-Enferadi, S., & Hosseini-Mazinani, M. (2015). Differential expression of fatty acid desaturases in Mari and Shengeh olive cultivars during fruit development and ripening. *European Journal of Lipid Science and Technology*, 117(4), 523–531. https://doi.org/10.1002/ejlt.201400327

Perez, A. G., Leon, L., Sanz, C., & de la Rosa, R. (2018). Fruit Phenolic Profiling: A New Selection Criterion in Olive Breeding Programs. *Frontiers in Plant Science*, 9. https://doi.org/10.3389/fpls.2018.00241

Periappuram, C., Steinhauer, L., Barton, D. L., Taylor, D. C., Chatson, B., & Zou, J. (2000). The plastidic phosphoglucomutase from arabidopsis. A reversible enzyme reaction with an important role in metabolic control. *Plant Physiology*, 122(4), 1193–1199. https://doi.org/10.1104/pp.122.4.1193

Rallo, L., Barranco, D., De la Rosa, R., & Leon, L. (2018). New olive cultivars and selections in Spain: results after 25 years of breeding. *Acta Horticulturae*, 1199, 21–26. https://doi.org/10.17660/ActaHortic.2018.1199.4 Riachy, M. El, Hamade, A., Ayoub, R., Dandachi, F., & Chalak, L. (2019). Oil Content, Fatty Acid and Phenolic Profiles of Some Olive Varieties Growing in Lebanon. 6(July). https://doi.org/10.3389/fnut.2019.00094

Rodriguez, J. M. (2013). De Novo Assembly and Functional Annotation of the Olive (Olea europaea) Transcriptome. November, 93–108. https://doi.org/10.1093/dnares/dss036

Rugini, E., Cristofori, V., & Silvestri, C. (2016). Genetic improvement of olive (Olea europaea L.) by conventional and in vitro biotechnology methods. In *Biotechnology Advances* (Vol. 34, Issue 5, pp. 687–696). Elsevier Inc. https://doi.org/10.1016/j.biotechadv.2016.03.004

Salas, J. J., Harwood, J. L., & Martinez-Force, E. (2013). Lipid Metabolism in Olive: Biosynthesis of Triacylglycerols and Aroma Components. In R. Aparicio & J. Harwood (Eds.), *Handbook of Olive Oil* (Second, Issue January 2014, pp. 97–127). Springer New York Heidelberg Dordrecht London. https://doi.org/10.1007/978-1-4614-7777-8

Sanchez, J., & Harwood, J. L. (2002). Biosynthesis of triacylglycerols and volatiles in olives. *European Journal of Lipid Science and Technology*, 104 (9–10), 564–573. https://doi.org/10.1002/1438-9312(200210)104:9/10<564::AID-EJLT564>3.0.CO;2-5

Santos-Antunes, F., Leon, L., de la Rosa, R., Alvarado, J., Mohedo, A., Trujillo, I., & Rallo, L. (2005). The Length of the Juvenile Period in Olive as Influenced by Vigor of the Seedlings and the Precocity of the Parents. *HortScience*, 40(5), 1213–1215. https://doi.org/10.21273/HORTSCI.40.5.1213

Sengupta, S., Mukherjee, S., Basak, P., & Majumder, A. L. (2015). Significance of galactinol and raffinose family oligosaccharide synthesis in plants. *Frontiers in Plant Science*, 6(AUG), 1–11. https://doi.org/10.3389/fpls.2015.00656

Sidorov, R. A., & Tsydendambaev, V. D. (2014). Biosynthesis of Fatty Oils in Higher Plants. 61(1), 1–18. https://doi.org/10.1134/S1021443714010130

Tan, W. J., Yang, Y. C., Zhou, Y., Huang, L. P., Xu, L., Chen, Q. F., Yu, L. J., & Xiao, S. (2018). DIACYLGLYCEROL ACYLTRANSFERASE and DIACYLGLYCEROL KINASE modulate triacylglycerol and phosphatidic acid production in the plant response to freezing stress. *Plant Physiology*, 177(3), 1303–1318. https://doi.org/10.1104/pp.18.00402

Torres, M., Pierantozzi, P., Searles, P., Rousseaux, M. C., García-Inza, G., Miserere, A., Bodoira, R., Contreras, C., & Maestri, D. (2017). Olive Cultivation in the Southern Hemisphere: Flowering, Water Requirements and Oil Quality Responses to New Crop Environments. *Frontiers in Plant Science*, 8. https://doi.org/10.3389/fpls.2017.01830

Unver, T., Wu, Z., Sterck, L., Turktas, M., Lohaus, R., Li, Z., Yang, M., He, L., Deng, T., Escalante, F. J., Llorens, C., Roig, F. J., Parmaksiz, I., Dundar, E., Xie, F., Zhang, B., Ipek, A., Uranbey, S., Erayman, M., ... Van de Peer, Y. (2017). Genome of wild olive and the evolution of oil biosynthesis. *Proceedings of the National Academy of Sciences*, 201708621. https://doi.org/10.1073/pnas.1708621114

Wang, L., Kazachkov, M., Shen, W., Bai, M., Wu, H., & Zou, J. (2014). Deciphering the roles of Arabidopsis LPCAT and PAH in phosphatidylcholine homeostasis and pathway coordination for chloroplast lipid synthesis. *The Plant Journal*, 80(6), 965–976. https://doi.org/10.1111/tpj.12683

Yanik, H., Turktas, M., Dundar, E., Hernandez, P., Dorado, G., & Unver, T. (2013). Genome-wide identification of alternate bearing-associated microRNAs (miRNAs) in olive (Olea europaea L.). *BMC Plant Biology*, 13(1), 1. https://doi.org/10.1186/1471-2229-13-10

Yubero-Serrano, E. M., Lopez-Moreno, J., Gomez-Delgado, F., & Lopez-Miranda, J. (2019). Extra virgin olive oil: More than a healthy fat. *European Journal of Clinical Nutrition*, 72(S1), 8–17. https://doi.org/10.1038/s41430-018-0304-x

Zeng, Y., Tan, X., Zhang, L., Jiang, N., & Cao, H. (2014). Identification and expression of fructose-1,6bisphosphate aldolase genes and their relations to oil content in developing seeds of tea oil tree (Camellia oleifera). *PLoS ONE*, 9(9). https://doi.org/10.1371/journal.pone.0107422

Additional files

Additional file1. Up and down-regulated identified genes of the galactose metabolism pathway in the biosynthesis of raffinose family oligosaccharides.

Additional file 2. Up and down-regulated identified genes of the glycolysis pathway in production of acetyl-CoA.

Additional file 3. Up and down-regulated identified genes of the pyruvate pathway in production of acetyl-CoA.

Additional file 4. Summary of important pathways and identified important genes in the meta-analysis.

Additional file 5. Identification of enriched genes in glycolysis and pentose phosphate pathways.

Additional file 6. Up and down-regulated identified genes in the Fatty acid biosynthesis pathway.

Additional file 7. Up and down-regulated identified genes in the Fatty acid degradation pathway.

Additional file 8. Up and down-regulated identified genes in the glycerolipid metabolism pathway.

Additional file 9. Up and down-regulated identified genes in the terpenoid backbone biosynthesis pathway.

Additional file 10. The results of search strategies and the information of thirty experiments was recorded.