

# Brain tumours, brain metastases, and neuroinflammation: Insights from neuroimaging studies.

Rafael Roesler<sup>1</sup>, Simone Dini<sup>2</sup>, and Gustavo Isolan<sup>2</sup>

<sup>1</sup>Federal University of Rio Grande do Sul

<sup>2,3</sup> The Center for Advanced Neurology and Neurosurgery (CEANNE)-Brazil, Porto Alegre, RS, Brazil

September 24, 2021

## Abstract

Brain tumors and brain metastases induce changes in brain tissue remodelling that lead to immunosuppression and trigger an inflammatory response within the tumor microenvironment. These immune and inflammatory changes can influence invasion and metastasis. Other neuroinflammatory and necrotic lesions may occur in patients with brain cancer or brain metastases as sequelae from treatment with radiotherapy. Glioblastoma (GBM) is the most aggressive primary malignant brain cancer in adults. Imaging methods such as positron-emission tomography (PET) and different magnetic resonance imaging (MRI) techniques are highly valuable for the diagnosis and therapeutic evaluation of GBM and other malignant brain tumors. However, differentiating between tumor tissue and inflamed brain tissue with imaging protocols remains a challenge. Here, we review recent advances in imaging methods that have helped to improve the specificity of primary tumor diagnosis versus evaluation of inflamed and necrotic brain lesions. We also comment on advances in differentiating metastasis from neuroinflammation processes. Recent advances include the radiosynthesis of 18F-FIMP, an L-type amino acid transporter 1 (LAT1)-specific PET probe that allows better differentiation between tumor tissue and inflammation compared to previous probes; and the combination different advanced imaging protocols with the inclusion of radiomics and machine learning algorithms.

## Clinical & Experimental Immunology

Invited Review for Special Issue on “Imaging Immunological Processes in Neuroinflammatory Diseases”

REVIEW ARTICLE

## Neuroinflammation and immunoregulation in glioblastoma and brain metastases: recent developments in imaging approaches

Rafael Roesler<sup>1,2,\*</sup>, Simone Afonso Dini<sup>3</sup>, Gustavo R. Isolan<sup>3,4,\*</sup>

1. *Department of Pharmacology, Institute for Basic Health Sciences, Federal University of Rio Grande do Sul, 90050-170 Porto Alegre, RS, Brazil; rafaelroesler@hcpa.edu.br*
2. *Cancer and Neurobiology Laboratory, Experimental Research Center, Clinical Hospital (CPE-HCPA), Federal University of Rio Grande do Sul, 90035-003 Porto Alegre, RS, Brazil*
3. *The Center for Advanced Neurology and Neurosurgery (CEANNE)-Brazil, Porto Alegre, RS, Brazil*
4. *Mackenzie Evangelical University of Paraná (FEMPAR), 80730-000 Curitiba, PR, Brazil; gustavo.isolan@fempar.edu.br*

\* Correspondence: these authors should be considered co-corresponding authors: rafaelroesler@hcpa.edu.br (R.R.); gustavo.isolan@fempar.edu.br (G.R.I.).

## Summary

Brain tumors and brain metastases induce changes in brain tissue remodelling that lead to immunosuppression and trigger an inflammatory response within the tumor microenvironment. These immune and inflammatory changes can influence invasion and metastasis. Other neuroinflammatory and necrotic lesions may occur in patients with brain cancer or brain metastases as sequelae from treatment with radiotherapy. Glioblastoma (GBM) is the most aggressive primary malignant brain cancer in adults. Imaging methods such as positron-emission tomography (PET) and different magnetic resonance imaging (MRI) techniques are highly valuable for the diagnosis and therapeutic evaluation of GBM and other malignant brain tumors. However, differentiating between tumor tissue and inflamed brain tissue with imaging protocols remains a challenge. Here, we review recent advances in imaging methods that have helped to improve the specificity of primary tumor diagnosis versus evaluation of inflamed and necrotic brain lesions. We also comment on advances in differentiating metastasis from neuroinflammation processes. Recent advances include the radiosynthesis of  $^{18}\text{F}$ -FIMP, an L-type amino acid transporter 1 (LAT1)-specific PET probe that allows better differentiation between tumor tissue and inflammation compared to previous probes; and the combination different advanced imaging protocols with the inclusion of radiomics and machine learning algorithms.

**Keywords** Glioblastoma; Brain metastases; Magnetic Resonance Imaging; Positron-Emission Tomography; Imaging; Brain tumor

## Introduction

Neuroimaging techniques are crucial tools for diagnosing, staging, and monitoring the treatment effects in patients with brain cancers [1]. Structural magnetic resonance imaging (MRI) helps to identify, classify, and grade brain tumors, as well as to guide surgery [2]. Complementary information is obtained with positron emission tomography (PET) imaging, where insights on tissue metabolism can be evaluated, which is particularly valuable for the measurement of fast cell proliferation in tumors and investigation of early-stage tumors [3-6].

Despite the current efficiency of these imaging techniques in brain tumor detection, some limitations remain to be overcome. For example, differentiating among relapsed brain tumors, solitary brain metastases, and inflammatory and necrotic lesions resulting from chemo- and radiotherapy remains a challenge. For example, the most commonly used PET probe for tumor imaging is 2-Deoxy-2- $^{18}\text{F}$ -fluoro-D-glucose (FDG), which is actively transported into cells via glucose transporters (GLUTs), accumulating not only in tumor tissue, which contains inflammatory processes itself, but also in other inflamed areas, where glucose metabolism is crucial for neuroinflammatory and neuroimmune responses. This can lead to false positive results for tumor diagnosis [7-9]. Also, both tumors and metastatic sites can display MRI patterns featuring peritumoral hyperintensities with similar intratumoral texture, thus representing a potential confounder for neuroradiologists [10]. In this review, we will discuss selected recent advances in imaging approaches aimed at improving the evaluation of brain tumors and brain metastases in relation to immunomodulation and neuroinflammatory processes. We will focus specifically on one type of brain tumor, glioblastoma (GBM), given that it represents the most aggressive primary malignant brain cancer in adults.

## Glioblastoma (GBM)

The current World Health Organization (WHO) classification divides GBM tumors into three groups depending on the status of the isocitrate dehydrogenase (*IDH*) gene: *IDH* wild-type (IDHwt) GBM, mutated *IDH*, or not specified GBM (NOS, unevaluated status). IDHwt GBMs represent around 90% of cases and typically appear in older patients, whereas around 10% of patients present tumors with an *IDH* mutation, which correspond to secondary GBMs which originate from lower grade gliomas, occur in younger patients, and have a better prognosis. Furthermore, GBMs are defined by other genetic biomarkers, including O6-methylguanine DNA methyltransferase (*MGMT*) promoter methylation, chromosome 1p/19q deletion, mutations of telomerase reverse transcriptase (*TERT*) promoter, tumor protein P53 (*TP53*), and phosphatase and tensin homolog (*PTEN*), and amplification of epidermal growth factor receptor (*EGFR*) and platelet-derived growth factor receptor A (*PDGFRA*). The prognosis for patients with GBM is dismal; current treatment, based on combining surgical resection, radiotherapy and adjuvant chemotherapy still

results in a median overall survival of less than 2 years [11-15].

### **Immunoregulation and inflammation in glioblastoma: brain tumors as neuroinflammatory diseases**

Inflammation is an integral component of cancer pathology, contributing to carcinogenesis and tumor progression. One common feature of GBM is tissue necrosis accompanied by microenvironment inflammation. Immunosuppressive inflammation with associated necrosis is typical of GBMs that display higher resistance to therapies and worst prognosis [16-20]. GBM cells express and secrete immune suppressive chemokines and cytokines including interleukin-6, interleukin-10, transforming growth factor (TGF)- $\beta$ , galectin-1, and prostaglandin-E, which act on infiltrating immune cells to hijack them by inducing a pro-tumor cellular phenotype. Thus, a population of non-neoplastic cells consisting mostly of tumor-associated macrophages, is established in the tumor site. These immunosuppressive changes enabled by inflammatory mediators stimulate GBM cell proliferation, migration, angiogenesis, and resistance to treatment. For example, signaling mediated by CXCR3 and CCR2 receptors recruit tumor-promoting immune cells such as T cells and myeloid-derived suppressor cells [16, 21-28]. In GBM cancer stem cells (CSCs), hypoxic tissue triggers expression of genes of the inflammatory reparative response [25, 29]. Cytokines such as interleukin (IL)-1 $\beta$  and TGF- $\beta$  cooperate to induce inflammatory gene expression and a proinflammatory phenotype in GBM CSCs, which in turn facilitates cellular proliferation and migration [30].

### **Neuroinflammation as a consequence of GBM treatment**

Inflammation and necrosis of healthy brain tissue is one of the main unwanted effects of radiotherapy used to treat GBM. Studies have found incidences of radiation-induced necrosis ranging from 2.5% to about 30% of GBM patient cohorts [31-33]. Necrosis can include underlying inflammation together with occlusive vasculopathy and perivascular parenchymal changes. Alterations in the integrity of the blood-brain barrier in inflamed sites leads to immune cell infiltration, fluid transudation into the interstitial space and brain edema. Infiltrated immune cells as well as reactive glial cells increase expression of mediators, amplifying the response [31, 34-36].

### **Consequences of inflammation for imaging diagnosis: differentiating between tumors and inflammatory lesions**

Conventional neuroradiological techniques do not always allow a differentiation between radiation necrosis and GBM recurrence, given that both recurrent tumors and necrosis (pseudoprogressions) may appear as similar lesions in MRI protocols such as T2- and gadolinium-enhanced T1-weighted imaging, with an increase in contrast enhancement and the image of a mass [31, 37-42]. PET imaging using  $^{18}\text{F}$ -Fluorodeoxyglucose (FDG-PET), in turn, can be useful in differentiating radiation-induced lesions from recurrent GBM [31, 43]. However, with this method false-positive and false-negative PET scan results can lead to low sensitivity and pose a severe limitation [44].

### **Imaging advances for the evaluation of brain tumors and neuroinflammation**

The measurement of metabolic and cell physiology parameters with improved MRI techniques provides more detailed information on tissue biochemical composition and blood perfusion, facilitating the distinction between tumors and necrosis. Proton magnetic resonance (MR) spectroscopic imaging is used to evaluate cell metabolism through the detection of proton metabolites and their distribution. Chemicals detected include compounds containing choline, creatine, lactate, lipid, and N-acetylaspartate. Choline, a component of cell membrane phospholipids, is particularly useful for differentiating tumors from non-tumoral tissues because its content is increased in highly proliferative cell populations. Overall, higher choline levels are associated with disease progression or recurrence, whereas low levels of choline are found in necrotic lesions. The analysis of increased ratios of choline content in relation to other chemicals can result in an accuracy of up to 97% in separating tumors from non-tumoral necrotic tissues [31, 42].

Conventional MRI, with the T1/T2 mismatch criterion, had a specificity of 75% and a sensitivity of only 44% in distinguishing between tumors and inflamed lesions. PET scan combined the best sensitivity and speci-

ficity, respectively of 92% and 69%. PET remained superior compared to NMR spectroscopy for choline/N-acetylaspartate and choline/creatin ratios across different thresholds [45]. A retrospective study of 57 GBM patients examined with T2\*-weighted dynamic susceptibility-weighted contrast material-enhanced (DSC) MRI found mean, maximum, and minimum relative peak height and relative cerebral blood volume were significantly higher in GBM cases compared to radiation necrosis cases. In contrast, mean, maximum, and minimum relative percentage of signal intensity recovery values were significantly lower in recurrent GBM compared to radiation necrosis [46]. Proton MR spectroscopy showed a temporary elevation of choline in 4 of 9 cases of necrosis, creating a confounding factor that could result in false positive findings for tumor recurrence [47]. Another study using proton MR spectroscopy in 11 patients who received high-dose radiotherapy revealed that cases of radiation necrosis had either increased lactate/creatine and phosphocreatine (Cr) ratio and decreased choline-containing compounds/phosphocreatine ratio compared to recurrent GBM, or reductions in all major metabolites [48]. A meta-analysis of 397 patients in 13 studies examined roles of several metabolites. MR spectroscopy and MR perfusion using Cho/NAA and Cho/Cr ratios and rCBV may increase the accuracy of differentiating necrosis from recurrent tumor in patients with primary brain tumors or brain metastases [49].

A meta-analysis of 6 studies with 118 patients and 134 scans indicated  $^{11}\text{C}$ -choline PET as an accurate diagnostic method for the differentiation of tumor relapse from radiation induced necrosis in gliomas [50]. A study with 55 patients, followed up for at least 11 months, with suspected brain tumor recurrence or necrosis after radiotherapy, examined MRI,  $^{18}\text{F}$ -FDG, and  $^{11}\text{C}$ -choline PET/CT, concluding for the superiority of  $^{11}\text{C}$ -choline PET/CT [51]. In a F98 orthotopic rat model of GBM, PET using  $^{18}\text{F}$ -FDG and  $^{18}\text{F}$ -FET PET were effective in discriminating GBM from radiation necrosis, with  $^{18}\text{F}$ -FDG delayed PET being particularly useful [52]. A study with 50 patients showed that  $^{11}\text{C}$ -methionine-PET was superior to both  $^{11}\text{C}$ -choline and  $^{18}\text{F}$ -FDG -PET for distinguishing GBM recurrence from radiation necrosis [53]. The LAT1 tumor-specific PET tracer 2- $^{18}\text{F}$ ]-FELP PET is able to differentiate glioblastoma from radiation necrosis, and 2- $^{18}\text{F}$ ]-FELP uptake is less likely to be contaminated by the presence of inflammation than the  $^{18}\text{F}$ ]-FDG signal [35].

Differentiation of brain tumors from brain infections and abscesses

Brain abscesses caused by aerobic bacteria can mimic GBM tumors in MRI image appearance and single voxel proton MR spectroscopy, leading to misdiagnosis. Based on the hypothesis that metabolite levels of choline would be decreased in the ring-enhancing portion of abscesses compared to GBM, one study using MR spectroscopic imaging found that metabolite ratios and maximum choline/choline-n, choline/creatine, and choline/ N-acetylaspartate ratios of the contrast-enhancing rim could be useful differentiating aerobic abscesses from GBM. In MRI procedures, the apparent diffusion coefficient (APC) may be useful in differentiating GBM from brain fungal abscess. Combining diffusion tensor imaging (DTI) and dynamic susceptibility contrast perfusion-weighted imaging (DSC-PWI) provides better results in differentiating lesions from brain infections than each technique used alone [54-56].

### **Brain metastases, inflammation, and imaging**

It is worth noting that CSCs constitute the main cell population mediating metastasis [57]. Brain metastases are more common than tumors originating primarily in the brain and confer grave prognosis to patients with various types of primary cancers originating in other sites, such as lung, breast, colorectal, and melanoma, given that available treatments show very limited efficacy. The tumor microenvironment is crucial to determine the establishment of brain metastases, and within this context, neuroinflammation plays a central role [58].

Brain metastases can produce a tissue lesion that induces a response comprising persistent astrocyte and microglia activation with cytokine and chemokine release, increased blood vessel permeability and recruitment of immune cells, resulting in neuroinflammation [58-61]. Also, non-neoplastic inflamed sites in the brain may facilitate the adhesion of circulating tumor cells from peripheral tumors to the activated endothelium in brain vessels, which is one of the possible mechanisms promoting metastasis initiation [62]. Inflammation

at distant sites promotes adhesion of CTCs to the activated endothelium and then initiates the formation of metastases. Many different types of mediators and immune cells are involved in brain metastasis, depending among other factors on the type of primary tumor of origin, metastatic site in the brain, and differential astrocyte and microglial activation, resulting in high biological heterogeneity [58, 63-67].

In terms of imaging, differentiating brain tumors from brain metastases pose a challenge by itself, as both present imaging patterns with similar peritumoral hyperintensities and intratumoral texture on MRI. Improvements have been obtained with the use of relative cerebral blood flow and volume analyses, diffusion tensor imaging, neurite orientation dispersion, density imaging to examine intracellular volume fraction, isotropic volume fraction, and extracellular volume fraction, and metabolite analysis with MR spectroscopy. Also, intratumoral creatine suggests GBM, whereas its absence indicates metastasis when single-voxel proton MR spectroscopy is used for imaging [68, 69]. A study examining the use of the mean apparent diffusion coefficient and absolute standard deviation derived from apparent diffusion coefficient measurements based on cellularity levels could not find marked differences between GBM tumors from brain metastasis [70]. However, another study found that the apparent diffusion coefficient could differentiate GBM from metastasis, and that homogeneity and the inverse difference moment of GBM were significantly higher than those of metastases in the regions of interest placements examined [71, 72].

Assessing the heterogeneity of both the tumor masses and peritumoral edema with MRI texture analysis revealed that the heterogeneity of the GBMs peritumoral edema was significantly higher than the edema surrounding MET, differentiating them with a sensitivity of 80% and specificity of 90% [73]. Combining arterial spin labeling perfusion (ASL)- and diffusion tensor imaging (DTI)-derived metrics showed to be promising in differentiating GBM and solitary brain metastases [74]. The use of 2D texture features extracted from images obtained with MRI may be a useful alternative for discriminating between GBM and brain metastases [75]. Computational-aided quantitative analysis of MRI images may improve the accuracy in differentiating GBM from metastases, and texture features are more significant than fractal-based features for that purpose [76]. Increasingly, machine learning algorithms have been applied to imaging data to improve the differentiation between GBM and brain metastases [77-81]. Novel diagnostic support systems based on radiomic features extracted from post-contrast 3DT1 MR images may help improving the distinction between solitary brain metastases and GBM with high diagnosis performance and generalizability [77]. Machine learning and deep learning-based models applied to conventional MR images may support preoperative discrimination between GBM and solitary brain metastasis conventional MR images [78-80], and deep learning network models that allow automated, on-site analysis of resected tumor specimens based on confocal laser endoscopic techniques image datasets have been developed [81]. Other parameters such as the cerebral blood volume gradient in the peritumoral brain zone may enable the differentiation of GBMs from metastases [82]. Another approach is to evaluate peritumoral areas with color map of phase difference enhanced imaging (Color PADRE) [83].

As with tumors originating in the brain, metastases are treated with radiotherapy and stereotactic radiosurgery, making incidence of radiation necrosis an important issue, and the distinction between metastasis and inflamed and necrotic tissue by MRI can be challenging. One study examined the hypothesis that methionine levels could be increased in metastatic tissue, whereas the inflammation marker translocator protein (PBR-TSPO), which can be quantified with specific PET tracers, would be increased in necrosis. Thus, the use of the [ $^{11}\text{C}$ ]methionine and [ $^{11}\text{C}$ ]PBR28 tracers in PET was evaluated in 5 patients previously treated brain metastases showing regrowth. The use of [ $^{11}\text{C}$ ]methionine could accurately identify pathologically confirmed tumor regrowth in all 7 lesions examined, whereas [ $^{11}\text{C}$ ]PBR28 could only identify 3 of 7 lesions, indicating that the former, but not the later tracer can be used as a reliable marker [84].

### Concluding remarks

As discussed above, the differentiation between GBM recurrence and radiation necrosis remains a crucial challenge (Fig. 1), given that the need for additional surgery in a previously operated brain region increases surgical risk in addition to raising treatment costs. An important current tool is multiparametric MRI using a combination of perfusion and diffusion parameters. Although previous studies [85-91] had found that apparent diffusion coefficient, volume transfer constant, and relative cerebral blood volume are very useful,

and the latter is the strongest parameter for differentiating radiation necrosis from GBM tumors [92]. Nael et al. [93] achieved a diagnostic accuracy of 92.8% by combining the use of volume transfer constant and relative cerebral blood volume.

In daily neuro-oncological practice, multiparametric MRI is the most used tool to achieve an accurate differential diagnosis between GBM, inflamed sites, and other brain diseases. Despite its advantages, the use of PET Scan remains limited in many regions of the world due to economical and logistic shortcomes. Overall, the integrated use of different MR and PET approaches, including multiparametric MRI, in addition to perfusion and diffusion parameters [94] may enable increasingly accurate diagnosis based on brain imaging.

### Acknowledgements

The writing of this article was supported by the National Council for Scientific and Technological Development (CNPq, MCTI, Brazil) grant 305647/2019-9 (R.R.); and The Center for Advanced Neurology and Neurosurgery (CEANNE)-Brazil (S.A.D., G.R.I.).

### Disclosure

The authors declare no conflicts of interest.

### References

1. Drake LR, Hillmer AT, Cai Z. Approaches to PET imaging of glioblastoma. *Molecules* 2020; **25**: 568. doi: 10.3390/molecules25030568
2. Reza SMS, Samad MD, Shboul ZA, Jones KA, Iftekharuddin KM. Glioma grading using structural magnetic resonance imaging and molecular data. *J Med Imaging (Bellingham)* 2019, **6**: 024501. doi: 10.1117/1.JMI.6.2.024501
3. Chiang GC, Kovanlikaya I, Choi C, Ramakrishna R, Magge R, Shungu DC. Magnetic resonance spectroscopy, positron emission tomography and radiogenomics-relevance to glioma. *Front Neurol* 2018, **9**: 33. doi: 10.3389/fneur.2018.00033
4. Frosina G. Positron emission tomography of high-grade gliomas. *J Neurooncol* 2016, **127**: 415-25. doi: 10.1007/s11060-016-2077-1
5. Holzgreve A, Albert NL, Galldiks N, Suchorska B. Use of PET imaging in neuro-oncological surgery. *Cancers (Basel)* 2021, **13**: 2093. doi: 10.3390/cancers13092093
6. Moreau A, Febvey O, Mognetti T, Frappaz D, Kryza D. Contribution of different positron emission tomography tracers in glioma management: focus on glioblastoma. *Front Oncol* 2019, **9**: 1134. doi: 10.3389/fonc.2019.01134
7. Cook GJ, Maisey MN, Fogelman I. Normal variants, artefacts and interpretative pitfalls in PET imaging with 18-fluoro-2-deoxyglucose and carbon-11 methionine. *Eur J Nucl Med* 1999, **26**: 1363–78. doi: 10.1007/s002590050597
8. Culverwell AD, Scarsbrook AF, Chowdhury FU. False-positive uptake on 2-[<sup>18</sup>F]-fluoro-2-deoxy-D-glucose (FDG) positron-emission tomography/computed tomography (PET/CT) in oncological imaging. *Clin Radiol* 2011, **66**: 366–82. doi: doi.org/10.1016/j.crad.2010.12.004
9. Nozaki S, Nakatani Y, Mawatari A, Shibata N, Hume WE, Hayashinaka E *et al.* 18F-FIMP: a LAT1-specific PET probe for discrimination between tumor tissue and inflammation. *Sci Rep* 2019, **9**: 15718. doi: 10.1038/s41598-019-52270-x
10. Fordham AJ, Hacherl CC, Patel N, Jones K, Myers B, Abraham M *et al.* Differentiating glioblastomas from solitary brain metastases: an update on the current literature of advanced imaging modalities. *Cancers (Basel)* 2021, **13**: 2960. doi: 10.3390/cancers13122960

11. Brennan CW, Verhaak RG, McKenna A, Campos B, Nounshmehr H, Salama SR *et al.* The somatic genomic landscape of glioblastoma. *Cell* 2013,**155**: 462-77. doi: 10.1016/j.cell.2013.09.034
12. Louis DN, Perry A, Reifenberger G, von Deimling A, Figarella-Branger D, Cavenee WK *et al.* The 2016 World Health Organization Classification of Tumors of the Central Nervous System: a summary. *Acta Neuropathol* 2016, **131**: 803-20. doi: 10.1007/s00401-016-1545-1
13. Roesler R, Brunetto AT, Abujamra AL, de Farias CB, Brunetto AL, Schwartzmann G. Current and emerging molecular targets in glioma. *Expert Rev. Anticancer Ther* 2010, **10**: 1735-51. doi: 10.1586/era.10.167
14. Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJ, *et al.* Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N Engl J Med* 2005, **352**: 987-96. doi: 10.1056/NEJMoa043330
15. Wen PY, Kesari S. Malignant gliomas in adults. *N Engl J Med* 2008,**359**: 492-507. doi: 10.1056/NEJMra0708126
16. Alghamri MS, McClellan BL, Hartlage MS, Haase S, Faisal SM, Thalla R, *et al.* Targeting neuroinflammation in brain cancer: uncovering mechanisms, pharmacological targets, and neuropharmaceutical developments. *Front Pharmacol* 2021, **12**: 680021. doi: 10.3389/fphar.2021.680021
17. Arimappamagan A, Somasundaram K, Thennarasu K, Peddagangannagari S, Srinivasan H, Shailaja BC, *et al.* A fourteen gene GBM prognostic signature identifies association of immune response pathway and mesenchymal subtype with high risk group. *PLoS One* 2013, **8**: e62042. doi: 10.1371/journal.pone.0062042
18. Carro MS, Lim WK, Alvarez MJ, Bollo RJ, Zhao X, Snyder EY, *et al.* The transcriptional network for mesenchymal transformation of brain tumours. *Nature* 2010, **463**: 318-25. doi: 10.1038/nature08712
19. DeCordova S, Shastri A, Tsolaki AG, Yasmin H, Klein L, Singh SV, *et al.* Molecular heterogeneity and immunosuppressive microenvironment in glioblastoma. *Front Immunol* 2020, **11**: 1402. doi: 10.3389/fimmu.2020.01402
20. Yeo ECF, Brown MP, Gargett T, Ebert LM. The role of cytokines and chemokines in shaping the immune microenvironment of glioblastoma: implications for immunotherapy. *Cells* 2021, **10**: 607. doi: 10.3390/cells10030607
21. Crane CA, Ahn BJ, Han SJ, Parsa AT. Soluble factors secreted by glioblastoma cell lines facilitate recruitment, survival, and expansion of regulatory T cells: implications for immunotherapy. *Neuro Oncol* 2012,**14**: 584-95. doi: 10.1093/neuonc/nos014
22. Groblewska M, Litman-Zawadzka A, Mroczko B. The role of selected chemokines and their receptors in the development of gliomas. *Int J Mol Sci* 2020, **21**: 3704. doi: 10.3390/ijms21103704
23. Huettner C, Paulus W, Roggendorf W. Messenger RNA expression of the immunosuppressive cytokine IL-10 in human gliomas. *Am J Pathol* 1995,**146**: 317-22
24. Perng P, Lim M. Immunosuppressive mechanisms of malignant gliomas: parallels at non-CNS sites. *Front Oncol* 2015, **5**: 153. doi: 10.3389/fonc.2015.00153
25. Tafani M, Di Vito M, Frati A, Pellegrini L, De Santis E, Sette G, *et al.* Pro-inflammatory gene expression in solid glioblastoma microenvironment and in hypoxic stem cells from human glioblastoma. *J Neuroinflammation* 2011, **8**: 32. doi: 10.1186/1742-2094-8-32
26. Urbantat RM, Vajkoczy P, Brandenburg S. Advances in chemokine signaling pathways as therapeutic targets in glioblastoma. *Cancers (Basel)* 2021, **13**: 2983. doi: 10.3390/cancers13122983
27. Van Meir E, Sawamura Y, Diserens AC, Hamou MF, de Tribolet N. Human glioblastoma cells release interleukin 6 in vivo and in vitro. *Cancer Res* 1990, **50**: 6683-8

28. Waters MR, Gupta AS, Mockenhaupt K, Brown LN, Biswas DD, Kordula T. RelB acts as a molecular switch driving chronic inflammation in glioblastoma multiforme. *Oncogenesis* 2019, **8**: 37. doi: 10.1038/s41389-019-0146-y
29. Papale M, Buccarelli M, Mollinari C, Russo MA, Pallini R, Ricci-Vitiani L, *et al.* Hypoxia, inflammation and necrosis as determinants of glioblastoma cancer Stem cells progression. *Int J Mol Sci* 2020, **21**: 2660. doi: 10.3390/ijms21082660
30. Wang L, Liu Z, Balivada S, Shrestha T, Bossmann S, Pyle M, *et al.* Interleukin-1 $\beta$  and transforming growth factor- $\beta$  cooperate to induce neurosphere formation and increase tumorigenicity of adherent LN-229 glioma cells. *Stem Cell Res Ther* 2012, **3**: 5. doi: 10.1186/scrt96
31. Brandes AA, Tosoni A, Spagnoli F, Frezza G, Leonardi M, Calbucci F, *et al.* Disease progression or pseudoprogression after concomitant radiochemotherapy treatment: pitfalls in neurooncology. *Neuro Oncol*. 2008, **10**: 361-7. doi: 10.1215/15228517-2008-008
32. DeAngelis LM, Delattre JY, Posner JB. Radiation-induced dementia in patients cured of brain metastases. *Neurology* 1989, **39**: 789-96. doi: 10.1212/wnl.39.6.789
33. Sheline GE, Wara WM, Smith V. Therapeutic irradiation and brain injury. *Int J Radiat Oncol Biol Phys* 1980, **6**: 1215-28. doi: 10.1016/0360-3016(80)90175-3
34. Bolcaen J, Descamps B, Deblaere K, Boterberg T, De Vos Pharm F, Kalala JP, *et al.* (18)F-fluoromethylcholine (FCho), (18)F-fluoroethyltyrosine (FET), and (18)F-fluorodeoxyglucose (FDG) for the discrimination between high-grade glioma and radiation necrosis in rats: a PET study. *Nucl Med Biol* 2015, **42**: 38-45. doi: 10.1016/j.nucmedbio.2014.07.006
35. Verhoeven J, Baguet T, Piron S, Pauwelyn G, Bouckaert C, Descamps B, Raedt R, Vanhove C, *et al.* 2-[18F]FELP, a novel LAT1-specific PET tracer, for the discrimination between glioblastoma, radiation necrosis and inflammation. *Nucl Med Biol* 2020, **82-83**: 9-16. doi: 10.1016/j.nucmedbio.2019.12.002
36. Sonar SA, Lal G. Blood-brain barrier and its function during inflammation and autoimmunity. *J Leukoc Biol* 2018, **103**: 839-53. doi: 10.1002/JLB.1RU1117-428R
37. Burger PC, Dubois PJ, Schold SC Jr, Smith KR Jr, Odom GL, Crafts DC, *et al.* Computerized tomographic and pathologic studies of the untreated, quiescent, and recurrent glioblastoma multiforme. *J Neurosurg* 1983, **58**: 159-69. doi: 10.3171/jns.1983.58.2.0159
38. Doms GC, Hecht S, Brant-Zawadzki M, Berthiaume Y, Norman D, Newton TH. Brain radiation lesions: MR imaging. *Radiology* 1986, **158**:149-55.
39. Jain R, Narang J, Sundgren PM, Hearshen D, Saksena S, Rock JP, *et al.* Treatment induced necrosis versus recurrent/progressing brain tumor: going beyond the boundaries of conventional morphologic imaging. *J Neurooncol* 2010, **100**: 17-29. doi: 10.1007/s11060-010-0139-3
40. Tihan T, Barletta J, Parney I, Lamborn K, Sneed PK, Chang S. Prognostic value of detecting recurrent glioblastoma multiforme in surgical specimens from patients after radiotherapy: should pathology evaluation alter treatment decisions? *Hum Pathol* 2006, **37**:272-82. doi: 10.1016/j.humpath.2005.11.010
41. Martínez-Bisbal MC, Celda B. Proton magnetic resonance spectroscopy imaging in the study of human brain cancer. *Q J Nucl Med Mol Imaging*. 2009, **53**: 618-30
42. Weybright P, Sundgren PC, Maly P, Hassan DG, Nan B, Rohrer S, *et al.* Differentiation between brain tumor recurrence and radiation injury using MR spectroscopy. *AJR Am J Roentgenol* 2005, **185**: 1471-6. doi: 10.2214/AJR.04.0933
43. Chen W. Clinical applications of PET in brain tumors. *J Nucl Med* 2007, **48**: 1468-81. doi: 10.2967/jnu-med.106.037689

44. Ricci PE, Karis JP, Heiserman JE, Fram EK, Bice AN, Drayer BP. Differentiating recurrent tumor from radiation necrosis: time for re-evaluation of positron emission tomography? *AJNR Am J Neuroradiol* 1998, **19**: 407-13
45. Menoux I, Noël G, Namer I, Antoni D. [PET scan and NMR spectroscopy for the differential diagnosis between brain radiation necrosis and tumour recurrence after stereotactic irradiation of brain metastases: Place in the decision tree]. *Cancer Radiother* 2017, **21**: 389-97. doi: 10.1016/j.canrad.2017.03.003
46. Barajas RF Jr, Chang JS, Segal MR, Parsa AT, McDermott MW, Berger MS, *et al.* Differentiation of recurrent glioblastoma multiforme from radiation necrosis after external beam radiation therapy with dynamic susceptibility-weighted contrast-enhanced perfusion MR imaging. *Radiology* 2009, **253**: 486-96. doi: 10.1148/radiol.2532090007
47. Nakajima T, Kumabe T, Kanamori M, Saito R, Tashiro M, Watanabe M, *et al.* Differential diagnosis between radiation necrosis and glioma progression using sequential proton magnetic resonance spectroscopy and methionine positron emission tomography. *Neurol Med Chir (Tokyo)* 2009, **49**: 394-401. doi: 10.2176/nmc.49.394
48. Kamada K, Houkin K, Abe H, Sawamura Y, Kashiwaba T. Differentiation of cerebral radiation necrosis from tumor recurrence by proton magnetic resonance spectroscopy. *Neurol Med Chir (Tokyo)* 1997, **37**:250-6. doi: 10.2176/nmc.37.250
49. Chuang MT, Liu YS, Tsai YS, Chen YC, Wang CK. Differentiating radiation-induced necrosis from recurrent brain tumor using MR perfusion and spectroscopy: a meta-analysis. *PLoS One* 2016, **11**: e0141438. doi: 10.1371/journal.pone.0141438
50. Gao L, Xu W, Li T, Zheng J, Chen G. Accuracy of 11C-choline positron emission tomography in differentiating glioma recurrence from radiation necrosis: A systematic review and meta-analysis. *Medicine (Baltimore)* 2018, **97**: e11556. doi: 10.1097/MD.00000000000011556
51. Tan H, Chen L, Guan Y, Lin X. Comparison of MRI, F-18 FDG, and 11C-choline PET/CT for their potentials in differentiating brain tumor recurrence from brain tumor necrosis following radiotherapy. *Clin Nucl Med* 2011, **36**: 978-81. doi: 10.1097/RLU.0b013e31822f68a6
52. Bolcaen J, Descamps B, Deblaere K, Boterberg T, De Vos Pharm F, Kalala JP, *et al.* (18)F-fluoromethylcholine (FCho), (18)F-fluoroethyltyrosine (FET), and (18)F-fluorodeoxyglucose (FDG) for the discrimination between high-grade glioma and radiation necrosis in rats: a PET study. *Nucl Med Biol* 2015, **42**: 38-45. doi: 10.1016/j.nucmedbio.2014.07.006
53. Takenaka S, Asano Y, Shinoda J, Nomura Y, Yonezawa S, Miwa K, Yano H, *et al.* Comparison of (11)C-methionine, (11)C-choline, and (18)F-fluorodeoxyglucose-PET for distinguishing glioma recurrence from radiation necrosis. *Neurol Med Chir (Tokyo)* 2014, **54**: 280-9. doi: 10.2176/nmc.0a2013-0117
54. Lai PH, Weng HH, Chen CY, Hsu SS, Ding S, Ko CW, *et al.* In vivo differentiation of aerobic brain abscesses and necrotic glioblastomas multiforme using proton MR spectroscopic imaging. *AJNR Am J Neuroradiol* 2008, **29**: 1511-8. doi: 10.3174/ajnr.A1130
55. Aziz K, Nawaz, Atif. 1411. Differentiation of fungal abscess of brain from brain glioblastoma by MRI scan ADC value. *Open Forum Infect Dis* 2019, **6(Suppl 2)**: S514. 2019 Oct 23. doi: 10.1093/ofid/ofz360.1275
56. Bink A, Gaa J, Franz K, Weidauer S, Yan B, Lanfermann H, *et al.* Importance of diffusion-weighted imaging in the diagnosis of cystic brain tumors and intracerebral abscesses. *Zentralbl Neurochir* 2005, **66**: 119-25. doi: 10.1055/s-2005-836478
57. Nandy SB, Lakshmanaswamy R. Cancer stem cells and metastasis. *Prog Mol Biol Transl Sci* 2017, **151**: 137-76. doi: 10.1016/bs.pmbts.2017.07.007

58. Doron H, Pukrop T, Erez N. A Blazing landscape: neuroinflammation shapes brain metastasis. *Cancer Res* 2019, **79**: 423-36. doi: 10.1158/0008-5472.CAN-18-1805
59. Gyoneva S, Ransohoff RM. Inflammatory reaction after traumatic brain injury: therapeutic potential of targeting cell-cell communication by chemokines. *Trends Pharmacol Sci* 2015, **36**: 471-80. doi: 10.1016/j.tips.2015.04.003
60. O'Callaghan JP, Sriram K, Miller DB. Defining "neuroinflammation". *Ann N Y Acad Sci* 2008, **1139**: 318-30. doi: 10.1196/annals.1432.032
61. Qian, B. Z. & Pollard, J. W. Macrophage diversity enhances tumor progression and metastasis. *Cell* **141**: 39-51, doi: 10.1016/j.cell.2010.03.014
62. Sikpa D, Whittingstall L, Fouquet JP, Radulska A, Tremblay L, Lebel R, *et al.* Cerebrovascular inflammation promotes the formation of brain metastases. *Int J Cancer* 2020, **147**: 244-55. doi: 10.1002/ijc.32902
63. Burda JE, Sofroniew MV. Reactive gliosis and the multicellular response to CNS damage and disease. *Neuron* 2014, **81**: 229-48. doi: 10.1016/j.neuron.2013.12.034
64. Doron H, Amer M, Ershaid N, Blazquez R, Shani O, Lahav TG, *et al.* Inflammatory activation of astrocytes facilitates melanoma brain tropism via the CXCL10-CXCR3 signaling axis. *Cell Rep* 2019, **28**:1785-98.e6. doi: 10.1016/j.celrep.2019.07.033
65. Klein A, Schwartz H, Sagi-Assif O, Meshel T, Izraely S, Ben Menachem S, *et al.* Astrocytes facilitate melanoma brain metastasis via secretion of IL-23. *J Pathol* 2015, **236**: 116-27. doi: 10.1002/path.4509
66. Seike T, Fujita K, Yamakawa Y, Kido MA, Takiguchi S, Teramoto N, *et al.* Interaction between lung cancer cells and astrocytes via specific inflammatory cytokines in the microenvironment of brain metastasis. *Clin Exp Metastasis* 2011, **28**: 13-25. doi: 10.1007/s10585-010-9354-8
67. Xing F, Kobayashi A, Okuda H, Watabe M, Pai SK, Pandey PR, *et al.* Reactive astrocytes promote the metastatic growth of breast cancer stem-like cells by activating Notch signalling in brain. *EMBO Mol Med* 2013, **5**: 384-96. doi: 10.1002/emmm.201201623
68. Fordham AJ, Hacherl CC, Patel N, Jones K, Myers B, Abraham M, *et al.* Differentiating glioblastomas from solitary brain metastases: an update on the current literature of advanced imaging modalities. *Cancers (Basel)* 2021, **13**: 2960. doi: 10.3390/cancers13122960
69. Ishimaru H, Morikawa M, Iwanaga S, Kaminogo M, Ochi M, Hayashi K. Differentiation between high-grade glioma and metastatic brain tumor using single-voxel proton MR spectroscopy. *Eur Radiol* 2001, **11**:1784-91. doi: 10.1007/s003300000814
70. Beig Zali S, Alinezhad F, Ranjkesh M, Daghighi MH, Poureisa M. Accuracy of apparent diffusion coefficient in differentiation of glioblastoma from metastasis. *Neuroradiol J* 2021, **34**: 205-12. doi: 10.1177/1971400920983678
71. Zhang G, Chen X, Zhang S, Ruan X, Gao C, Liu Z, *et al.* Discrimination between solitary brain metastasis and glioblastoma multiforme by using ADC-based texture analysis: a comparison of two different ROI placements. *Acad Radiol* 2019, **26**: 1466-72. doi: 10.1016/j.acra.2019.01.010
72. Fordham AJ, Hacherl CC, Patel N, Jones K, Myers B, Abraham M, *et al.* Differentiating glioblastomas from solitary brain metastases: an update on the current literature of advanced imaging modalities. *Cancers (Basel)* 2021, **13**: 2960. doi: 10.3390/cancers13122960
73. Skogen K, Schulz A, Helseth E, Ganeshan B, Dormagen JB, Server A. Texture analysis on diffusion tensor imaging: discriminating glioblastoma from single brain metastasis. *Acta Radiol* 2019, **60**: 356-66. doi: 10.1177/0284185118780889

74. Abdel Razek AAK, Talaat M, El-Serougy L, Abdelsalam M, Gaballa G. Differentiating glioblastomas from solitary brain metastases using arterial spin labeling perfusion- and diffusion tensor imaging-derived metrics. *World Neurosurg* 2019, 127, e593-e598. doi: 10.1016/j.wneu.2019.03.213
75. Ortiz-Ramón R, Ruiz-España S, Mollá-Olmos E, Moratal D. Glioblastomas and brain metastases differentiation following an MRI texture analysis-based radiomics approach. *Phys Med* 2020, **76**:44-54. doi: 10.1016/j.ejmp.2020.06.016
76. Petrujkić K, Milošević N, Rajković N, Stanisavljević D, Gavrilović S, Dželebdžić D, *et al.* Computational quantitative MR image features - a potential useful tool in differentiating glioblastoma from solitary brain metastasis. *Eur J Radiol* 2019, **119**: 108634. doi: 10.1016/j.ejrad.2019.08.003
77. de Causans A, Carré A, Roux A, Tauziède-Espariat A, Ammari S, Dezamis E, *et al.* Development of a machine learning classifier based on radiomic features extracted from post-contrast 3D T1-weighted MR images to distinguish glioblastoma from solitary brain metastasis. *Front Oncol* 2021, **11**: 638262. doi: 10.3389/fonc.2021.638262
78. Shin I, Kim H, Ahn SS, Sohn B, Bae S, Park JE, *et al.* Development and validation of a deep learning-based model to distinguish glioblastoma from solitary brain metastasis using conventional MR images. *AJNR Am J Neuroradiol* 2021, **42**: 838-44. doi: 10.3174/ajnr.A7003
79. Swinburne NC, Schefflein J, Sakai Y, Oermann EK, Titano JJ, Chen I, *et al.* Machine learning for semi-automated classification of glioblastoma, brain metastasis and central nervous system lymphoma using magnetic resonance advanced imaging. *Ann Transl Med* 2019, **7**:232. doi: 10.21037/atm.2018.08.05
80. Tateishi M, Nakaura T, Kitajima M, Uetani H, Nakagawa M, Inoue T, *et al.* An initial experience of machine learning based on multi-sequence texture parameters in magnetic resonance imaging to differentiate glioblastoma from brain metastases. *J Neurol Sci* 2020, **410**:116514. doi: 10.1016/j.jns.2019.116514
81. Ziebart A, Stadniczuk D, Roos V, Ratliff M, von Deimling A, Hänggi D, *et al.* Deep neural network for differentiation of brain tumor tissue displayed by confocal laser endomicroscopy. *Front Oncol* 2021, **11**: 668273. doi: 10.3389/fonc.2021.668273
82. She D, Xing Z, Cao D. Differentiation of glioblastoma and solitary brain metastasis by gradient of relative cerebral blood volume in the peritumoral brain zone derived from dynamic susceptibility contrast perfusion magnetic resonance imaging. *J Comput Assist Tomogr* 2019, **43**: 13-7. doi: 10.1097/RCT.0000000000000771
83. Doishita S, Sakamoto S, Yoneda T, Uda T, Tsukamoto T, Yamada E, *et al.* Differentiation of brain metastases and gliomas based on color map of phase difference enhanced imaging. *Front Neurol* 2018, **9**:788. doi: 10.3389/fneur.2018.00788
84. Tran TT, Gallezot JD, Jilaveanu LB, Zito C, Turcu G, Lim K, *et al.* [11C]methionine and [11C]PBR28 as PET imaging tracers to differentiate metastatic tumor recurrence or radiation necrosis. *Mol Imaging* 2020, **19**: 1536012120968669. doi: 10.1177/1536012120968669
85. Asao C, Korogi Y, Kitajima M, Hirai T, Baba Y, Makino K, *et al.* Diffusion-weighted imaging of radiation-induced brain injury for differentiation from tumor recurrence. *AJNR Am J Neuroradiol* 2005, **26**: 1455-60
86. Hein PA, Eskey CJ, Dunn JF, Hug EB. Diffusion-weighted imaging in the follow-up of treated high-grade gliomas: tumor recurrence versus radiation injury. *AJNR Am J Neuroradiol* 2004, **25**: 201-9
87. Prager AJ, Martinez N, Beal K, Omuro A, Zhang Z, Young RJ. Diffusion and perfusion MRI to differentiate treatment-related changes including pseudoprogression from recurrent tumors in high-grade gliomas with histopathologic evidence. *AJNR Am J Neuroradiol* 2015, **36**: 877-85. doi: 10.3174/ajnr.A4218
88. Sugahara T, Korogi Y, Tomiguchi S, Shigematsu Y, Ikushima I, Kira T, *et al.* Posttherapeutic intraaxial brain tumor: the value of perfusion-sensitive contrast-enhanced MR imaging for differentiating tumor

- recurrence from nonneoplastic contrast-enhancing tissue. *AJNR Am J Neuroradiol* 2000, 21: 901-9.
89. Xu JL, Li YL, Lian JM, Dou SW, Yan FS, Wu H, *et al.* Distinction between postoperative recurrent glioma and radiation injury using MR diffusion tensor imaging. *Neuroradiology* 2010, **52**:1193-9. doi: 10.1007/s00234-010-0731-4
90. Young RJ, Gupta A, Shah AD, Graber JJ, Chan TA, Zhang Z, *et al.* MRI perfusion in determining pseudoprogression in patients with glioblastoma. *Clin Imaging* 2013, 37: 41-9. doi: 10.1016/j.clinimag.2012.02.016
91. Yun TJ, Park CK, Kim TM, Lee SH, Kim JH, Sohn CH, *et al.* Glioblastoma treated with concurrent radiation therapy and temozolomide chemotherapy: differentiation of true progression from pseudoprogression with quantitative dynamic contrast-enhanced MR imaging. *Radiology* 2015, 274: 830-40. doi: 10.1148/radiol.14132632
92. Wesseling P, Ruiter DJ, Burger PC. Angiogenesis in brain tumors; pathobiological and clinical aspects. *J Neurooncol* 1997, 32: 253-65. doi: 10.1023/a:1005746320099
93. Nael K, Bauer AH, Hormigo A, Lemole M, Germano IM, Puig J, *et al.* Multiparametric MRI for differentiation of radiation necrosis from recurrent tumor in patients with treated glioblastoma. *AJR Am J Roentgenol* 2018, 210: 18-23. doi: 10.2214/AJR.17.18003
94. Soni N, Ora M, Mohindra N, Menda Y, Bathla G. Diagnostic performance of PET and perfusion-weighted imaging in differentiating tumor recurrence or progression from radiation necrosis in posttreatment gliomas: a review of literature. *AJNR Am J Neuroradiol* 2020, 41: 1550-7. doi: 10.3174/ajnr.A6685

### Legend for Figure 1

**Fig. 1.** Illustrations of **A.** A pseudoprogression resulting from radiation-induced inflamed necrosis. MR images taken post-operatively show axial T2, FLAIR, T1 contrast-enhanced, and perfusion, with enhancement in the right temporal lobe, areas of T2/FLAIR hyperintensity in the surrounding parenchima, indicating a lesion induced by radiation therapy. There is no increase in relative cerebral blood volume in the enhancing portion of the lesion, which further suggests pseudoprogression. **B.** A tumor relapse shown by images taken from the same patient three months later. MR images taken post-operatively show axial T2, FLAIR, T1 contrast-enhanced, and perfusion, with heterogeneous enhancement in the right temporal lobe and great extension of the T2/FLAIR hyperintensity area, indicating a combination of tumor spread and vasogenic edema involving the right peritrigonal and insular regions. There is increased relative cerebral blood volume in the enhancing portion of the lesion, representing tumoral progression.

